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(54) Title: SECRETED PROTEINS

(57) Abstract: The invention provides human secreted proteins (SECP) and polynucleotides which identify and encode SECP. The invention also provides expression vectors, host cells, antibodies, agonists, and antagonists. The invention also provides methods for diagnosing, treating, or preventing disorders associated with aberrant expression of SECP.

SECRETED PROTEINS

TECHNICAL FIELD

This invention relates to nucleic acid and amino acid sequences of secreted proteins and to the use of these sequences in the diagnosis, treatment, and prevention of cell proliferative, autoimmune/inflammatory, cardiovascular, neurological, and developmental disorders, and in the assessment of the effects of exogenous compounds on the expression of nucleic acid and amino acid sequences of secreted proteins.

BACKGROUND OF THE INVENTION

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Protein transport and secretion are essential for cellular function. Protein transport is mediated by a signal peptide located at the amino terminus of the protein to be transported or secreted. The signal peptide is comprised of about ten to twenty hydrophobic amino acids which target the nascent protein from the ribosome to a particular membrane bound compartment such as the endoplasmic reticulum (ER). Proteins targeted to the ER may either proceed through the secretory pathway or remain in any of the secretory organelles such as the ER, Golgi apparatus, or lysosomes. Proteins that transit through the secretory pathway are either secreted into the extracellular space or retained in the plasma membrane. Proteins that are retained in the plasma membrane contain one or more transmembrane domains, each comprised of about 20 hydrophobic amino acid residues. Secreted proteins are generally synthesized as inactive precursors that are activated by posttranslational processing events during transit through the secretory pathway. Such events include glycosylation, proteolysis, and removal of the signal peptide by a signal peptidase. Other events that may occur during protein transport include chaperone-dependent unfolding and folding of the nascent protein and interaction of the protein with a receptor or pore complex. Examples of secreted proteins with amino terminal signal peptides are discussed below and include proteins with important roles in cell-to-cell signaling. Such proteins include transmembrane receptors and cell surface markers, extracellular matrix molecules, cytokines, hormones, growth and differentiation factors, enzymes, neuropeptides, vasomediators, cell surface markers, and antigen recognition molecules. (Reviewed in Alberts, B. et al. (1994) Molecular Biology of The Cell, Garland Publishing, New York, NY, pp. 557-560, 582-592.)

Cell surface markers include cell surface antigens identified on leukocytic cells of the immune system. These antigens have been identified using systematic, monoclonal antibody (mAb)-based "shot gun" techniques. These techniques have resulted in the production of hundreds of mAbs directed against unknown cell surface leukocytic antigens. These antigens have been grouped into "clusters of differentiation" based on common immunocytochemical localization patterns in various

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differentiated and undifferentiated leukocytic cell types. Antigens in a given cluster are presumed to identify a single cell surface protein and are assigned a "cluster of differentiation" or "CD" designation. Some of the genes encoding proteins identified by CD antigens have been cloned and verified by standard molecular biology techniques. CD antigens have been characterized as both transmembrane proteins and cell surface proteins anchored to the plasma membrane via covalent attachment to fatty acid-containing glycolipids such as glycosylphosphatidylinositol (GPI). (Reviewed in Barclay, A.N. et al. (1995) <u>The Leucocyte Antigen Facts Book</u>, Academic Press, San Diego, CA, pp. 17-20.)

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Matrix proteins (MPs) are transmembrane and extracellular proteins which function in formation, growth, remodeling, and maintenance of tissues and as important mediators and regulators of the inflammatory response. The expression and balance of MPs may be perturbed by biochemical changes that result from congenital, epigenetic, or infectious diseases. In addition, MPs affect leukocyte migration, proliferation, differentiation, and activation in the immune response. MPs are frequently characterized by the presence of one or more domains which may include collagen-like domains, EGF-like domains, immunoglobulin-like domains, and fibronectin-like domains. In addition, MPs may be heavily glycosylated and may contain an Arginine-Glycine-Aspartate (RGD) tripeptide motif which may play a role in adhesive interactions. MPs include extracellular proteins such as fibronectin, collagen, galectin, vitronectin and its proteolytic derivative somatomedin B; and cell adhesion receptors such as cell adhesion molecules (CAMs), cadherins, and integrins. (Reviewed in Ayad, S. et al. (1994) The Extracellular Matrix Facts Book, Academic Press, San Diego, CA, pp. 2-16; Ruoslahti, E. (1997) Kidney Int. 51:1413-1417; Sjaastad, M.D. and Nelson, W.J. (1997) BioEssays 19:47-55.)

Mucins are highly glycosylated glycoproteins that are the major structural component of the mucus gel. The physiological functions of mucins are cytoprotection, mechanical protection, maintenance of viscosity in secretions, and cellular recognition. MUC6 is a human gastric mucin that is also found in gall bladder, pancreas, seminal vesicles, and female reproductive tract (Toribara, N.W. et al. (1997) J. Biol. Chem. 272:16398-16403). The MUC6 gene has been mapped to human chromosome 11 (Toribara, N.W. et al. (1993) J. Biol. Chem. 268:5879-5885). Hemomucin is a novel Drosophila surface mucin that may be involved in the induction of antibacterial effector molecules (Theopold, U. et al. (1996) J. Biol. Chem. 217:12708-12715).

Tuftelins are one of four different enamel matrix proteins that have been identified so far. The other three known enamel matrix proteins are the amelogenins, enamelin and ameloblastin. Assembly of the enamel extracellular matrix from these component proteins is believed to be critical in producing a matrix competent to undergo mineral replacement. (Paine, C.T. et al. (1998) Connect Tissue Res. 38:257-267). Tuftelin mRNA has been found to be expressed in human ameloblastoma

tumor, a non-mineralized odontogenic tumor (Deutsch, D. et al. (1998) Connect. Tissue Res. 39:177-184).

Olfactomedin-related proteins are extracellular matrix, secreted glycoproteins with conserved C-terminal motifs. They are expressed in a wide variety of tissues and in broad range of species, from Caenorhabditis elegans to Homo sapiens. Olfactomedin-related proteins comprise a gene family with at least 5 family members in humans. One of the five, TIGR/myocilin protein, is expressed in the eye and is associated with the pathogenesis of glaucoma (Kulkarni, N.H. et al. (2000) Genet. Res. 76:41-50). Research by Yokoyama et al. (1996) found a 135-amino acid protein, termed AMY, having 96% sequence identity with rat neuronal olfactomedin-releated ER localized protein in a neuroblastoma cell line cDNA library, suggesting an essential role for AMY in nerve tissue (Yokoyama, M. et al. (1996) DNA Res. 3:311-320). Neuron-specific olfactomedin-related glycoproteins isolated from rat brain cDNA libraries show strong sequence similarity with olfactomedin. This similarity is suggestive of a matrix-related function of these glycoproteins in neurons and neurosecretory cells (Danielson, P.E. et al. (1994) J. Neurosci. Res. 38:468-478).

Mac-2 binding protein is a 90-kD serum protein (90K), a secreted glycoprotein isolated from both the human breast carcinoma cell line SK-BR-3, and human breast milk. It specifically binds to a human macrophage-associated lectin, Mac-2. Structurally, the mature protein is 567 amino acids in length and is proceeded by an 18-amino acid leader. There are 16 cysteines and seven potential N-linked glycosylation sites. The first 106 amino acids represent a domain very similar to an ancient protein superfamily defined by a macrophage scavenger receptor cysteine-rich domain (Koths, K. et al. (1993) J. Biol. Chem. 268:14245-14249). 90K is elevated in the serum of subpopulations of AIDS patients and is expressed at varying levels in primary tumor samples and tumor cell lines. Ullrich et al. (1994) have demonstrated that 90K stimulates host defense systems and can induce interleukin-2 secretion. This immune stimulation is proposed to be a result of oncogenic transformation, viral infection or pathogenic invasion (Ullrich, A. et al. (1994) J. Biol. Chem. 269:18401-18407).

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Semaphorins are a large group of axonal guidance molecules consisting of at least 30 different members and are found in vertebrates, invertebrates, and even certain viruses. All semaphorins contain the sema domain which is approximately 500 amino acids in length. Neuropilin, a semaphorin receptor, has been shown to promote neurite outgrowth in vitro. The extracellular region of neuropilins consists of three different domains: CUB, discoidin, and MAM domains. The CUB and the MAM motifs of neuropilin have been suggested to have roles in protein-protein interactions and are thought to be involved in the binding of semaphorins through the sema and the C-terminal domains (reviewed in Raper, J.A. (2000) Curr. Opin. Neurobiol. 10:88-94). Plexins are neuronal cell surface molecules that mediate cell adhesion via a homophilic binding mechanism in the presence of calcium ions. Plexins have been shown to be expressed in the receptors and neurons of

particular sensory systems (Ohta, K. et al. (1995) Cell 14:1189-1199). There is evidence that suggests that some plexins function to control motor and CNS axon guidance in the developing nervous system. Plexins, which themselves contain complete semaphorin domains, may be both the ancestors of classical semaphorins and binding partners for semaphorins (Winberg, M.L. et al (1998) Cell 95:903-916).

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Human pregnancy-specific beta 1-glycoprotein (PSG) is a family of closely related glycoproteins of molecular weights of 72 KDa, 64KDa, 62KDa, and 54KDa. Together with the carcinoembryonic antigen, they comprise a subfamily within the immunoglobulin superfamily (Plouzek, C.A. and Chou, J.Y. (1991) Endocrinology 129:950-958) Different subpopulations of PSG have been found to be produced by the trophoblasts of the human placenta, and the amnionic and chorionic membranes (Plouzek, C.A. et al. (1993) Placenta 14:277-285).

Autocrine motility factor (AMF) is one of the motility cytokines regulating tumor cell migration; therefore identification of the signaling pathway coupled with it has critical importance. Autocrine motility factor receptor (AMFR) expression has been found to be associated with tumor progression in thymoma (Ohta Y. et al. (2000) Int. J. Oncol. 17:259-264). AMFR is a cell surface glycoprotein of molecular weight 78KDa.

Hormones are signaling molecules that coordinately regulate basic physiological processes from embryogenesis throughout adulthood. These processes include metabolism, respiration, reproduction, excretion, fetal tissue differentiation and organogenesis, growth and development, homeostasis, and the stress response. Hormonal secretions and the nervous system are tightly integrated and interdependent. Hormones are secreted by endocrine glands, primarily the hypothalamus and pituitary, the thyroid and parathyroid, the pancreas, the adrenal glands, and the ovaries and testes.

The secretion of hormones into the circulation is tightly controlled. Hormones are often secreted in diurnal, pulsatile, and cyclic patterns. Hormone secretion is regulated by perturbations in blood biochemistry, by other upstream-acting hormones, by neural impulses, and by negative feedback loops. Blood hormone concentrations are constantly monitored and adjusted to maintain optimal, steady-state levels. Once secreted, hormones act only on those target cells that express specific receptors.

Most disorders of the endocrine system are caused by either hyposecretion or hypersecretion of hormones. Hyposecretion often occurs when a hormone's gland of origin is damaged or otherwise impaired. Hypersecretion often results from the proliferation of tumors derived from hormone-secreting cells. Inappropriate hormone levels may also be caused by defects in regulatory feedback loops or in the processing of hormone precursors. Endocrine malfunction may also occur when the target cell fails to respond to the hormone.

Hormones can be classified biochemically as polypeptides, steroids, eicosanoids, or amines. Polypeptide hormones, which include diverse hormones such as insulin and growth hormone, vary in size and function and are often synthesized as inactive precursors that are processed intracellularly into mature, active forms. Amine hormones, which include epinephrine and dopamine, are amino acid derivatives that function in neuroendocrine signaling. Steroid hormones, which include the cholesterol-derived hormones estrogen and testosterone, function in sexual development and reproduction. Eicosanoid hormones, which include prostaglandins and prostacyclins, are fatty acid derivatives that function in a variety of processes. Most polypeptide hormones and some amine hormones are soluble in the circulation where they are highly susceptible to proteolytic degradation within seconds after their secretion. Steroid hormones and eicosanoid hormones are insoluble and must be transported in the circulation by carrier proteins. The following discussion will focus primarily on polypeptide hormones.

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Hormones secreted by the hypothalamus and pituitary gland play a critical role in endocrine function by coordinately regulating hormonal secretions from other endocrine glands in response to neural signals. Hypothalamic hormones include thyrotropin-releasing hormone, gonadotropin-releasing hormone, somatostatin, growth-hormone releasing factor, corticotropin-releasing hormone, substance P, dopamine, and prolactin-releasing hormone. These hormones directly regulate the secretion of hormones from the anterior lobe of the pituitary. Hormones secreted by the anterior pituitary include adrenocorticotropic hormone (ACTH), melanocyte-stimulating hormone, somatotropic hormones such as growth hormone and prolactin, glycoprotein hormones such as thyroid-stimulating hormone, luteinizing hormone (LH), and follicle-stimulating hormone (FSH), β -lipotropin, and β -endorphins. These hormones regulate hormonal secretions from the thyroid, pancreas, and adrenal glands, and act directly on the reproductive organs to stimulate ovulation and spermatogenesis. The posterior pituitary synthesizes and secretes antidiuretic hormone (ADH, vasopressin) and oxytocin.

Disorders of the hypothalamus and pituitary often result from lesions such as primary brain tumors, adenomas, infarction associated with pregnancy, hypophysectomy, aneurysms, vascular malformations, thrombosis, infections, immunological disorders, and complications due to head trauma. Such disorders have profound effects on the function of other endocrine glands. Disorders associated with hypopituitarism include hypogonadism, Sheehan syndrome, diabetes insipidus, Kallman's disease, Hand-Schuller-Christian disease, Letterer-Siwe disease, sarcoidosis, empty sella syndrome, and dwarfism. Disorders associated with hyperpituitarism include acromegaly, giantism, and syndrome of inappropriate ADH secretion (SIADH), often caused by benign adenomas.

Hormones secreted by the thyroid and parathyroid primarily control metabolic rates and the regulation of serum calcium levels, respectively. Thyroid hormones include calcitonin, somatostatin,

and thyroid hormone. The parathyroid secretes parathyroid hormone. Disorders associated with hypothyroidism include goiter, myxedema, acute thyroiditis associated with bacterial infection, subacute thyroiditis associated with viral infection, autoimmune thyroiditis (Hashimoto's disease), and cretinism. Disorders associated with hyperthyroidism include thyrotoxicosis and its various forms, Grave's disease, pretibial myxedema, toxic multinodular goiter, thyroid carcinoma, and Plummer's disease. Disorders associated with hyperparathyroidism include Conn disease (chronic hypercalemia) leading to bone resorption and parathyroid hyperplasia.

Hormones secreted by the pancreas regulate blood glucose levels by modulating the rates of carbohydrate, fat, and protein metabolism. Pancreatic hormones include insulin, glucagon, amylin, γ-aminobutyric acid, gastrin, somatostatin, and pancreatic polypeptide. The principal disorder associated with pancreatic dysfunction is diabetes mellitus caused by insufficient insulin activity. Diabetes mellitus is generally classified as either Type I (insulin-dependent, juvenile diabetes) or Type II (non-insulin-dependent, adult diabetes). The treatment of both forms by insulin replacement therapy is well known. Diabetes mellitus often leads to acute complications such as hypoglycemia (insulin shock), coma, diabetic ketoacidosis, lactic acidosis, and chronic complications leading to disorders of the eye, kidney, skin, bone, joint, cardiovascular system, nervous system, and to decreased resistance to infection.

The anatomy, physiology, and diseases related to hormonal function are reviewed in McCance, K. L. and Huether, S. E. (1994) <u>Pathophysiology: The Biological Basis for Disease in Adults and Children</u>, Mosby-Year Book, Inc., St. Louis, MO; Greenspan, F. S. and Baxter, J. D. (1994) <u>Basic and Clinical Endocrinology</u>, Appleton and Lange, East Norwalk, CT.

Growth factors are secreted proteins that mediate intercellular communication. Unlike hormones, which travel great distances via the circulatory system, most growth factors are primarily local mediators that act on neighboring cells. Most growth factors contain a hydrophobic N-terminal signal peptide sequence which directs the growth factor into the secretory pathway. Most growth factors also undergo post-translational modifications within the secretory pathway. These modifications can include proteolysis, glycosylation, phosphorylation, and intramolecular disulfide bond formation. Once secreted, growth factors bind to specific receptors on the surfaces of neighboring target cells, and the bound receptors trigger intracellular signal transduction pathways. These signal transduction pathways elicit specific cellular responses in the target cells. These responses can include the modulation of gene expression and the stimulation or inhibition of cell division, cell differentiation, and cell motility.

Growth factors fall into at least two broad and overlapping classes. The broadest class includes the large polypeptide growth factors, which are wide-ranging in their effects. These factors include epidermal growth factor (EGF), fibroblast growth factor (FGF), transforming growth factor- β

 $(TGF-\beta)$, insulin-like growth factor (IGF), nerve growth factor (NGF), and platelet-derived growth factor (PDGF), each defining a family of numerous related factors. The large polypeptide growth factors, with the exception of NGF, act as mitogens on diverse cell types to stimulate wound healing, bone synthesis and remodeling, extracellular matrix synthesis, and proliferation of epithelial, epidermal, and connective tissues. Members of the TGF-B, EGF, and FGF families also function as inductive signals in the differentiation of embryonic tissue. NGF functions specifically as a neurotrophic factor, promoting neuronal growth and differentiation.

Another class of growth factors includes the hematopoietic growth factors, which are narrow in their target specificity. These factors stimulate the proliferation and differentiation of blood cells such as B-lymphocytes, T-lymphocytes, erythrocytes, platelets, eosinophils, basophils, neutrophils, macrophages, and their stem cell precursors. These factors include the colony-stimulating factors (G-CSF, M-CSF, GM-CSF, and CSF1-3), erythropoietin, and the cytokines. The cytokines are specialized hematopoietic factors secreted by cells of the immune system and are discussed in detail below.

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Hormones travel through the circulation and bind to specific receptors on the surface of, or within, target cells. Although they have diverse biochemical compositions and mechanisms of action, hormones can be grouped into two categories. One category includes small lipophilic hormones that diffuse through the plasma membrane of target cells, bind to cytosolic or nuclear receptors, and form a complex that alters gene expression. Examples of these molecules include retinoic acid, thyroxine, and the cholesterol-derived steroid hormones such as progesterone, estrogen, testosterone, cortisol, and aldosterone. The second category includes hydrophilic hormones that function by binding to cell surface receptors that transduce signals across the plasma membrane. Examples of such hormones include amino acid derivatives such as catecholamines (epinephrine, norepinephrine) and histamine, and peptide hormones such as glucagon, insulin, gastrin, secretin, cholecystokinin, adrenocorticotropic hormone, follicle stimulating hormone, luteinizing hormone, thyroid stimulating hormone, and vasopressin. (See, for example, Lodish et al. (1995) Molecular Cell Biology, Scientific American Books Inc., New York, NY, pp. 856-864.)

Pro-opiomelanocortin (POMC) is the precursor polypeptide of corticotropin (ACTH), a hormone synthesized by the anterior pituitary gland, which functions in the stimulation of the adrenal 30 cortex. POMC is also the precursor polypeptide of the hormone beta-lipotropin (beta-LPH). Each hormone includes smaller peptides with distinct biological activities: alpha-melanotropin (alpha-MSH) and corticotropin-like intermediate lobe peptide (CLIP) are formed from ACTH; gammalipotropin (gamma-LPH) and beta-endorphin are peptide components of beta-LPH; while beta-MSH is contained within gamma-LPH. Adrenal insufficiency due to ACTH deficiency, resulting from a genetic mutation in exons 2 and 3 of POMC results in an endocrine disorder characterized by early-

onset obesity, adrenal insufficiency, and red hair pigmentation (Chretien, M. et al. (1979) Can. J. Biochem. 57:1111-1121; Krude, H. et al. (1998) Nat. Genet. 19:155-157; Online Mendelian Inheritance in Man (OMIM) 176830).

Growth and differentiation factors are secreted proteins which function in intercellular communication. Some factors require oligomerization or association with membrane proteins for activity. Complex interactions among these factors and their receptors trigger intracellular signal transduction pathways that stimulate or inhibit cell division, cell differentiation, cell signaling, and cell motility. Most growth and differentiation factors act on cells in their local environment (paracrine signaling). There are three broad classes of growth and differentiation factors. The first class includes the large polypeptide growth factors such as epidermal growth factor (EGF), fibroblast growth factor, transforming growth factor, insulin-like growth factor (IGF), and platelet-derived growth factor. EGF includes a 30-40 residue EGF repeat domain, composed of conserved cysteine and glycine residues, found in a variety of proteins involved in cell proliferation, including the leukocyte antigen CD97 and the Notch family proteins (Greener, M. (2000) Mol. Med. Today 6:139-140). IGF forms a heterotrimeric complex with IGF-binding-protein 3 and the acid-labile subunit (ALS). ALS is largely composed of 18-20 leucine-rich repeats of 24 amino acids (Leong, S.R. et al. (1992) Mol. Endocrinol. 6:870-876). The second class includes the hematopoietic growth factors such as the colony stimulating factors (CSFs). Hematopoietic growth factors stimulate the proliferation and differentiation of blood cells such as B-lymphocytes, T-lymphocytes, erythrocytes, platelets, eosinophils, basophils, neutrophils, macrophages, and their stem cell precursors. The third class includes small peptide factors such as bombesin, vasopressin, oxytocin, endothelin, transferrin, angiotensin II, vasoactive intestinal peptide, and bradykinin, which function as hormones to regulate cellular functions other than proliferation.

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Growth and differentiation factors play critical roles in neoplastic transformation of cells in vitro and in tumor progression in vivo. Inappropriate expression of growth factors by tumor cells may contribute to vascularization and metastasis of tumors. During hematopoiesis, growth factor misregulation can result in anemias, leukemias, and lymphomas. Certain growth factors such as interferon are cytotoxic to tumor cells both in vivo and in vitro. Moreover, some growth factors and growth factor receptors are related both structurally and functionally to oncoproteins. In addition, growth factors affect transcriptional regulation of both proto-oncogenes and oncosuppressor genes. (Reviewed in Pimentel, E. (1994) Handbook of Growth Factors, CRC Press, Ann Arbor, MI, pp. 1-9.)

In addition, some of the large polypeptide growth factors play crucial roles in the induction of the primordial germ layers in the developing embryo. This induction ultimately results in the formation of the embryonic mesoderm, ectoderm, and endoderm which in turn provide the framework for the entire adult body plan. Disruption of this inductive process would be catastrophic

to embryonic development. One such growth factor, wnt, is a secreted glycoprotein that has activity as both a short-range inducer and as a long-range morphogen (for a review, see Howes, R. and S. Bray (2000) Current Biology 10:R222-R226). Wnt signaling is implicated in diseases including cancer and Alzheimer's Disease (Bienz, M. and H. Clevers (2000) Cell 103:311-320; Polakis, P. (2000) Genes Dev. 14:1837-1851; De Ferrari, G.V. and N.C. Inestrosa (2000) Brain Res. Brain. Res. Rev. 33:1-12). Chordin is a developmental protein that binds to ventralizing TGF-beta-like bone morphogenetic proteins (BMPs) and sequesters them in latent complexes, causing dorsalization of tissue (Pappano, W. N. et al. (1998) Genomics 52:236-239). Other developmental proteins that regulate BMPs include noggin, cerberus, dan, and gremlin (Schmitt, J.M. et al. (1999) J. Orthop. Res. 17:269-278).

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The Slit protein, first identified in Drosophila, is critical in central nervous system midline formation and potentially in nervous tissue histogenesis and axonal pathfinding. Itoh et al. ((1998) Brain Res. Mol. Brain Res. 62:175-186) have identified mammalian homologues of the slit gene (human Slit-1, Slit-2, Slit-3 and rat Slit-1). The encoded proteins are putative secreted proteins containing EGF-like motifs and leucine-rich repeats, both of which are conserved protein-protein interaction domains. Slit-1, -2, and -3 mRNAs are expressed in the brain, spinal cord, and thyroid, respectively (Itoh, A. et al., supra). The Slit family of proteins are indicated to be functional ligands of glypican-1 in nervous tissue and it is suggested that their interactions may be critical in certain stages during central nervous system histogenesis (Liang, Y. et al. (1999) J. Biol. Chem. 274:17885-17892).

Neuropeptides and vasomediators (NP/VM) comprise a large family of endogenous signaling molecules. Included in this family are neuropeptides and neuropeptide hormones such as bombesin, neuropeptide Y, neurotensin, neuromedin N, melanocortins, opioids, galanin, somatostatin, tachykinins, urotensin II and related peptides involved in smooth muscle stimulation, vasopressin, vasoactive intestinal peptide, and circulatory system-borne signaling molecules such as angiotensin, complement, calcitonin, endothelins, formyl-methionyl peptides, glucagon, cholecystokinin and gastrin. NP/VMs can transduce signals directly, modulate the activity or release of other neurotransmitters and hormones, and act as catalytic enzymes in cascades. The effects of NP/VMs range from extremely brief to long-lasting. (Reviewed in Martin, C.R. et al. (1985) Endocrine Physiology, Oxford University Press, New York, NY, pp. 57-62.)

NP/VMs are involved in numerous neurological and cardiovascular disorders. For example, neuropeptide Y is involved in hypertension, congestive heart failure, affective disorders, and appetite regulation. Somatostatin inhibits secretion of growth hormone and prolactin in the anterior pituitary, as well as inhibiting secretion in intestine, pancreatic acinar cells, and pancreatic beta-cells. A reduction in somatostatin levels has been reported in Alzheimer's disease and Parkinson's disease.

Vasopressin acts in the kidney to increase water and sodium absorption, and in higher concentrations stimulates contraction of vascular smooth muscle, platelet activation, and glycogen breakdown in the liver. Vasopressin and its analogues are used clinically to treat diabetes insipidus. Endothelin and angiotensin are involved in hypertension, and drugs, such as captopril, which reduce plasma levels of angiotensin, are used to reduce blood pressure (Watson, S. and S. Arkinstall (1994) The G-protein Linked Receptor Facts Book, Academic Press, San Diego CA, pp. 194; 252; 284; 55; 111).

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Neuropeptides have also been shown to have roles in nociception (pain). Vasoactive intestinal peptide appears to play an important role in chronic neuropathic pain. Nociceptin, an endogenous ligand for for the opioid receptor-like 1 receptor, is thought to have a predominantly antinociceptive effect, and has been shown to have analgesic properties in different animal models of tonic or chronic pain (Dickinson, T. and Fleetwood-Walker, S.M. (1998) Trends Pharmacol. Sci. 19:346-348).

Cytokines comprise a family of signaling molecules that modulate the immune system and the inflammatory response. Cytokines are usually secreted by leukocytes, or white blood cells, in response to injury or infection. Cytokines function as growth and differentiation factors that act primarily on cells of the immune system such as B- and T-lymphocytes, monocytes, macrophages, and granulocytes. Like other signaling molecules, cytokines bind to specific plasma membrane receptors and trigger intracellular signal transduction pathways which alter gene expression patterns. There is considerable potential for the use of cytokines in the treatment of inflammation and immune system disorders.

Cytokine structure and function have been extensively characterized <u>in vitro</u>. Most cytokines are small polypeptides of about 30 kilodaltons or less. Over 50 cytokines have been identified from human and rodent sources. Examples of cytokine subfamilies include the interferons (IFN- α , - β , and - γ), the interleukins (IL1-IL13), the tumor necrosis factors (TNF- α and - β), and the chemokines. Many cytokines have been produced using recombinant DNA techniques, and the activities of individual cytokines have been determined <u>in vitro</u>. These activities include regulation of leukocyte proliferation, differentiation, and motility.

The activity of an individual cytokine <u>in vitro</u> may not reflect the full scope of that cytokine's activity <u>in vivo</u>. Cytokines are not expressed individually <u>in vivo</u> but are instead expressed in combination with a multitude of other cytokines when the organism is challenged with a stimulus. Together, these cytokines collectively modulate the immune response in a manner appropriate for that particular stimulus. Therefore, the physiological activity of a cytokine is determined by the stimulus itself and by complex interactive networks among co-expressed cytokines which may demonstrate both synergistic and antagonistic relationships.

Chemokines comprise a cytokine subfamily with over 30 members. (Reviewed in Wells, T.

N. C. and Peitsch, M. C. (1997) J. Leukoc. Biol. 61:545-550.) Chemokines were initially identified as chemotactic proteins that recruit monocytes and macrophages to sites of inflammation. Recent evidence indicates that chemokines may also play key roles in hematopoiesis and HIV-1 infection. Chemokines are small proteins which range from about 6-15 kilodaltons in molecular weight. Chemokines are further classified as C, CC, CXC, or CX₃C based on the number and position of critical cysteine residues. The CC chemokines, for example, each contain a conserved motif consisting of two consecutive cysteines followed by two additional cysteines which occur downstream at 24- and 16-residue intervals, respectively (ExPASy PROSITE database, documents PS00472 and PDOC00434). The presence and spacing of these four cysteine residues are highly conserved, whereas the intervening residues diverge significantly. However, a conserved tyrosine located about 15 residues downstream of the cysteine doublet seems to be important for chemotactic activity. Most of the human genes encoding CC chemokines are clustered on chromosome 17, although there are a few examples of CC chemokine genes that map elsewhere. Other chemokines include lymphotactin (C chemokine); macrophage chemotactic and activating factor (MCAF/MCP-1; CC chemokine); platelet factor 4 and IL-8 (CXC chemokines); and fractalkine and neurotractin 15 (CX₃C chemokines). (Reviewed in Luster, A. D. (1998) N. Engl. J. Med. 338:436-445.)

Other proteins that contain signal peptides include secreted proteins with enzymatic activity. Such activity includes, for example, oxidoreductase/dehydrogenase activity, transferase activity, hydrolase activity, lyase activity, isomerase activity, or ligase activity. For example, matrix metalloproteinases are secreted hydrolytic enzymes that degrade the extracellular matrix and thus play an important role in tumor metastasis, tissue morphogenesis, and arthritis (Reponen, P. et al. (1995) Dev. Dyn. 202:388-396; Firestein, G.S. (1992) Curr. Opin. Rheumatol. 4:348-354; Ray, J.M. and Stetler-Stevenson, W.G. (1994) Eur. Respir. J. 7:2062-2072; and Mignatti, P. and Rifkin, D.B. (1993) Physiol. Rev. 73:161-195). The catalytic protein disulfide isomerase (PDI) is found in membrane-bound eukaryotic compartments such as the endoplasmic reticulum (ER). It facilitates disulfide bond exchange as well as correct glycosylation. Edman et al. (1995; Nature 317:267-70) reported that rat PDI is useful for the in vitro production and folding of recombinant human proteins. Likewise, purified PDI is also commercially useful for the production and folding of recombinant, therapeutic human proteins such as tissue plasminogen activator (tPA). Ceruloplasmin is a serum multicopper oxidase which plays a role in iron metabolism. Aceruloplasminemia is characterized by diabetes, retinal degeneration, and neurologic symptoms (for a review, see Gitlin, J.D. (1998) Pediatr. Res. 4:271-276). Additional examples are the acetyl-CoA synthetases which activate acetate for use in lipid synthesis or energy generation (Luong, A. et al. (2000) J. Biol. Chem. 275:26458-26466). The result of acetyl-CoA synthetase activity is the formation of acetyl-CoA from acetate and CoA. Acetyl-CoA sythetases share a region of sequence similarity identified as the AMP-binding domain

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signature. Acetyl-CoA synthetase has been shown to be associated with hypertension (Toh, H. (1991) Protein Seq. Data Anal. 4:111-117; and Iwai, N. et al. (1994) Hypertension 23:375-380).

A number of isomerases catalyze steps in protein folding, phototransduction, and various anabolic and catabolic pathways. One class of isomerases is known as peptidyl-prolyl *cis-trans* isomerases (PPIases). PPIases catalyze the *cis* to *trans* isomerization of certain proline imidic bonds in proteins. Two families of PPIases are the FK506 binding proteins (FKBPs), and cyclophilins (CyPs). FKBPs bind the potent immunosuppressants FK506 and rapamycin, thereby inhibiting signaling pathways in T-cells. Specifically, the PPIase activity of FKBPs is inhibited by binding of FK506 or rapamycin. There are five members of the FKBP family which are named according to their calculated molecular masses (FKBP12, FKBP13, FKBP25, FKBP52, and FKBP65), and localized to different regions of the cell where they associate with different protein complexes (Coss, M. et al. (1995) J. Biol. Chem. 270:29336-29341; Schreiber, S.L. (1991) Science 251:283-287).

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The peptidyl-prolyl isomerase activity of CyP may be part of the signaling pathway that leads to T-cell activation. CyP isomerase activity is associated with protein folding and protein trafficking, and may also be involved in assembly/disassembly of protein complexes and regulation of protein activity. For example, in *Drosophila*, the CyP NinaA is required for correct localization of rhodopsins, while a mammalian CyP (Cyp40) is part of the Hsp90/Hsc70 complex that binds steroid receptors. The mammalian CypA has been shown to bind the *gag* protein from human immunodeficiency virus 1 (HIV-1), an interaction that can be inhibited by cyclosporin. Since cyclosporin has potent anti-HIV-1 activity, CypA may play an essential function in HIV-1 replication. Finally, Cyp40 has been shown to bind and inactivate the transcription factor c-Myb, an effect that is reversed by cyclosporin. This effect implicates CyPs in the regulation of transcription, transformation, and differentiation (Bergsma, D.J. et al (1991) J. Biol. Chem. 266:23204-23214; Hunter, T. (1998) Cell 92:141-143; and Leverson, J.D. and Ness, S.A. (1998) Mol. Cell. 1:203-211).

Gamma-carboxyglutamic acid (Gla) proteins rich in proline (PRGPs) are members of a family of vitamin K-dependent single-pass integral membrane proteins. These proteins are characterized by an extracellular amino terminal domain of approximately 45 amino acids rich in Gla. The intracellular carboxyl terminal region contains one or two copies of the sequence PPXY, a motif present in a variety of proteins involved in such diverse cellular functions as signal transduction, cell cycle progression, and protein turnover (Kulman, J.D. et al. (2001) Proc. Natl. Acad. Sci. USA 98:1370-1375). The process of post-translational modification of glutamic residues to form Gla is Vitamin K-dependent carboxylation. Proteins which contain Gla include plasma proteins involved in blood coagulation. These proteins are prothrombin, proteins C, S, and Z, and coagulation factors VII, IX, and X. Osteocalcin (bone-Gla protein, BGP) and matrix Gla-protein (MGP) also contain Gla (Friedman, P.A. and C.T. Przysiecki (1987) Int. J. Biochem. 19:1-7; C. Vermeer (1990) Biochem. J.

266:625-636).

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Immunoglobulins

Antigen recognition molecules are key players in the sophisticated and complex immune systems which all vertebrates have developed to provide protection from viral, bacterial, fungal, and parasitic infections. A key feature of the immune system is its ability to distinguish foreign molecules, or antigens, from "self" molecules. This ability is mediated primarily by secreted and transmembrane proteins expressed by leukocytes (white blood cells) such as lymphocytes, granulocytes, and monocytes. Most of these proteins belong to the immunoglobulin (Ig) superfamily, members of which contain one or more repeats of a conserved structural domain. This Ig domain is comprised of antiparallel β sheets joined by a disulfide bond in an arrangement called the Ig fold. The criteria for a protein to be a member of the Ig superfamily is to have one or more Ig domains, which are regions of 70-110 amino acid residues in length homologous to either Ig variable-like (V) or Ig constant-like (C) domains. Members of the Ig superfamily include antibodies (Ab), T cell receptors (TCRs), class I and II major histocompatibility (MHC) proteins and immune cell-specific surface markers such as the "cluster of differentiation" or CD antigens, CD2, CD3, CD4, CD8, poly-Ig receptors, Fc receptors, neural cell-adhesion molecule (NCAM) and platelet-derived growth factor receptor (PDGFR).

Ig domains (V and C) are regions of conserved amino acid residues that give a polypeptide a globular tertiary structure called an immunoglobulin (or antibody) fold, which consists of two approximately parallel layers of β -sheets. Conserved cysteine residues form an intrachain disulfide-bonded loop, 55-75 amino acid residues in length, which connects the two layers of β -sheets. Each β -sheet has three or four anti-parallel β -strands of 5-10 amino acid residues. Hydrophobic and hydrophilic interactions of amino acid residues within the β -strands stabilize the Ig fold (hydrophobic on inward facing amino acid residues and hydrophilic on the amino acid residues in the outward facing portion of the strands). A V domain consists of a longer polypeptide than a C domain, with an additional pair of β -strands in the Ig fold.

A consistent feature of Ig superfamily genes is that each sequence of an Ig domain is encoded by a single exon. It is possible that the superfamily evolved from a gene coding for a single Ig domain involved in mediating cell-cell interactions. New members of the superfamily then arose by exon and gene duplications. Modern Ig superfamily proteins contain different numbers of V and/or C domains. Another evolutionary feature of this superfamily is the ability to undergo DNA rearrangements, a unique feature retained by the antigen receptor members of the family.

Many members of the Ig superfamily are integral plasma membrane proteins with extracellular Ig domains. The hydrophobic amino acid residues of their transmembrane domains and

their cytoplasmic tails are very diverse, with little or no homology among Ig family members or to known signal-transducing structures. There are exceptions to this general superfamily description. For example, the cytoplasmic tail of PDGFR has tyrosine kinase activity. In addition Thy-1 is a glycoprotein found on thymocytes and T cells. This protein has no cytoplasmic tail, but is instead attached to the plasma membrane by a covalent glycophosphatidylinositol linkage.

Another common feature of many Ig superfamily proteins is the interactions between Ig domains which are essential for the function of these molecules. Interactions between Ig domains of a multimeric protein can be either homophilic or heterophilic (i.e., between the same or different Ig domains). Antibodies are multimeric proteins which have both homophilic and heterophilic interactions between Ig domains. Pairing of constant regions of heavy chains forms the Fc region of an antibody and pairing of variable regions of light and heavy chains form the antigen binding site of an antibody. Heterophilic interactions also occur between Ig domains of different molecules. These interactions provide adhesion between cells for significant cell-cell interactions in the immune system and in the developing and mature nervous system. (Reviewed in Abbas, A.K. et al. (1991) Cellular and Molecular Immunology, W.B. Saunders Company, Philadelphia, PA, pp.142-145.)

MHC proteins are cell surface markers that bind to and present foreign antigens to T cells. MHC molecules are classified as either class I or class II. Class I MHC molecules (MHC I) are expressed on the surface of almost all cells and are involved in the presentation of antigen to cytotoxic T cells. For example, a cell infected with virus will degrade intracellular viral proteins and express the protein fragments bound to MHC I molecules on the cell surface. The MHC I/antigen complex is recognized by cytotoxic T-cells which destroy the infected cell and the virus within. Class II MHC molecules are expressed primarily on specialized antigen-presenting cells of the immune system, such as B-cells and macrophages. These cells ingest foreign proteins from the extracellular fluid and express MHC II/antigen complex on the cell surface. This complex activates helper T-cells, which then secrete cytokines and other factors that stimulate the immune response. MHC molecules also play an important role in organ rejection following transplantation. Rejection occurs when the recipient's T-cells respond to foreign MHC molecules on the transplanted organ in the same way as to self MHC molecules bound to foreign antigen. (Reviewed in Alberts, B. et al. (1994) Molecular Biology of the Cell, Garland Publishing, New York, NY, pp. 1229-1246.)

Antibodies are multimeric members of the Ig superfamily which are either expressed on the surface of B-cells or secreted by B-cells into the circulation. Antibodies bind and neutralize foreign antigens in the blood and other extracellular fluids. The prototypical antibody is a tetramer consisting of two identical heavy polypeptide chains (H-chains) and two identical light polypeptide chains (L-chains) interlinked by disulfide bonds. This arrangement confers the characteristic Y-shape to

antibody molecules. Antibodies are classified based on their H-chain composition. The five antibody classes, IgA, IgD, IgE, IgG and IgM, are defined by the α , δ , ϵ , γ , and μ H-chain types. There are two types of L-chains, κ and λ , either of which may associate as a pair with any H-chain pair. IgG, the most common class of antibody found in the circulation, is tetrameric, while the other classes of antibodies are generally variants or multimers of this basic structure.

H-chains and L-chains each contain an N-terminal variable region and a C-terminal constant region. The constant region consists of about 110 amino acids in L-chains and about 330 or 440 amino acids in H-chains. The amino acid sequence of the constant region is nearly identical among H- or L-chains of a particular class. The variable region consists of about 110 amino acids in both H- and L-chains. However, the amino acid sequence of the variable region differs among H- or L-chains of a particular class. Within each H- or L-chain variable region are three hypervariable regions of extensive sequence diversity, each consisting of about 5 to 10 amino acids. In the antibody molecule, the H- and L-chain hypervariable regions come together to form the antigen recognition site. (Reviewed in Alberts, B. et al. supra, pp. 1206-1213 and 1216-1217.)

Both H-chains and L-chains contain the repeated Ig domains of members of the Ig superfamily. For example, a typical H-chain contains four Ig domains, three of which occur within the constant region and one of which occurs within the variable region and contributes to the formation of the antigen recognition site. Likewise, a typical L-chain contains two Ig domains, one of which occurs within the constant region and one of which occurs within the variable region.

The immune system is capable of recognizing and responding to any foreign molecule that enters the body. Therefore, the immune system must be armed with a full repertoire of antibodies against all potential antigens. Such antibody diversity is generated by somatic rearrangement of gene segments encoding variable and constant regions. These gene segments are joined together by site-specific recombination which occurs between highly conserved DNA sequences that flank each gene segment. Because there are hundreds of different gene segments, millions of unique genes can be generated combinatorially. In addition, imprecise joining of these segments and an unusually high rate of somatic mutation within these segments further contribute to the generation of a diverse antibody population.

The discovery of new secreted proteins, and the polynucleotides encoding them, satisfies a need in the art by providing new compositions which are useful in the diagnosis, prevention, and treatment of cell proliferative, autoimmune/inflammatory, cardiovascular, neurological, and developmental disorders, and in the assessment of the effects of exogenous compounds on the expression of nucleic acid and amino acid sequences of secreted proteins.

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The invention features purified polypeptides, secreted proteins, referred to collectively as "SECP" and individually as "SECP-1," "SECP-2," "SECP-3," "SECP-4," "SECP-5," "SECP-6." "SECP-7," "SECP-8," "SECP-9," "SECP-10," "SECP-11," "SECP-12," "SECP-13," "SECP-14," "SECP-15," "SECP-16," "SECP-17," "SECP-18," "SECP-19," "SECP-20," "SECP-21," "SECP-22." "SECP-23," "SECP-24," "SECP-25," "SECP-26," "SECP-27," "SECP-28," "SECP-29," "SECP-30," "SECP-31," "SECP-32," "SECP-33," "SECP-34," "SECP-35," "SECP-36," "SECP-37," "SECP-38." "SECP-39," "SECP-40," "SECP-41," "SECP-42," "SECP-43," "SECP-44," "SECP-45." "SECP-46." "SECP-47," "SECP-48," "SECP-49," "SECP-50," "SECP-51," "SECP-52," "SECP-53," and "SECP-54." In one aspect, the invention provides an isolated polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEO ID NO:1-54, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-54. In one alternative, the invention provides an isolated polypeptide comprising the amino acid sequence of SEO ID NO:1-54.

The invention further provides an isolated polynucleotide encoding a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-54. In one alternative, the polynucleotide encodes a polypeptide selected from the group consisting of SEQ ID NO:1-54. In another alternative, the polynucleotide is selected from the group consisting of SEQ ID NO:55-108.

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Additionally, the invention provides a recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide encoding a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-54. In one alternative, the invention provides a cell transformed with the recombinant polynucleotide. In another alternative, the

invention provides a transgenic organism comprising the recombinant polynucleotide.

The invention also provides a method for producing a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-54. The method comprises a) culturing a cell under conditions suitable for expression of the polypeptide, wherein said cell is transformed with a recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide encoding the polypeptide, and b) recovering the polypeptide so expressed.

Additionally, the invention provides an isolated antibody which specifically binds to a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-54.

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The invention further provides an isolated polynucleotide selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:55-108, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:55-108, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). In one alternative, the polynucleotide comprises at least 60 contiguous nucleotides.

Additionally, the invention provides a method for detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:55-108, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:55-108, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). The method comprises a) hybridizing the sample with a probe comprising at least 20 contiguous nucleotides comprising a sequence complementary to said target polynucleotide in the sample, and

which probe specifically hybridizes to said target polynucleotide, under conditions whereby a hybridization complex is formed between said probe and said target polynucleotide or fragments thereof, and b) detecting the presence or absence of said hybridization complex, and optionally, if present, the amount thereof. In one alternative, the probe comprises at least 60 contiguous nucleotides.

The invention further provides a method for detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:55-108, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:55-108, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). The method comprises a) amplifying said target polynucleotide or fragment thereof using polymerase chain reaction amplification, and b) detecting the presence or absence of said amplified target polynucleotide or fragment thereof, and, optionally, if present, the amount thereof.

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The invention further provides a composition comprising an effective amount of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, and a pharmaceutically acceptable excipient. In one embodiment, the composition comprises an amino acid sequence selected from the group consisting of SEQ ID NO:1-54. The invention additionally provides a method of treating a disease or condition associated with decreased expression of functional SECP, comprising administering to a patient in need of such treatment the composition.

The invention also provides a method for screening a compound for effectiveness as an agonist of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-54. The method comprises a) exposing a sample comprising the

polypeptide to a compound, and b) detecting agonist activity in the sample. In one alternative, the invention provides a composition comprising an agonist compound identified by the method and a pharmaceutically acceptable excipient. In another alternative, the invention provides a method of treating a disease or condition associated with decreased expression of functional SECP, comprising administering to a patient in need of such treatment the composition.

Additionally, the invention provides a method for screening a compound for effectiveness as an antagonist of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-54. The method comprises a) exposing a sample comprising the polypeptide to a compound, and b) detecting antagonist activity in the sample. In one alternative, the invention provides a composition comprising an antagonist compound identified by the method and a pharmaceutically acceptable excipient. In another alternative, the invention provides a method of treating a disease or condition associated with overexpression of functional SECP, comprising administering to a patient in need of such treatment the composition.

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The invention further provides a method of screening for a compound that specifically binds to a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-54. The method comprises a) combining the polypeptide with at least one test compound under suitable conditions, and b) detecting binding of the polypeptide to the test compound, thereby identifying a compound that specifically binds to the polypeptide.

The invention further provides a method of screening for a compound that modulates the activity of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group

consisting of SEQ ID NO:1-54. The method comprises a) combining the polypeptide with at least one test compound under conditions permissive for the activity of the polypeptide, b) assessing the activity of the polypeptide in the presence of the test compound, and c) comparing the activity of the polypeptide in the presence of the test compound with the activity of the polypeptide in the absence of the test compound, wherein a change in the activity of the polypeptide in the presence of the test compound is indicative of a compound that modulates the activity of the polypeptide.

The invention further provides a method for screening a compound for effectiveness in altering expression of a target polynucleotide, wherein said target polynucleotide comprises a polynucleotide sequence selected from the group consisting of SEQ ID NO:55-108, the method comprising a) exposing a sample comprising the target polynucleotide to a compound, b) detecting altered expression of the target polynucleotide, and c) comparing the expression of the target polynucleotide in the presence of varying amounts of the compound and in the absence of the compound.

The invention further provides a method for assessing toxicity of a test compound, said method comprising a) treating a biological sample containing nucleic acids with the test compound; b) hybridizing the nucleic acids of the treated biological sample with a probe comprising at least 20 contiguous nucleotides of a polynucleotide selected from the group consisting of i) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:55-108, ii) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:55-108, iii) a polynucleotide having a sequence complementary to i), iv) a polynucleotide complementary to the polynucleotide of ii), and v) an RNA equivalent of i)-iv). Hybridization occurs under conditions whereby a specific hybridization complex is formed between said probe and a target polynucleotide in the biological sample, said target polynucleotide selected from the group consisting of i) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEO ID NO:55-108, ii) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:55-108, iii) a polynucleotide complementary to the polynucleotide of i), iv) a polynucleotide complementary to the polynucleotide of ii), and v) an RNA equivalent of i)-iv). Alternatively, the target polynucleotide comprises a fragment of a polynucleotide sequence selected from the group consisting of i)-v) above; c) quantifying the amount of hybridization complex; and d) comparing the amount of hybridization complex in the treated biological sample with the amount of hybridization complex in an untreated biological sample, wherein a difference in the amount of hybridization complex in the treated biological sample is indicative of toxicity of the test compound.

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BRIEF DESCRIPTION OF THE TABLES

Table 1 summarizes the nomenclature for the full length polynucleotide and polypeptide sequences of the present invention.

Table 2 shows the GenBank identification number and annotation of the nearest GenBank homolog for polypeptides of the invention. The probability scores for the matches between each polypeptide and its homolog(s) are also shown.

Table 3 shows structural features of polypeptide sequences of the invention, including predicted motifs and domains, along with the methods, algorithms, and searchable databases used for analysis of the polypeptides.

Table 4 lists the cDNA and/or genomic DNA fragments which were used to assemble polynucleotide sequences of the invention, along with selected fragments of the polynucleotide sequences.

Table 5 shows the representative cDNA library for polynucleotides of the invention.

Table 6 provides an appendix which describes the tissues and vectors used for construction of the cDNA libraries shown in Table 5.

Table 7 shows the tools, programs, and algorithms used to analyze the polynucleotides and polypeptides of the invention, along with applicable descriptions, references, and threshold parameters.

DESCRIPTION OF THE INVENTION

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Before the present proteins, nucleotide sequences, and methods are described, it is understood that this invention is not limited to the particular machines, materials and methods described, as these may vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to limit the scope of the present invention which will be limited only by the appended claims.

It must be noted that as used herein and in the appended claims, the singular forms "a," "an," and "the" include plural reference unless the context clearly dictates otherwise. Thus, for example, a reference to "a host cell" includes a plurality of such host cells, and a reference to "an antibody" is a reference to one or more antibodies and equivalents thereof known to those skilled in the art, and so forth.

Unless defined otherwise, all technical and scientific terms used herein have the same meanings as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any machines, materials, and methods similar or equivalent to those described herein can be used to practice or test the present invention, the preferred machines, materials and methods are now described. All publications mentioned herein are cited for the purpose of describing and disclosing

the cell lines, protocols, reagents and vectors which are reported in the publications and which might be used in connection with the invention. Nothing herein is to be construed as an admission that the invention is not entitled to antedate such disclosure by virtue of prior invention.

DEFINITIONS

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"SECP" refers to the amino acid sequences of substantially purified SECP obtained from any species, particularly a mammalian species, including bovine, ovine, porcine, murine, equine, and human, and from any source, whether natural, synthetic, semi-synthetic, or recombinant.

The term "agonist" refers to a molecule which intensifies or mimics the biological activity of SECP. Agonists may include proteins, nucleic acids, carbohydrates, small molecules, or any other compound or composition which modulates the activity of SECP either by directly interacting with SECP or by acting on components of the biological pathway in which SECP participates.

An "allelic variant" is an alternative form of the gene encoding SECP. Allelic variants may result from at least one mutation in the nucleic acid sequence and may result in altered mRNAs or in polypeptides whose structure or function may or may not be altered. A gene may have none, one, or many allelic variants of its naturally occurring form. Common mutational changes which give rise to allelic variants are generally ascribed to natural deletions, additions, or substitutions of nucleotides. Each of these types of changes may occur alone, or in combination with the others, one or more times in a given sequence.

"Altered" nucleic acid sequences encoding SECP include those sequences with deletions, insertions, or substitutions of different nucleotides, resulting in a polypeptide the same as SECP or a polypeptide with at least one functional characteristic of SECP. Included within this definition are polymorphisms which may or may not be readily detectable using a particular oligonucleotide probe of the polynucleotide encoding SECP, and improper or unexpected hybridization to allelic variants, with a locus other than the normal chromosomal locus for the polynucleotide sequence encoding SECP. The encoded protein may also be "altered," and may contain deletions, insertions, or substitutions of amino acid residues which produce a silent change and result in a functionally equivalent SECP. Deliberate amino acid substitutions may be made on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues, as long as the biological or immunological activity of SECP is retained. For example, negatively charged amino acids may include aspartic acid and glutamic acid, and positively charged amino acids may include lysine and arginine. Amino acids with uncharged polar side chains having similar hydrophilicity values may include: asparagine and glutamine; and serine and threonine. Amino acids with uncharged side chains having similar hydrophilicity values may include: leucine, isoleucine, and valine; glycine and alanine; and phenylalanine and tyrosine.

The terms "amino acid" and "amino acid sequence" refer to an oligopeptide, peptide,

polypeptide, or protein sequence, or a fragment of any of these, and to naturally occurring or synthetic molecules. Where "amino acid sequence" is recited to refer to a sequence of a naturally occurring protein molecule, "amino acid sequence" and like terms are not meant to limit the amino acid sequence to the complete native amino acid sequence associated with the recited protein molecule.

"Amplification" relates to the production of additional copies of a nucleic acid sequence.

Amplification is generally carried out using polymerase chain reaction (PCR) technologies well known in the art.

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The term "antagonist" refers to a molecule which inhibits or attenuates the biological activity of SECP. Antagonists may include proteins such as antibodies, nucleic acids, carbohydrates, small molecules, or any other compound or composition which modulates the activity of SECP either by directly interacting with SECP or by acting on components of the biological pathway in which SECP participates.

The term "antibody" refers to intact immunoglobulin molecules as well as to fragments thereof, such as Fab, F(ab')₂, and Fv fragments, which are capable of binding an epitopic determinant. Antibodies that bind SECP polypeptides can be prepared using intact polypeptides or using fragments containing small peptides of interest as the immunizing antigen. The polypeptide or oligopeptide used to immunize an animal (e.g., a mouse, a rat, or a rabbit) can be derived from the translation of RNA, or synthesized chemically, and can be conjugated to a carrier protein if desired. Commonly used carriers that are chemically coupled to peptides include bovine serum albumin, thyroglobulin, and keyhole limpet hemocyanin (KLH). The coupled peptide is then used to immunize the animal.

The term "antigenic determinant" refers to that region of a molecule (i.e., an epitope) that makes contact with a particular antibody. When a protein or a fragment of a protein is used to immunize a host animal, numerous regions of the protein may induce the production of antibodies which bind specifically to antigenic determinants (particular regions or three-dimensional structures on the protein). An antigenic determinant may compete with the intact antigen (i.e., the immunogen used to elicit the immune response) for binding to an antibody.

The term "aptamer" refers to a nucleic acid or oligonucleotide molecule that binds to a specific molecular target. Aptamers are derived from an <u>in vitro</u> evolutionary process (e.g., SELEX (Systematic Evolution of Ligands by EXponential Enrichment), described in U.S. Patent No. 5,270,163), which selects for target-specific aptamer sequences from large combinatorial libraries. Aptamer compositions may be double-stranded or single-stranded, and may include deoxyribonucleotides, ribonucleotides, nucleotide derivatives, or other nucleotide-like molecules. The nucleotide components of an aptamer may have modified sugar groups (e.g., the 2'-OH group of a ribonucleotide may be replaced by 2'-F or 2'-NH₂), which may improve a desired property, e.g., resistance to nucleases or longer lifetime in blood. Aptamers may be conjugated to other molecules,

e.g., a high molecular weight carrier to slow clearance of the aptamer from the circulatory system. Aptamers may be specifically cross-linked to their cognate ligands, e.g., by photo-activation of a cross-linker. (See, e.g., Brody, E.N. and L. Gold (2000) J. Biotechnol. 74:5-13.)

The term "intramer" refers to an aptamer which is expressed <u>in vivo</u>. For example, a vaccinia virus-based RNA expression system has been used to express specific RNA aptamers at high levels in the cytoplasm of leukocytes (Blind, M. et al. (1999) Proc. Natl Acad. Sci. USA 96:3606-3610).

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The term "spiegelmer" refers to an aptamer which includes L-DNA, L-RNA, or other left-handed nucleotide derivatives or nucleotide-like molecules. Aptamers containing left-handed nucleotides are resistant to degradation by naturally occurring enzymes, which normally act on substrates containing right-handed nucleotides.

The term "antisense" refers to any composition capable of base-pairing with the "sense" (coding) strand of a specific nucleic acid sequence. Antisense compositions may include DNA; RNA; peptide nucleic acid (PNA); oligonucleotides having modified backbone linkages such as phosphorothioates, methylphosphonates, or benzylphosphonates; oligonucleotides having modified sugar groups such as 2'-methoxyethyl sugars or 2'-methoxyethoxy sugars; or oligonucleotides having modified bases such as 5-methyl cytosine, 2'-deoxyuracil, or 7-deaza-2'-deoxyguanosine. Antisense molecules may be produced by any method including chemical synthesis or transcription. Once introduced into a cell, the complementary antisense molecule base-pairs with a naturally occurring nucleic acid sequence produced by the cell to form duplexes which block either transcription or translation. The designation "negative" or "minus" can refer to the antisense strand, and the designation "positive" or "plus" can refer to the sense strand of a reference DNA molecule.

The term "biologically active" refers to a protein having structural, regulatory, or biochemical functions of a naturally occurring molecule. Likewise, "immunologically active" or "immunogenic" refers to the capability of the natural, recombinant, or synthetic SECP, or of any oligopeptide thereof, to induce a specific immune response in appropriate animals or cells and to bind with specific antibodies.

"Complementary" describes the relationship between two single-stranded nucleic acid sequences that anneal by base-pairing. For example, 5'-AGT-3' pairs with its complement, 3'-TCA-5'.

A "composition comprising a given polynucleotide sequence" and a "composition comprising a given amino acid sequence" refer broadly to any composition containing the given polynucleotide or amino acid sequence. The composition may comprise a dry formulation or an aqueous solution. Compositions comprising polynucleotide sequences encoding SECP or fragments of SECP may be employed as hybridization probes. The probes may be stored in freeze-dried form and may be associated with a stabilizing agent such as a carbohydrate. In hybridizations, the probe may be

deployed in an aqueous solution containing salts (e.g., NaCl), detergents (e.g., sodium dodecyl sulfate; SDS), and other components (e.g., Denhardt's solution, dry milk, salmon sperm DNA, etc.).

"Consensus sequence" refers to a nucleic acid sequence which has been subjected to repeated DNA sequence analysis to resolve uncalled bases, extended using the XL-PCR kit (Applied Biosystems, Foster City CA) in the 5' and/or the 3' direction, and resequenced, or which has been assembled from one or more overlapping cDNA, EST, or genomic DNA fragments using a computer program for fragment assembly, such as the GELVIEW fragment assembly system (GCG, Madison WI) or Phrap (University of Washington, Seattle WA). Some sequences have been both extended and assembled to produce the consensus sequence.

"Conservative amino acid substitutions" are those substitutions that are predicted to least interfere with the properties of the original protein, i.e., the structure and especially the function of the protein is conserved and not significantly changed by such substitutions. The table below shows amino acids which may be substituted for an original amino acid in a protein and which are regarded as conservative amino acid substitutions.

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15	Original Residue	Conservative Substitution
	Ala	Gly, Ser
	Arg	His, Lys
	Asn	Asp, Gln, His
	Asp	Asn, Glu
20	. Cys	Ala, Ser
	Gln	Asn, Glu, His
	Glu	Asp, Gln, His
	Gly	Ala
	His	Asn, Arg, Gln, Glu
25	Ile	Leu, Val
	Leu	Ile, Val
	Lys	Arg, Gln, Glu
	Met	Leu, Ile
	Phe	His, Met, Leu, Trp, Tyr
30	Ser	Cys, Thr
	Thr	Ser, Val
	Trp	Phe, Tyr
	Tyr	His, Phe, Trp
	Val	Ile, Leu, Thr

Conservative amino acid substitutions generally maintain (a) the structure of the polypeptide backbone in the area of the substitution, for example, as a beta sheet or alpha helical conformation, (b) the charge or hydrophobicity of the molecule at the site of the substitution, and/or (c) the bulk of the side chain.

A "deletion" refers to a change in the amino acid or nucleotide sequence that results in the absence of one or more amino acid residues or nucleotides.

The term "derivative" refers to a chemically modified polynucleotide or polypeptide.

Chemical modifications of a polynucleotide can include, for example, replacement of hydrogen by an alkyl, acyl, hydroxyl, or amino group. A derivative polynucleotide encodes a polypeptide which retains at least one biological or immunological function of the natural molecule. A derivative polypeptide is one modified by glycosylation, pegylation, or any similar process that retains at least one biological or immunological function of the polypeptide from which it was derived.

A "detectable label" refers to a reporter molecule or enzyme that is capable of generating a measurable signal and is covalently or noncovalently joined to a polynucleotide or polypeptide.

"Differential expression" refers to increased or upregulated; or decreased, downregulated, or absent gene or protein expression, determined by comparing at least two different samples. Such comparisons may be carried out between, for example, a treated and an untreated sample, or a diseased and a normal sample.

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"Exon shuffling" refers to the recombination of different coding regions (exons). Since an exon may represent a structural or functional domain of the encoded protein, new proteins may be assembled through the novel reassortment of stable substructures, thus allowing acceleration of the evolution of new protein functions.

A "fragment" is a unique portion of SECP or the polynucleotide encoding SECP which is identical in sequence to but shorter in length than the parent sequence. A fragment may comprise up to the entire length of the defined sequence, minus one nucleotide/amino acid residue. For example, a fragment may comprise from 5 to 1000 contiguous nucleotides or amino acid residues. A fragment used as a probe, primer, antigen, therapeutic molecule, or for other purposes, may be at least 5, 10, 15, 16, 20, 25, 30, 40, 50, 60, 75, 100, 150, 250 or at least 500 contiguous nucleotides or amino acid residues in length. Fragments may be preferentially selected from certain regions of a molecule. For example, a polypeptide fragment may comprise a certain length of contiguous amino acids selected from the first 250 or 500 amino acids (or first 25% or 50%) of a polypeptide as shown in a certain defined sequence. Clearly these lengths are exemplary, and any length that is supported by the specification, including the Sequence Listing, tables, and figures, may be encompassed by the present embodiments.

A fragment of SEQ ID NO:55-108 comprises a region of unique polynucleotide sequence that specifically identifies SEQ ID NO:55-108, for example, as distinct from any other sequence in the genome from which the fragment was obtained. A fragment of SEQ ID NO:55-108 is useful, for example, in hybridization and amplification technologies and in analogous methods that distinguish SEQ ID NO:55-108 from related polynucleotide sequences. The precise length of a fragment of SEQ ID NO:55-108 and the region of SEQ ID NO:55-108 to which the fragment corresponds are routinely determinable by one of ordinary skill in the art based on the intended purpose for the fragment.

A fragment of SEQ ID NO:1-54 is encoded by a fragment of SEQ ID NO:55-108. A

fragment of SEQ ID NO:1-54 comprises a region of unique amino acid sequence that specifically identifies SEQ ID NO:1-54. For example, a fragment of SEQ ID NO:1-54 is useful as an immunogenic peptide for the development of antibodies that specifically recognize SEQ ID NO:1-54. The precise length of a fragment of SEQ ID NO:1-54 and the region of SEQ ID NO:1-54 to which the fragment corresponds are routinely determinable by one of ordinary skill in the art based on the intended purpose for the fragment.

A "full length" polynucleotide sequence is one containing at least a translation initiation codon (e.g., methionine) followed by an open reading frame and a translation termination codon. A "full length" polynucleotide sequence encodes a "full length" polypeptide sequence.

"Homology" refers to sequence similarity or, interchangeably, sequence identity, between two or more polynucleotide sequences or two or more polypeptide sequences.

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The terms "percent identity" and "% identity," as applied to polynucleotide sequences, refer to the percentage of residue matches between at least two polynucleotide sequences aligned using a standardized algorithm. Such an algorithm may insert, in a standardized and reproducible way, gaps in the sequences being compared in order to optimize alignment between two sequences, and therefore achieve a more meaningful comparison of the two sequences.

Percent identity between polynucleotide sequences may be determined using the default parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e sequence alignment program. This program is part of the LASERGENE software package, a suite of molecular biological analysis programs (DNASTAR, Madison WI). CLUSTAL V is described in Higgins, D.G. and P.M. Sharp (1989) CABIOS 5:151-153 and in Higgins, D.G. et al. (1992) CABIOS 8:189-191. For pairwise alignments of polynucleotide sequences, the default parameters are set as follows: Ktuple=2, gap penalty=5, window=4, and "diagonals saved"=4. The "weighted" residue weight table is selected as the default. Percent identity is reported by CLUSTAL V as the "percent similarity" between aligned polynucleotide sequences.

Alternatively, a suite of commonly used and freely available sequence comparison algorithms is provided by the National Center for Biotechnology Information (NCBI) Basic Local Alignment Search Tool (BLAST) (Altschul, S.F. et al. (1990) J. Mol. Biol. 215:403-410), which is available from several sources, including the NCBI, Bethesda, MD, and on the Internet at http://www.ncbi.nlm.nih.gov/BLAST/. The BLAST software suite includes various sequence analysis programs including "blastn," that is used to align a known polynucleotide sequence with other polynucleotide sequences from a variety of databases. Also available is a tool called "BLAST 2 Sequences" that is used for direct pairwise comparison of two nucleotide sequences. "BLAST 2 Sequences" can be accessed and used interactively at http://www.ncbi.nlm.nih.gov/gorf/bl2.html. The "BLAST 2 Sequences" tool can be used for both blastn and blastp (discussed below). BLAST

programs are commonly used with gap and other parameters set to default settings. For example, to compare two nucleotide sequences, one may use blastn with the "BLAST 2 Sequences" tool Version 2.0.12 (April-21-2000) set at default parameters. Such default parameters may be, for example:

Matrix: BLOSUM62

5 Reward for match: 1

Penalty for mismatch: -2

Open Gap: 5 and Extension Gap: 2 penalties

Gap x drop-off: 50

Expect: 10

10 Word Size: 11

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Filter: on

Percent identity may be measured over the length of an entire defined sequence, for example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined sequence, for instance, a fragment of at least 20, at least 30, at least 40, at least 50, at least 70, at least 100, or at least 200 contiguous nucleotides. Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures, or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

Nucleic acid sequences that do not show a high degree of identity may nevertheless encode similar amino acid sequences due to the degeneracy of the genetic code. It is understood that changes in a nucleic acid sequence can be made using this degeneracy to produce multiple nucleic acid sequences that all encode substantially the same protein.

The phrases "percent identity" and "% identity," as applied to polypeptide sequences, refer to the percentage of residue matches between at least two polypeptide sequences aligned using a standardized algorithm. Methods of polypeptide sequence alignment are well-known. Some alignment methods take into account conservative amino acid substitutions. Such conservative substitutions, explained in more detail above, generally preserve the charge and hydrophobicity at the site of substitution, thus preserving the structure (and therefore function) of the polypeptide.

Percent identity between polypeptide sequences may be determined using the default parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e sequence alignment program (described and referenced above). For pairwise alignments of polypeptide sequences using CLUSTAL V, the default parameters are set as follows: Ktuple=1, gap penalty=3, window=5, and "diagonals saved"=5. The PAM250 matrix is selected as the default residue weight table. As with polynucleotide alignments, the percent identity is reported by CLUSTAL V as the "percent similarity" between aligned polypeptide sequence pairs.

Alternatively the NCBI BLAST software suite may be used. For example, for a pairwise comparison of two polypeptide sequences, one may use the "BLAST 2 Sequences" tool Version 2.0.12 (April-21-2000) with blastp set at default parameters. Such default parameters may be, for example:

5 Matrix: BLOSUM62

Open Gap: 11 and Extension Gap: 1 penalties

Gap x drop-off: 50

Expect: 10
Word Size: 3

10 Filter: on

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Percent identity may be measured over the length of an entire defined polypeptide sequence, for example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined polypeptide sequence, for instance, a fragment of at least 15, at least 20, at least 30, at least 40, at least 50, at least 70 or at least 150 contiguous residues. Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

"Human artificial chromosomes" (HACs) are linear microchromosomes which may contain DNA sequences of about 6 kb to 10 Mb in size and which contain all of the elements required for chromosome replication, segregation and maintenance.

The term "humanized antibody" refers to an antibody molecule in which the amino acid sequence in the non-antigen binding regions has been altered so that the antibody more closely resembles a human antibody, and still retains its original binding ability.

"Hybridization" refers to the process by which a polynucleotide strand anneals with a complementary strand through base pairing under defined hybridization conditions. Specific hybridization is an indication that two nucleic acid sequences share a high degree of complementarity. Specific hybridization complexes form under permissive annealing conditions and remain hybridized after the "washing" step(s). The washing step(s) is particularly important in determining the stringency of the hybridization process, with more stringent conditions allowing less non-specific binding, i.e., binding between pairs of nucleic acid strands that are not perfectly matched. Permissive conditions for annealing of nucleic acid sequences are routinely determinable by one of ordinary skill in the art and may be consistent among hybridization experiments, whereas wash conditions may be varied among experiments to achieve the desired stringency, and therefore hybridization specificity. Permissive annealing conditions occur, for example, at 68°C in the presence of about 6 x SSC, about 1% (w/v) SDS, and about 100 μg/ml sheared, denatured salmon sperm DNA.

Generally, stringency of hybridization is expressed, in part, with reference to the temperature under which the wash step is carried out. Such wash temperatures are typically selected to be about 5° C to 20° C lower than the thermal melting point (T_{m}) for the specific sequence at a defined ionic strength and pH. The T_{m} is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. An equation for calculating T_{m} and conditions for nucleic acid hybridization are well known and can be found in Sambrook, J. et al. (1989) Molecular Cloning: A Laboratory Manual, 2^{nd} ed., vol. 1-3, Cold Spring Harbor Press, Plainview NY; specifically see volume 2, chapter 9.

High stringency conditions for hybridization between polynucleotides of the present invention include wash conditions of 68°C in the presence of about 0.2 x SSC and about 0.1% SDS, for 1 hour. Alternatively, temperatures of about 65°C, 60°C, 55°C, or 42°C may be used. SSC concentration may be varied from about 0.1 to 2 x SSC, with SDS being present at about 0.1%. Typically, blocking reagents are used to block non-specific hybridization. Such blocking reagents include, for instance, sheared and denatured salmon sperm DNA at about 100-200 μ g/ml. Organic solvent, such as formamide at a concentration of about 35-50% v/v, may also be used under particular circumstances, such as for RNA:DNA hybridizations. Useful variations on these wash conditions will be readily apparent to those of ordinary skill in the art. Hybridization, particularly under high stringency conditions, may be suggestive of evolutionary similarity between the nucleotides. Such similarity is strongly indicative of a similar role for the nucleotides and their encoded polypeptides.

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The term "hybridization complex" refers to a complex formed between two nucleic acid sequences by virtue of the formation of hydrogen bonds between complementary bases. A hybridization complex may be formed in solution (e.g., C_0 t or R_0 t analysis) or formed between one nucleic acid sequence present in solution and another nucleic acid sequence immobilized on a solid support (e.g., paper, membranes, filters, chips, pins or glass slides, or any other appropriate substrate to which cells or their nucleic acids have been fixed).

The words "insertion" and "addition" refer to changes in an amino acid or nucleotide sequence resulting in the addition of one or more amino acid residues or nucleotides, respectively.

"Immune response" can refer to conditions associated with inflammation, trauma, immune disorders, or infectious or genetic disease, etc. These conditions can be characterized by expression of various factors, e.g., cytokines, chemokines, and other signaling molecules, which may affect cellular and systemic defense systems.

An "immunogenic fragment" is a polypeptide or oligopeptide fragment of SECP which is capable of eliciting an immune response when introduced into a living organism, for example, a mammal. The term "immunogenic fragment" also includes any polypeptide or oligopeptide fragment of SECP which is useful in any of the antibody production methods disclosed herein or known in the

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The term "microarray" refers to an arrangement of a plurality of polynucleotides, polypeptides, or other chemical compounds on a substrate.

The terms "element" and "array element" refer to a polynucleotide, polypeptide, or other chemical compound having a unique and defined position on a microarray.

The term "modulate" refers to a change in the activity of SECP. For example, modulation may cause an increase or a decrease in protein activity, binding characteristics, or any other biological, functional, or immunological properties of SECP.

The phrases "nucleic acid" and "nucleic acid sequence" refer to a nucleotide, oligonucleotide, polynucleotide, or any fragment thereof. These phrases also refer to DNA or RNA of genomic or synthetic origin which may be single-stranded or double-stranded and may represent the sense or the antisense strand, to peptide nucleic acid (PNA), or to any DNA-like or RNA-like material.

"Operably linked" refers to the situation in which a first nucleic acid sequence is placed in a functional relationship with a second nucleic acid sequence. For instance, a promoter is operably linked to a coding sequence if the promoter affects the transcription or expression of the coding sequence. Operably linked DNA sequences may be in close proximity or contiguous and, where necessary to join two protein coding regions, in the same reading frame.

"Peptide nucleic acid" (PNA) refers to an antisense molecule or anti-gene agent which comprises an oligonucleotide of at least about 5 nucleotides in length linked to a peptide backbone of amino acid residues ending in lysine. The terminal lysine confers solubility to the composition. PNAs preferentially bind complementary single stranded DNA or RNA and stop transcript elongation, and may be pegylated to extend their lifespan in the cell.

"Post-translational modification" of an SECP may involve lipidation, glycosylation, phosphorylation, acetylation, racemization, proteolytic cleavage, and other modifications known in the art. These processes may occur synthetically or biochemically. Biochemical modifications will vary by cell type depending on the enzymatic milieu of SECP.

"Probe" refers to nucleic acid sequences encoding SECP, their complements, or fragments thereof, which are used to detect identical, allelic or related nucleic acid sequences. Probes are isolated oligonucleotides or polynucleotides attached to a detectable label or reporter molecule. Typical labels include radioactive isotopes, ligands, chemiluminescent agents, and enzymes. "Primers" are short nucleic acids, usually DNA oligonucleotides, which may be annealed to a target polynucleotide by complementary base-pairing. The primer may then be extended along the target DNA strand by a DNA polymerase enzyme. Primer pairs can be used for amplification (and identification) of a nucleic acid sequence, e.g., by the polymerase chain reaction (PCR).

Probes and primers as used in the present invention typically comprise at least 15 contiguous

nucleotides of a known sequence. In order to enhance specificity, longer probes and primers may also be employed, such as probes and primers that comprise at least 20, 25, 30, 40, 50, 60, 70, 80, 90, 100, or at least 150 consecutive nucleotides of the disclosed nucleic acid sequences. Probes and primers may be considerably longer than these examples, and it is understood that any length supported by the specification, including the tables, figures, and Sequence Listing, may be used.

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Methods for preparing and using probes and primers are described in the references, for example Sambrook, J. et al. (1989) Molecular Cloning: A Laboratory Manual, 2nd ed., vol. 1-3, Cold Spring Harbor Press, Plainview NY; Ausubel, F.M. et al. (1987) Current Protocols in Molecular Biology, Greene Publ. Assoc. & Wiley-Intersciences, New York NY; Innis, M. et al. (1990) PCR Protocols, A Guide to Methods and Applications, Academic Press, San Diego CA. PCR primer pairs can be derived from a known sequence, for example, by using computer programs intended for that purpose such as Primer (Version 0.5, 1991, Whitehead Institute for Biomedical Research, Cambridge MA).

Oligonucleotides for use as primers are selected using software known in the art for such purpose. For example, OLIGO 4.06 software is useful for the selection of PCR primer pairs of up to 100 nucleotides each, and for the analysis of oligonucleotides and larger polynucleotides of up to 5,000 nucleotides from an input polynucleotide sequence of up to 32 kilobases. Similar primer selection programs have incorporated additional features for expanded capabilities. For example, the PrimOU primer selection program (available to the public from the Genome Center at University of Texas South West Medical Center, Dallas TX) is capable of choosing specific primers from megabase sequences and is thus useful for designing primers on a genome-wide scope. The Primer3 primer selection program (available to the public from the Whitehead Institute/MIT Center for Genome Research, Cambridge MA) allows the user to input a "mispriming library," in which sequences to avoid as primer binding sites are user-specified. Primer3 is useful, in particular, for the selection of oligonucleotides for microarrays. (The source code for the latter two primer selection programs may also be obtained from their respective sources and modified to meet the user's specific needs.) The PrimeGen program (available to the public from the UK Human Genome Mapping Project Resource Centre, Cambridge UK) designs primers based on multiple sequence alignments, thereby allowing selection of primers that hybridize to either the most conserved or least conserved regions of aligned nucleic acid sequences. Hence, this program is useful for identification of both unique and conserved oligonucleotides and polynucleotide fragments. The oligonucleotides and polynucleotide fragments identified by any of the above selection methods are useful in hybridization technologies, for example, as PCR or sequencing primers, microarray elements, or specific probes to identify fully or partially complementary polynucleotides in a sample of nucleic acids. Methods of oligonucleotide selection are not limited to those described above.

A "recombinant nucleic acid" is a sequence that is not naturally occurring or has a sequence that is made by an artificial combination of two or more otherwise separated segments of sequence. This artificial combination is often accomplished by chemical synthesis or, more commonly, by the artificial manipulation of isolated segments of nucleic acids, e.g., by genetic engineering techniques such as those described in Sambrook, <u>supra</u>. The term recombinant includes nucleic acids that have been altered solely by addition, substitution, or deletion of a portion of the nucleic acid. Frequently, a recombinant nucleic acid may include a nucleic acid sequence operably linked to a promoter sequence. Such a recombinant nucleic acid may be part of a vector that is used, for example, to transform a cell.

Alternatively, such recombinant nucleic acids may be part of a viral vector, e.g., based on a vaccinia virus, that could be use to vaccinate a mammal wherein the recombinant nucleic acid is expressed, inducing a protective immunological response in the mammal.

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A "regulatory element" refers to a nucleic acid sequence usually derived from untranslated regions of a gene and includes enhancers, promoters, introns, and 5' and 3' untranslated regions (UTRs). Regulatory elements interact with host or viral proteins which control transcription, translation, or RNA stability.

"Reporter molecules" are chemical or biochemical moieties used for labeling a nucleic acid, amino acid, or antibody. Reporter molecules include radionuclides; enzymes; fluorescent, chemiluminescent, or chromogenic agents; substrates; cofactors; inhibitors; magnetic particles; and other moieties known in the art.

An "RNA equivalent," in reference to a DNA sequence, is composed of the same linear sequence of nucleotides as the reference DNA sequence with the exception that all occurrences of the nitrogenous base thymine are replaced with uracil, and the sugar backbone is composed of ribose instead of deoxyribose.

The term "sample" is used in its broadest sense. A sample suspected of containing SECP, nucleic acids encoding SECP, or fragments thereof may comprise a bodily fluid; an extract from a cell, chromosome, organelle, or membrane isolated from a cell; a cell; genomic DNA, RNA, or cDNA, in solution or bound to a substrate; a tissue; a tissue print; etc.

The terms "specific binding" and "specifically binding" refer to that interaction between a protein or peptide and an agonist, an antibody, an antagonist, a small molecule, or any natural or synthetic binding composition. The interaction is dependent upon the presence of a particular structure of the protein, e.g., the antigenic determinant or epitope, recognized by the binding molecule. For example, if an antibody is specific for epitope "A," the presence of a polypeptide comprising the epitope A, or the presence of free unlabeled A, in a reaction containing free labeled A and the antibody will reduce the amount of labeled A that binds to the antibody.

The term "substantially purified" refers to nucleic acid or amino acid sequences that are removed from their natural environment and are isolated or separated, and are at least 60% free, preferably at least 75% free, and most preferably at least 90% free from other components with which they are naturally associated.

A "substitution" refers to the replacement of one or more amino acid residues or nucleotides by different amino acid residues or nucleotides, respectively.

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"Substrate" refers to any suitable rigid or semi-rigid support including membranes, filters, chips, slides, wafers, fibers, magnetic or nonmagnetic beads, gels, tubing, plates, polymers, microparticles and capillaries. The substrate can have a variety of surface forms, such as wells, trenches, pins, channels and pores, to which polynucleotides or polypeptides are bound.

A "transcript image" or "expression profile" refers to the collective pattern of gene expression by a particular cell type or tissue under given conditions at a given time.

"Transformation" describes a process by which exogenous DNA is introduced into a recipient cell. Transformation may occur under natural or artificial conditions according to various methods well known in the art, and may rely on any known method for the insertion of foreign nucleic acid sequences into a prokaryotic or eukaryotic host cell. The method for transformation is selected based on the type of host cell being transformed and may include, but is not limited to, bacteriophage or viral infection, electroporation, heat shock, lipofection, and particle bombardment. The term "transformed cells" includes stably transformed cells in which the inserted DNA is capable of replication either as an autonomously replicating plasmid or as part of the host chromosome, as well as transiently transformed cells which express the inserted DNA or RNA for limited periods of time.

A "transgenic organism," as used herein, is any organism, including but not limited to animals and plants, in which one or more of the cells of the organism contains heterologous nucleic acid introduced by way of human intervention, such as by transgenic techniques well known in the art. The nucleic acid is introduced into the cell, directly or indirectly by introduction into a precursor of the cell, by way of deliberate genetic manipulation, such as by microinjection or by infection with a recombinant virus. The term genetic manipulation does not include classical cross-breeding, or in vitro fertilization, but rather is directed to the introduction of a recombinant DNA molecule. The transgenic organisms contemplated in accordance with the present invention include bacteria, cyanobacteria, fungi, plants and animals. The isolated DNA of the present invention can be introduced into the host by methods known in the art, for example infection, transfection, transformation or transconjugation. Techniques for transferring the DNA of the present invention into such organisms are widely known and provided in references such as Sambrook et al. (1989), supra.

A "variant" of a particular nucleic acid sequence is defined as a nucleic acid sequence having

at least 40% sequence identity to the particular nucleic acid sequence over a certain length of one of the nucleic acid sequences using blastn with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such a pair of nucleic acids may show, for example, at least 50%, at least 60%, at least 70%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% or greater sequence identity over a certain defined length. A variant may be described as, for example, an "allelic" (as defined above), "splice," "species," or "polymorphic" variant. A splice variant may have significant identity to a reference molecule, but will generally have a greater or lesser number of polynucleotides due to alternate splicing of exons during mRNA processing. The corresponding polypeptide may possess additional functional domains or lack domains that are present in the reference molecule. Species variants are polynucleotide sequences that vary from one species to another. The resulting polypeptides will generally have significant amino acid identity relative to each other. A polymorphic variant is a variation in the polynucleotide sequence of a particular gene between individuals of a given species. Polymorphic variants also may encompass "single nucleotide polymorphisms" (SNPs) in which the polynucleotide sequence varies by one nucleotide base. The presence of SNPs may be indicative of, for example, a certain population, a disease state, or a propensity for a disease state.

A "variant" of a particular polypeptide sequence is defined as a polypeptide sequence having at least 40% sequence identity to the particular polypeptide sequence over a certain length of one of the polypeptide sequences using blastp with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such a pair of polypeptides may show, for example, at least 50%, at least 60%, at least 70%, at least 80%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% or greater sequence identity over a certain defined length of one of the polypeptides.

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THE INVENTION

The invention is based on the discovery of new human secreted proteins (SECP), the polynucleotides encoding SECP, and the use of these compositions for the diagnosis, treatment, or prevention of cell proliferative, autoimmune/inflammatory, cardiovascular, neurological, and developmental disorders.

Table 1 summarizes the nomenclature for the full length polynucleotide and polypeptide sequences of the invention. Each polynucleotide and its corresponding polypeptide are correlated to a single Incyte project identification number (Incyte Project ID). Each polypeptide sequence is denoted by both a polypeptide sequence identification number (Polypeptide SEQ ID NO:) and an Incyte polypeptide sequence number (Incyte Polypeptide ID) as shown. Each polynucleotide sequence is

denoted by both a polynucleotide sequence identification number (Polynucleotide SEQ ID NO:) and an Incyte polynucleotide consensus sequence number (Incyte Polynucleotide ID) as shown.

Table 2 shows sequences with homology to the polypeptides of the invention as identified by BLAST analysis against the GenBank protein (genpept) database. Columns 1 and 2 show the polypeptide sequence identification number (Polypeptide SEQ ID NO:) and the corresponding Incyte polypeptide sequence number (Incyte Polypeptide ID) for polypeptides of the invention. Column 3 shows the GenBank identification number (GenBank ID NO:) of the nearest GenBank homolog. Column 4 shows the probability scores for the matches between each polypeptide and its homolog(s). Column 5 shows the annotation of the GenBank homolog(s) along with relevant citations where applicable, all of which are expressly incorporated by reference herein.

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Table 3 shows various structural features of the polypeptides of the invention. Columns 1 and 2 show the polypeptide sequence identification number (SEQ ID NO:) and the corresponding Incyte polypeptide sequence number (Incyte Polypeptide ID) for each polypeptide of the invention. Column 3 shows the number of amino acid residues in each polypeptide. Column 4 shows potential phosphorylation sites, and column 5 shows potential glycosylation sites, as determined by the MOTIFS program of the GCG sequence analysis software package (Genetics Computer Group, Madison WI). Column 6 shows amino acid residues comprising signature sequences, domains, and motifs, including the locations of signal peptides (as indicated by "Signal Peptide" and/or "signal_cleavage".). Column 7 shows analytical methods for protein structure/function analysis and in some cases, searchable databases to which the analytical methods were applied.

Together, Tables 2 and 3 summarize the properties of polypeptides of the invention, and these properties establish that the claimed polypeptides are secreted proteins. For example, SEQ ID NO:2 is 99% identical to a novel human AMP-binding enzyme similar to acetyl-coenzyme A synthethase (acetate-coA ligase) (GenBank ID g6996429) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 5.8e-262, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:2 also contains an AMP-binding domain signature as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS, MOTIFS, and PROFILESCAN analyses provide further corroborative evidence that SEQ ID NO:2 is an AMP-binding enzyme (note that "AMP-binding domains" are shared regions of sequence similarity within a number of prokaryotic and eukaryotic enzymes which most likely act via an ATP-dependent covalent binding of AMP to their substrate, PROSITE:PDOC00427).

As a further example, SEQ ID NO:3 is 33% identical from residues E44 to L530 to bovine
PDI (protein disulfide isomerase) (GenBank ID g163497) as determined by the Basic Local

Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 1.1e-70, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:3 also contains a thioredoxin domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS and PROFILESCAN analyses provide further corroborative evidence that SEQ ID NO:3 is a protein disulfide isomerase.

As a further example, SEQ ID NO:4 is 56% identical to human preceruloplasmin (GenBank ID g180256) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 0.0, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:4 contains a signal peptide and a multicopper oxidase active site domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) The presence of this domain is confirmed by BLIMPS, MOTIFS, and PROFILESCAN analyses, providing further corroborative evidence that SEQ ID NO:4 is a secreted multicopper oxidase.

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In another example, SEQ ID NO:16 is 79% identical to human growth hormone hGH-V2 (GenBank ID g183178) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 5.6e-106, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:16 also contains a signal peptide and a somatotropin hormone family signature as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) The presence of these motifs is confirmed by BLIMPS, MOTIFS, SPSCAN, and PROFILESCAN analyses, providing further corroborative evidence that SEQ ID NO:16 is a secreted hormone.

As a further example, SEQ ID NO:27 is 49% identical to mouse Fca/m receptor (GenBank ID g11071950) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 2.2e-115, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:27 also contains an immunoglobulin domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from additional BLAST analyses provide further corroborative evidence that SEQ ID NO:27 is an immunoglobulin domain-containing receptor.

In another example, SEQ ID NO:41 is 99% identical to human chordin (GenBank ID g3822218) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 0.0, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:41 also contains a von Willebrand factor growth

regulator domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from MOTIFS analyses provide further corroborative evidence that SEQ ID NO:41 is a growth regulation molecule.

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SEQ ID NO:50 contains a signal peptide as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 2.) The presence of the signal peptide is confirmed by data from SPSCAN. SEQ ID NO:1, SEQ ID NO:5-15, SEQ ID NO:17-26, SEQ ID NO:28-40, SEQ ID NO:42-49 and SEQ ID NO:51-54, which were analyzed and annotated in a similar manner, all contain signal peptides as determined by SPSCAN or HMMER analysis. The algorithms and parameters for the analysis of SEQ ID NO:1-54 are described in Table 7.

As shown in Table 4, the full length polynucleotide sequences of the present invention were assembled using cDNA sequences or coding (exon) sequences derived from genomic DNA, or any combination of these two types of sequences. Column 1 lists the polynucleotide sequence identification number (Polynucleotide SEQ ID NO:), the corresponding Incyte polynucleotide consensus sequence number (Incyte ID) for each polynucleotide of the invention, and the length of each polynucleotide sequence in basepairs. Column 2 shows the nucleotide start (5') and stop (3') positions of the cDNA and/or genomic sequences used to assemble the full length polynucleotide sequences of the invention, and of fragments of the polynucleotide sequences which are useful, for example, in hybridization or amplification technologies that identify SEQ ID NO:55-108 or that distinguish between SEQ ID NO:55-108 and related polynucleotide sequences.

The polynucleotide fragments described in Column 2 of Table 4 may refer specifically, for example, to Incyte cDNAs derived from tissue-specific cDNA libraries or from pooled cDNA libraries. Alternatively, the polynucleotide fragments described in column 2 may refer to GenBank cDNAs or ESTs which contributed to the assembly of the full length polynucleotide sequences. In addition, the polynucleotide fragments described in column 2 may identify sequences derived from the ENSEMBL (The Sanger Centre, Cambridge, UK) database (i.e., those sequences including the designation "ENST"). Alternatively, the polynucleotide fragments described in column 2 may be derived from the NCBI RefSeq Nucleotide Sequence Records Database (i.e., those sequences including the designation "NM" or "NT") or the NCBI RefSeq Protein Sequence Records (i.e., those sequences including the designation "NP"). Alternatively, the polynucleotide fragments described in column 2 may refer to assemblages of both cDNA and Genscan-predicted exons brought together by an "exon stitching" algorithm. For example, a polynucleotide sequence identified as FL_XXXXXX_N₁_N₂_YYYYY_N₃_N₄ represents a "stitched" sequence in which XXXXXX is the identification number of the cluster of sequences to which the algorithm was applied, and YYYYY is

the number of the prediction generated by the algorithm, and $N_{1,2,3...}$, if present, represent specific exons that may have been manually edited during analysis (See Example V). Alternatively, the polynucleotide fragments in column 2 may refer to assemblages of exons brought together by an "exon-stretching" algorithm. For example, a polynucleotide sequence identified as

5 FLXXXXXX_gAAAAA_gBBBBB_1_N is a "stretched" sequence, with XXXXXX being the Incyte project identification number, gAAAAA being the GenBank identification number of the human genomic sequence to which the "exon-stretching" algorithm was applied, gBBBBB being the GenBank identification number or NCBI RefSeq identification number of the nearest GenBank protein homolog, and N referring to specific exons (See Example V). In instances where a RefSeq sequence was used as a protein homolog for the "exon-stretching" algorithm, a RefSeq identifier (denoted by "NM," "NP," or "NT") may be used in place of the GenBank identifier (i.e., gBBBBB).

Alternatively, a prefix identifies component sequences that were hand-edited, predicted from genomic DNA sequences, or derived from a combination of sequence analysis methods. The following Table lists examples of component sequence prefixes and corresponding sequence analysis methods associated with the prefixes (see Example IV and Example V).

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Prefix	Type of analysis and/or examples of programs
GNN, GFG, ENST	Exon prediction from genomic sequences using, for example, GENSCAN (Stanford University, CA, USA) or FGENES
ENST.	(Computer Genomics Group, The Sanger Centre, Cambridge, UK).
GBI	Hand-edited analysis of genomic sequences.
FL	Stitched or stretched genomic sequences (see Example V).
INCY	Full length transcript and exon prediction from mapping of EST sequences to the genome. Genomic location and EST composition
	data are combined to predict the exons and resulting transcript.

In some cases, Incyte cDNA coverage redundant with the sequence coverage shown in Table 4 was obtained to confirm the final consensus polynucleotide sequence, but the relevant Incyte cDNA identification numbers are not shown.

Table 5 shows the representative cDNA libraries for those full length polynucleotide sequences which were assembled using Incyte cDNA sequences. The representative cDNA library is the Incyte cDNA library which is most frequently represented by the Incyte cDNA sequences which were used to assemble and confirm the above polynucleotide sequences. The tissues and vectors which were used to construct the cDNA libraries shown in Table 5 are described in Table 6.

The invention also encompasses SECP variants. A preferred SECP variant is one which has at least about 80%, or alternatively at least about 90%, or even at least about 95% amino acid

sequence identity to the SECP amino acid sequence, and which contains at least one functional or structural characteristic of SECP.

The invention also encompasses polynucleotides which encode SECP. In a particular embodiment, the invention encompasses a polynucleotide sequence comprising a sequence selected from the group consisting of SEQ ID NO:55-108, which encodes SECP. The polynucleotide sequences of SEQ ID NO:55-108, as presented in the Sequence Listing, embrace the equivalent RNA sequences, wherein occurrences of the nitrogenous base thymine are replaced with uracil, and the sugar backbone is composed of ribose instead of deoxyribose.

The invention also encompasses a variant of a polynucleotide sequence encoding SECP. In particular, such a variant polynucleotide sequence will have at least about 70%, or alternatively at least about 85%, or even at least about 95% polynucleotide sequence identity to the polynucleotide sequence encoding SECP. A particular aspect of the invention encompasses a variant of a polynucleotide sequence comprising a sequence selected from the group consisting of SEQ ID NO:55-108 which has at least about 70%, or alternatively at least about 85%, or even at least about 15 95% polynucleotide sequence identity to a nucleic acid sequence selected from the group consisting of SEQ ID NO:55-108. Any one of the polynucleotide variants described above can encode an amino acid sequence which contains at least one functional or structural characteristic of SECP.

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In addition, or in the alternative, a polynucleotide variant of the invention is a splice variant of a polynucleotide sequence encoding SECP. A splice variant may have portions which have significant sequence identity to the polynucleotide sequence encoding SECP, but will generally have a greater or lesser number of polynucleotides due to additions or deletions of blocks of sequence arising from alternate splicing of exons during mRNA processing. A splice variant may have less than about 70%, or alternatively less than about 60%, or alternatively less than about 50% polynucleotide sequence identity to the polynucleotide sequence encoding SECP over its entire length; however, portions of the splice variant will have at least about 70%, or alternatively at least about 85%, or alternatively at least about 95%, or alternatively 100% polynucleotide sequence identity to portions of the polynucleotide sequence encoding SECP. For example, a polynucleotide comprising a sequence of SEQ ID NO:108 is a splice variant of a polynucleotide comprising a sequence of SEQ ID NO:94. Any one of the splice variants described above can encode an amino acid sequence which contains at least one functional or structural characteristic of SECP.

It will be appreciated by those skilled in the art that as a result of the degeneracy of the genetic code, a multitude of polynucleotide sequences encoding SECP, some bearing minimal similarity to the polynucleotide sequences of any known and naturally occurring gene, may be produced. Thus, the invention contemplates each and every possible variation of polynucleotide sequence that could be made by selecting combinations based on possible codon choices. These

combinations are made in accordance with the standard triplet genetic code as applied to the polynucleotide sequence of naturally occurring SECP, and all such variations are to be considered as being specifically disclosed.

Although nucleotide sequences which encode SECP and its variants are generally capable of hybridizing to the nucleotide sequence of the naturally occurring SECP under appropriately selected conditions of stringency, it may be advantageous to produce nucleotide sequences encoding SECP or its derivatives possessing a substantially different codon usage, e.g., inclusion of non-naturally occurring codons. Codons may be selected to increase the rate at which expression of the peptide occurs in a particular prokaryotic or eukaryotic host in accordance with the frequency with which particular codons are utilized by the host. Other reasons for substantially altering the nucleotide sequence encoding SECP and its derivatives without altering the encoded amino acid sequences include the production of RNA transcripts having more desirable properties, such as a greater half-life, than transcripts produced from the naturally occurring sequence.

The invention also encompasses production of DNA sequences which encode SECP and SECP derivatives, or fragments thereof, entirely by synthetic chemistry. After production, the synthetic sequence may be inserted into any of the many available expression vectors and cell systems using reagents well known in the art. Moreover, synthetic chemistry may be used to introduce mutations into a sequence encoding SECP or any fragment thereof.

Also encompassed by the invention are polynucleotide sequences that are capable of hybridizing to the claimed polynucleotide sequences, and, in particular, to those shown in SEQ ID NO:55-108 and fragments thereof under various conditions of stringency. (See, e.g., Wahl, G.M. and S.L. Berger (1987) Methods Enzymol. 152:399-407; Kimmel, A.R. (1987) Methods Enzymol. 152:507-511.) Hybridization conditions, including annealing and wash conditions, are described in "Definitions."

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Methods for DNA sequencing are well known in the art and may be used to practice any of the embodiments of the invention. The methods may employ such enzymes as the Klenow fragment of DNA polymerase I, SEQUENASE (US Biochemical, Cleveland OH), Taq polymerase (Applied Biosystems), thermostable T7 polymerase (Amersham Pharmacia Biotech, Piscataway NJ), or combinations of polymerases and proofreading exonucleases such as those found in the ELONGASE amplification system (Life Technologies, Gaithersburg MD). Preferably, sequence preparation is automated with machines such as the MICROLAB 2200 liquid transfer system (Hamilton, Reno NV), PTC200 thermal cycler (MJ Research, Watertown MA) and ABI CATALYST 800 thermal cycler (Applied Biosystems). Sequencing is then carried out using either the ABI 373 or 377 DNA sequencing system (Applied Biosystems), the MEGABACE 1000 DNA sequencing system (Molecular Dynamics, Sunnyvale CA), or other systems known in the art. The resulting sequences

are analyzed using a variety of algorithms which are well known in the art. (See, e.g., Ausubel, F.M. (1997) Short Protocols in Molecular Biology, John Wiley & Sons, New York NY, unit 7.7; Meyers, R.A. (1995) Molecular Biology and Biotechnology, Wiley VCH, New York NY, pp. 856-853.)

The nucleic acid sequences encoding SECP may be extended utilizing a partial nucleotide sequence and employing various PCR-based methods known in the art to detect upstream sequences, such as promoters and regulatory elements. For example, one method which may be employed, restriction-site PCR, uses universal and nested primers to amplify unknown sequence from genomic DNA within a cloning vector. (See, e.g., Sarkar, G. (1993) PCR Methods Applic. 2:318-322.) Another method, inverse PCR, uses primers that extend in divergent directions to amplify unknown sequence from a circularized template. The template is derived from restriction fragments comprising a known genomic locus and surrounding sequences. (See, e.g., Triglia, T. et al. (1988) Nucleic Acids Res. 16:8186.) A third method, capture PCR, involves PCR amplification of DNA fragments adjacent to known sequences in human and yeast artificial chromosome DNA. (See, e.g., Lagerstrom, M. et al. (1991) PCR Methods Applic. 1:111-119.) In this method, multiple restriction enzyme digestions and ligations may be used to insert an engineered double-stranded sequence into a region of unknown sequence before performing PCR. Other methods which may be used to retrieve unknown sequences are known in the art. (See, e.g., Parker, J.D. et al. (1991) Nucleic Acids Res. 19:3055-3060). Additionally, one may use PCR, nested primers, and PROMOTERFINDER libraries (Clontech, Palo Alto CA) to walk genomic DNA. This procedure avoids the need to screen libraries and is useful in finding intron/exon junctions. For all PCR-based methods, primers may be designed using commercially available software, such as OLIGO 4.06 primer analysis software (National Biosciences, Plymouth MN) or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the template at temperatures of about 68°C to 72°C.

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When screening for full length cDNAs, it is preferable to use libraries that have been size-selected to include larger cDNAs. In addition, random-primed libraries, which often include sequences containing the 5' regions of genes, are preferable for situations in which an oligo d(T) library does not yield a full-length cDNA. Genomic libraries may be useful for extension of sequence into 5' non-transcribed regulatory regions.

Capillary electrophoresis systems which are commercially available may be used to analyze the size or confirm the nucleotide sequence of sequencing or PCR products. In particular, capillary sequencing may employ flowable polymers for electrophoretic separation, four different nucleotide-specific, laser-stimulated fluorescent dyes, and a charge coupled device camera for detection of the emitted wavelengths. Output/light intensity may be converted to electrical signal using appropriate software (e.g., GENOTYPER and SEQUENCE NAVIGATOR, Applied Biosystems), and the entire

process from loading of samples to computer analysis and electronic data display may be computer controlled. Capillary electrophoresis is especially preferable for sequencing small DNA fragments which may be present in limited amounts in a particular sample.

In another embodiment of the invention, polynucleotide sequences or fragments thereof which encode SECP may be cloned in recombinant DNA molecules that direct expression of SECP, or fragments or functional equivalents thereof, in appropriate host cells. Due to the inherent degeneracy of the genetic code, other DNA sequences which encode substantially the same or a functionally equivalent amino acid sequence may be produced and used to express SECP.

The nucleotide sequences of the present invention can be engineered using methods generally known in the art in order to alter SECP-encoding sequences for a variety of purposes including, but not limited to, modification of the cloning, processing, and/or expression of the gene product. DNA shuffling by random fragmentation and PCR reassembly of gene fragments and synthetic oligonucleotides may be used to engineer the nucleotide sequences. For example, oligonucleotide-mediated site-directed mutagenesis may be used to introduce mutations that create new restriction sites, alter glycosylation patterns, change codon preference, produce splice variants, and so forth.

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The nucleotides of the present invention may be subjected to DNA shuffling techniques such as MOLECULARBREEDING (Maxygen Inc., Santa Clara CA; described in U.S. Patent No. 5,837,458; Chang, C.-C. et al. (1999) Nat. Biotechnol. 17:793-797; Christians, F.C. et al. (1999) Nat. Biotechnol. 17:259-264; and Crameri, A. et al. (1996) Nat. Biotechnol. 14:315-319) to alter or improve the biological properties of SECP, such as its biological or enzymatic activity or its ability to bind to other molecules or compounds. DNA shuffling is a process by which a library of gene variants is produced using PCR-mediated recombination of gene fragments. The library is then subjected to selection or screening procedures that identify those gene variants with the desired properties. These preferred variants may then be pooled and further subjected to recursive rounds of DNA shuffling and selection/screening. Thus, genetic diversity is created through "artificial" breeding and rapid molecular evolution. For example, fragments of a single gene containing random point mutations may be recombined, screened, and then reshuffled until the desired properties are optimized. Alternatively, fragments of a given gene may be recombined with fragments of homologous genes in the same gene family, either from the same or different species, thereby maximizing the genetic diversity of multiple naturally occurring genes in a directed and controllable manner.

In another embodiment, sequences encoding SECP may be synthesized, in whole or in part, using chemical methods well known in the art. (See, e.g., Caruthers, M.H. et al. (1980) Nucleic Acids Symp. Ser. 7:215-223; and Horn, T. et al. (1980) Nucleic Acids Symp. Ser. 7:225-232.)

Alternatively, SECP itself or a fragment thereof may be synthesized using chemical methods. For

example, peptide synthesis can be performed using various solution-phase or solid-phase techniques. (See, e.g., Creighton, T. (1984) Proteins, Structures and Molecular Properties, WH Freeman, New York NY, pp. 55-60; and Roberge, J.Y. et al. (1995) Science 269:202-204.) Automated synthesis may be achieved using the ABI 431A peptide synthesizer (Applied Biosystems). Additionally, the amino acid sequence of SECP, or any part thereof, may be altered during direct synthesis and/or combined with sequences from other proteins, or any part thereof, to produce a variant polypeptide or a polypeptide having a sequence of a naturally occurring polypeptide.

The peptide may be substantially purified by preparative high performance liquid chromatography. (See, e.g., Chiez, R.M. and F.Z. Regnier (1990) Methods Enzymol. 182:392-421.) The composition of the synthetic peptides may be confirmed by amino acid analysis or by sequencing. (See, e.g., Creighton, supra, pp. 28-53.)

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In order to express a biologically active SECP, the nucleotide sequences encoding SECP or derivatives thereof may be inserted into an appropriate expression vector, i.e., a vector which contains the necessary elements for transcriptional and translational control of the inserted coding sequence in a suitable host. These elements include regulatory sequences, such as enhancers, constitutive and inducible promoters, and 5' and 3' untranslated regions in the vector and in polynucleotide sequences encoding SECP. Such elements may vary in their strength and specificity. Specific initiation signals may also be used to achieve more efficient translation of sequences encoding SECP. Such signals include the ATG initiation codon and adjacent sequences, e.g. the Kozak sequence. In cases where sequences encoding SECP and its initiation codon and upstream regulatory sequences are inserted into the appropriate expression vector, no additional transcriptional or translational control signals may be needed. However, in cases where only coding sequence, or a fragment thereof, is inserted, exogenous translational control signals including an in-frame ATG initiation codon should be provided by the vector. Exogenous translational elements and initiation codons may be of various origins, both natural and synthetic. The efficiency of expression may be enhanced by the inclusion of enhancers appropriate for the particular host cell system used. (See, e.g., Scharf, D. et al. (1994) Results Probl. Cell Differ. 20:125-162.)

Methods which are well known to those skilled in the art may be used to construct expression vectors containing sequences encoding SECP and appropriate transcriptional and translational control elements. These methods include <u>in vitro</u> recombinant DNA techniques, synthetic techniques, and <u>in vivo</u> genetic recombination. (See, e.g., Sambrook, J. et al. (1989) <u>Molecular Cloning, A Laboratory Manual</u>, Cold Spring Harbor Press, Plainview NY, ch. 4, 8, and 16-17; Ausubel, F.M. et al. (1995) <u>Current Protocols in Molecular Biology</u>, John Wiley & Sons, New York NY, ch. 9, 13, and 16.)

A variety of expression vector/host systems may be utilized to contain and express sequences encoding SECP. These include, but are not limited to, microorganisms such as bacteria transformed

with recombinant bacteriophage, plasmid, or cosmid DNA expression vectors; yeast transformed with yeast expression vectors; insect cell systems infected with viral expression vectors (e.g., baculovirus); plant cell systems transformed with viral expression vectors (e.g., cauliflower mosaic virus, CaMV, or tobacco mosaic virus, TMV) or with bacterial expression vectors (e.g., Ti or pBR322 plasmids); or animal cell systems. (See, e.g., Sambrook, supra; Ausubel, supra; Van Heeke, G. and S.M. Schuster (1989) J. Biol. Chem. 264:5503-5509; Engelhard, E.K. et al. (1994) Proc. Natl. Acad. Sci. USA 91:3224-3227; Sandig, V. et al. (1996) Hum. Gene Ther. 7:1937-1945; Takamatsu, N. (1987) EMBO J. 6:307-311; The McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York NY, pp. 191-196; Logan, J. and T. Shenk (1984) Proc. Natl. Acad. Sci. USA 81:3655-3659; and Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355.) Expression vectors derived from retroviruses, adenoviruses, or herpes or vaccinia viruses, or from various bacterial plasmids, may be used for delivery of nucleotide sequences to the targeted organ, tissue, or cell population. (See, e.g., Di Nicola, M. et al. (1998) Cancer Gen. Ther. 5(6):350-356; Yu, M. et al. (1993) Proc. Natl. Acad. Sci. USA 90(13):6340-6344; Buller, R.M. et al. (1985) Nature 317(6040):813-815; McGregor, D.P. et al. (1994) Mol. Immunol. 31(3):219-226; and Verma, I.M. and N. Somia (1997) Nature 389:239-242.) The invention is not limited by the host cell employed.

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In bacterial systems, a number of cloning and expression vectors may be selected depending upon the use intended for polynucleotide sequences encoding SECP. For example, routine cloning, subcloning, and propagation of polynucleotide sequences encoding SECP can be achieved using a multifunctional <u>E. coli</u> vector such as PBLUESCRIPT (Stratagene, La Jolla CA) or PSPORT1 plasmid (Life Technologies). Ligation of sequences encoding SECP into the vector's multiple cloning site disrupts the *lacZ* gene, allowing a colorimetric screening procedure for identification of transformed bacteria containing recombinant molecules. In addition, these vectors may be useful for in vitro transcription, dideoxy sequencing, single strand rescue with helper phage, and creation of nested deletions in the cloned sequence. (See, e.g., Van Heeke, G. and S.M. Schuster (1989) J. Biol. Chem. 264:5503-5509.) When large quantities of SECP are needed, e.g. for the production of antibodies, vectors which direct high level expression of SECP may be used. For example, vectors containing the strong, inducible SP6 or T7 bacteriophage promoter may be used.

Yeast expression systems may be used for production of SECP. A number of vectors containing constitutive or inducible promoters, such as alpha factor, alcohol oxidase, and PGH promoters, may be used in the yeast <u>Saccharomyces cerevisiae</u> or <u>Pichia pastoris</u>. In addition, such vectors direct either the secretion or intracellular retention of expressed proteins and enable integration of foreign sequences into the host genome for stable propagation. (See, e.g., Ausubel, 1995, <u>supra</u>; Bitter, G.A. et al. (1987) Methods Enzymol. 153:516-544; and Scorer, C.A. et al. (1994) Bio/Technology 12:181-184.)

Plant systems may also be used for expression of SECP. Transcription of sequences encoding SECP may be driven by viral promoters, e.g., the 35S and 19S promoters of CaMV used alone or in combination with the omega leader sequence from TMV (Takamatsu, N. (1987) EMBO J. 6:307-311). Alternatively, plant promoters such as the small subunit of RUBISCO or heat shock promoters may be used. (See, e.g., Coruzzi, G. et al. (1984) EMBO J. 3:1671-1680; Broglie, R. et al. (1984) Science 224:838-843; and Winter, J. et al. (1991) Results Probl. Cell Differ. 17:85-105.) These constructs can be introduced into plant cells by direct DNA transformation or pathogen-mediated transfection. (See, e.g., The McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York NY, pp. 191-196.)

In mammalian cells, a number of viral-based expression systems may be utilized. In cases where an adenovirus is used as an expression vector, sequences encoding SECP may be ligated into an adenovirus transcription/translation complex consisting of the late promoter and tripartite leader sequence. Insertion in a non-essential E1 or E3 region of the viral genome may be used to obtain infective virus which expresses SECP in host cells. (See, e.g., Logan, J. and T. Shenk (1984) Proc. Natl. Acad. Sci. USA 81:3655-3659.) In addition, transcription enhancers, such as the Rous sarcoma virus (RSV) enhancer, may be used to increase expression in mammalian host cells. SV40 or EBV-based vectors may also be used for high-level protein expression.

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Human artificial chromosomes (HACs) may also be employed to deliver larger fragments of DNA than can be contained in and expressed from a plasmid. HACs of about 6 kb to 10 Mb are constructed and delivered via conventional delivery methods (liposomes, polycationic amino polymers, or vesicles) for therapeutic purposes. (See, e.g., Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355.)

For long term production of recombinant proteins in mammalian systems, stable expression of SECP in cell lines is preferred. For example, sequences encoding SECP can be transformed into cell lines using expression vectors which may contain viral origins of replication and/or endogenous expression elements and a selectable marker gene on the same or on a separate vector. Following the introduction of the vector, cells may be allowed to grow for about 1 to 2 days in enriched media before being switched to selective media. The purpose of the selectable marker is to confer resistance to a selective agent, and its presence allows growth and recovery of cells which successfully express the introduced sequences. Resistant clones of stably transformed cells may be propagated using tissue culture techniques appropriate to the cell type.

Any number of selection systems may be used to recover transformed cell lines. These include, but are not limited to, the herpes simplex virus thymidine kinase and adenine phosphoribosyltransferase genes, for use in tk and apr cells, respectively. (See, e.g., Wigler, M. et al. (1977) Cell 11:223-232; Lowy, I. et al. (1980) Cell 22:817-823.) Also, antimetabolite, antibiotic,

or herbicide resistance can be used as the basis for selection. For example, dhfr confers resistance to methotrexate; neo confers resistance to the aminoglycosides neomycin and G-418; and als and pat confer resistance to chlorsulfuron and phosphinotricin acetyltransferase, respectively. (See, e.g., Wigler, M. et al. (1980) Proc. Natl. Acad. Sci. USA 77:3567-3570; Colbere-Garapin, F. et al. (1981) J. Mol. Biol. 150:1-14.) Additional selectable genes have been described, e.g., trpB and hisD, which alter cellular requirements for metabolites. (See, e.g., Hartman, S.C. and R.C. Mulligan (1988) Proc. Natl. Acad. Sci. USA 85:8047-8051.) Visible markers, e.g., anthocyanins, green fluorescent proteins (GFP; Clontech), ß glucuronidase and its substrate ß-glucuronide, or luciferase and its substrate luciferin may be used. These markers can be used not only to identify transformants, but also to quantify the amount of transient or stable protein expression attributable to a specific vector system. (See, e.g., Rhodes, C.A. (1995) Methods Mol. Biol. 55:121-131.)

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Although the presence/absence of marker gene expression suggests that the gene of interest is also present, the presence and expression of the gene may need to be confirmed. For example, if the sequence encoding SECP is inserted within a marker gene sequence, transformed cells containing sequences encoding SECP can be identified by the absence of marker gene function. Alternatively, a marker gene can be placed in tandem with a sequence encoding SECP under the control of a single promoter. Expression of the marker gene in response to induction or selection usually indicates expression of the tandem gene as well.

In general, host cells that contain the nucleic acid sequence encoding SECP and that express SECP may be identified by a variety of procedures known to those of skill in the art. These procedures include, but are not limited to, DNA-DNA or DNA-RNA hybridizations, PCR amplification, and protein bioassay or immunoassay techniques which include membrane, solution, or chip based technologies for the detection and/or quantification of nucleic acid or protein sequences.

Immunological methods for detecting and measuring the expression of SECP using either specific polyclonal or monoclonal antibodies are known in the art. Examples of such techniques include enzyme-linked immunosorbent assays (ELISAs), radioimmunoassays (RIAs), and fluorescence activated cell sorting (FACS). A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering epitopes on SECP is preferred, but a competitive binding assay may be employed. These and other assays are well known in the art. (See, e.g., Hampton, R. et al. (1990) Serological Methods, a Laboratory Manual, APS Press, St. Paul MN, Sect. IV; Coligan, J.E. et al. (1997) Current Protocols in Immunology, Greene Pub. Associates and Wiley-Interscience, New York NY; and Pound, J.D. (1998) Immunochemical Protocols, Humana Press, Totowa NJ.)

A wide variety of labels and conjugation techniques are known by those skilled in the art and may be used in various nucleic acid and amino acid assays. Means for producing labeled

hybridization or PCR probes for detecting sequences related to polynucleotides encoding SECP include oligolabeling, nick translation, end-labeling, or PCR amplification using a labeled nucleotide. Alternatively, the sequences encoding SECP, or any fragments thereof, may be cloned into a vector for the production of an mRNA probe. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes in vitro by addition of an appropriate RNA polymerase such as T7, T3, or SP6 and labeled nucleotides. These procedures may be conducted using a variety of commercially available kits, such as those provided by Amersham Pharmacia Biotech, Promega (Madison WI), and US Biochemical. Suitable reporter molecules or labels which may be used for ease of detection include radionuclides, enzymes, fluorescent, chemiluminescent, or chromogenic agents, as well as substrates, cofactors, inhibitors, magnetic particles, and the like.

Host cells transformed with nucleotide sequences encoding SECP may be cultured under conditions suitable for the expression and recovery of the protein from cell culture. The protein produced by a transformed cell may be secreted or retained intracellularly depending on the sequence and/or the vector used. As will be understood by those of skill in the art, expression vectors containing polynucleotides which encode SECP may be designed to contain signal sequences which direct secretion of SECP through a prokaryotic or eukaryotic cell membrane.

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In addition, a host cell strain may be chosen for its ability to modulate expression of the inserted sequences or to process the expressed protein in the desired fashion. Such modifications of the polypeptide include, but are not limited to, acetylation, carboxylation, glycosylation, phosphorylation, lipidation, and acylation. Post-translational processing which cleaves a "prepro" or "pro" form of the protein may also be used to specify protein targeting, folding, and/or activity. Different host cells which have specific cellular machinery and characteristic mechanisms for post-translational activities (e.g., CHO, HeLa, MDCK, HEK293, and WI38) are available from the American Type Culture Collection (ATCC, Manassas VA) and may be chosen to ensure the correct modification and processing of the foreign protein.

In another embodiment of the invention, natural, modified, or recombinant nucleic acid sequences encoding SECP may be ligated to a heterologous sequence resulting in translation of a fusion protein in any of the aforementioned host systems. For example, a chimeric SECP protein containing a heterologous moiety that can be recognized by a commercially available antibody may facilitate the screening of peptide libraries for inhibitors of SECP activity. Heterologous protein and peptide moieties may also facilitate purification of fusion proteins using commercially available affinity matrices. Such moieties include, but are not limited to, glutathione S-transferase (GST), maltose binding protein (MBP), thioredoxin (Trx), calmodulin binding peptide (CBP), 6-His, FLAG, *c-myc*, and hemagglutinin (HA). GST, MBP, Trx, CBP, and 6-His enable purification of their cognate fusion proteins on immobilized glutathione, maltose, phenylarsine oxide, calmodulin, and

metal-chelate resins, respectively. FLAG, c-myc, and hemagglutinin (HA) enable immunoaffinity purification of fusion proteins using commercially available monoclonal and polyclonal antibodies that specifically recognize these epitope tags. A fusion protein may also be engineered to contain a proteolytic cleavage site located between the SECP encoding sequence and the heterologous protein sequence, so that SECP may be cleaved away from the heterologous moiety following purification. Methods for fusion protein expression and purification are discussed in Ausubel (1995, supra, ch. 10). A variety of commercially available kits may also be used to facilitate expression and purification of fusion proteins.

In a further embodiment of the invention, synthesis of radiolabeled SECP may be achieved <u>in</u> <u>vitro</u> using the TNT rabbit reticulocyte lysate or wheat germ extract system (Promega). These systems couple transcription and translation of protein-coding sequences operably associated with the T7, T3, or SP6 promoters. Translation takes place in the presence of a radiolabeled amino acid precursor, for example, ³⁵S-methionine.

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SECP of the present invention or fragments thereof may be used to screen for compounds that specifically bind to SECP. At least one and up to a plurality of test compounds may be screened for specific binding to SECP. Examples of test compounds include antibodies, oligonucleotides, proteins (e.g., receptors), or small molecules.

In one embodiment, the compound thus identified is closely related to the natural ligand of SECP, e.g., a ligand or fragment thereof, a natural substrate, a structural or functional mimetic, or a natural binding partner. (See, e.g., Coligan, J.E. et al. (1991) <u>Current Protocols in Immunology</u> 1(2): Chapter 5.) Similarly, the compound can be closely related to the natural receptor to which SECP binds, or to at least a fragment of the receptor, e.g., the ligand binding site. In either case, the compound can be rationally designed using known techniques. In one embodiment, screening for these compounds involves producing appropriate cells which express SECP, either as a secreted protein or on the cell membrane. Preferred cells include cells from mammals, yeast, <u>Drosophila</u>, or <u>E. coli</u>. Cells expressing SECP or cell membrane fractions which contain SECP are then contacted with a test compound and binding, stimulation, or inhibition of activity of either SECP or the compound is analyzed.

An assay may simply test binding of a test compound to the polypeptide, wherein binding is detected by a fluorophore, radioisotope, enzyme conjugate, or other detectable label. For example, the assay may comprise the steps of combining at least one test compound with SECP, either in solution or affixed to a solid support, and detecting the binding of SECP to the compound. Alternatively, the assay may detect or measure binding of a test compound in the presence of a labeled competitor. Additionally, the assay may be carried out using cell-free preparations, chemical libraries, or natural product mixtures, and the test compound(s) may be free in solution or affixed to a

solid support.

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SECP of the present invention or fragments thereof may be used to screen for compounds that modulate the activity of SECP. Such compounds may include agonists, antagonists, or partial or inverse agonists. In one embodiment, an assay is performed under conditions permissive for SECP activity, wherein SECP is combined with at least one test compound, and the activity of SECP in the presence of a test compound is compared with the activity of SECP in the absence of the test compound. A change in the activity of SECP in the presence of the test compound is indicative of a compound that modulates the activity of SECP. Alternatively, a test compound is combined with an in vitro or cell-free system comprising SECP under conditions suitable for SECP activity, and the assay is performed. In either of these assays, a test compound which modulates the activity of SECP may do so indirectly and need not come in direct contact with the test compound. At least one and up to a plurality of test compounds may be screened.

In another embodiment, polynucleotides encoding SECP or their mammalian homologs may be "knocked out" in an animal model system using homologous recombination in embryonic stem (ES) cells. Such techniques are well known in the art and are useful for the generation of animal models of human disease. (See, e.g., U.S. Patent No. 5,175,383 and U.S. Patent No. 5,767,337.) For example, mouse ES cells, such as the mouse 129/SvJ cell line, are derived from the early mouse embryo and grown in culture. The ES cells are transformed with a vector containing the gene of interest disrupted by a marker gene, e.g., the neomycin phosphotransferase gene (neo; Capecchi, M.R. (1989) Science 244:1288-1292). The vector integrates into the corresponding region of the host genome by homologous recombination. Alternatively, homologous recombination takes place using the Cre-loxP system to knockout a gene of interest in a tissue- or developmental stage-specific manner (Marth, J.D. (1996) Clin. Invest. 97:1999-2002; Wagner, K.U. et al. (1997) Nucleic Acids Res. 25:4323-4330). Transformed ES cells are identified and microinjected into mouse cell . blastocysts such as those from the C57BL/6 mouse strain. The blastocysts are surgically transferred to pseudopregnant dams, and the resulting chimeric progeny are genotyped and bred to produce heterozygous or homozygous strains. Transgenic animals thus generated may be tested with potential therapeutic or toxic agents.

Polynucleotides encoding SECP may also be manipulated <u>in vitro</u> in ES cells derived from human blastocysts. Human ES cells have the potential to differentiate into at least eight separate cell lineages including endoderm, mesoderm, and ectodermal cell types. These cell lineages differentiate into, for example, neural cells, hematopoietic lineages, and cardiomyocytes (Thomson, J.A. et al. (1998) Science 282:1145-1147).

Polynucleotides encoding SECP can also be used to create "knockin" humanized animals (pigs) or transgenic animals (mice or rats) to model human disease. With knockin technology, a

region of a polynucleotide encoding SECP is injected into animal ES cells, and the injected sequence integrates into the animal cell genome. Transformed cells are injected into blastulae, and the blastulae are implanted as described above. Transgenic progeny or inbred lines are studied and treated with potential pharmaceutical agents to obtain information on treatment of a human disease. Alternatively, a mammal inbred to overexpress SECP, e.g., by secreting SECP in its milk, may also serve as a convenient source of that protein (Janne, J. et al. (1998) Biotechnol. Annu. Rev. 4:55-74). THERAPEUTICS

Chemical and structural similarity, e.g., in the context of sequences and motifs, exists between regions of SECP and secreted proteins. In addition, the expression of SECP is closely associated with breast, reproductive, digestive, urinary, fibroblastic, diseased, tumorous, testicular, pituitary, adenoid, lymph node, monocyte, ileum, coronary artery endothelium, uterine endometrial and brain tissues. Examples can also be found in Table 6. Therefore, SECP appears to play a role in cell proliferative, autoimmune/inflammatory, cardiovascular, neurological, and developmental disorders. In the treatment of disorders associated with increased SECP expression or activity, it is desirable to decrease the expression or activity of SECP. In the treatment of disorders associated with decreased SECP expression or activity of SECP.

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Therefore, in one embodiment, SECP or a fragment or derivative thereof may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of SECP. Examples of such disorders include, but are not limited to, a cell proliferative disorder such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease (MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia, and cancers including adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, a cancer of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus; an autoimmune/inflammatory disorder such as acquired immunodeficiency syndrome (AIDS), Addison's disease, adult respiratory distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, autoimmune polyendocrinopathycandidiasis-ectodermal dystrophy (APECED), bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, episodic lymphopenia with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis, glomerulonephritis, Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple sclerosis, myasthenia gravis, myocardial or

pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, psoriasis, Reiter's syndrome, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome, complications of cancer, hemodialysis, and extracorporeal circulation, viral, bacterial, fungal, parasitic, protozoal, and helminthic infections, and trauma; a cardiovascular disorder such as congestive heart failure, ischemic heart disease, angina pectoris, myocardial infarction, hypertensive heart disease, degenerative valvular heart disease, calcific aortic valve stenosis, congenitally bicuspid aortic valve, mitral annular calcification, mitral valve prolapse, rheumatic fever and rheumatic heart disease, infective endocarditis, nonbacterial thrombotic endocarditis, endocarditis of systemic lupus erythematosus, carcinoid heart disease, cardiomyopathy, myocarditis, pericarditis, neoplastic heart disease, congenital heart disease, complications of cardiac transplantation, arteriovenous fistula, atherosclerosis, hypertension, vasculitis, Raynaud's disease, aneurysms, arterial dissections, varicose veins, thrombophlebitis and phlebothrombosis, vascular tumors, and complications of thrombolysis, balloon angioplasty, vascular replacement, and coronary artery bypass graft surgery; a neurological disorder such as epilepsy, ischemic cerebrovascular disease, stroke, cerebral neoplasms, Alzheimer's disease, Pick's disease, Huntington's disease, dementia, Parkinson's disease and other extrapyramidal disorders, amyotrophic lateral sclerosis and other motor neuron disorders, progressive neural muscular atrophy, retinitis pigmentosa, hereditary ataxias, multiple sclerosis and other demyelinating diseases, bacterial and viral meningitis, brain abscess, subdural empyema, epidural abscess, suppurative intracranial thrombophlebitis, myelitis and radiculitis, viral central nervous system disease, prion diseases including kuru, Creutzfeldt-Jakob disease, and Gerstmann-Straussler-Scheinker syndrome, fatal familial insomnia, nutritional and metabolic diseases of the nervous system, neurofibromatosis, tuberous sclerosis, cerebelloretinal hemangioblastomatosis. encephalotrigeminal syndrome, mental retardation and other developmental disorders of the central nervous system including Down syndrome, cerebral palsy, neuroskeletal disorders, autonomic nervous system disorders, cranial nerve disorders, spinal cord diseases, muscular dystrophy and other neuromuscular disorders, peripheral nervous system disorders, dermatomyositis and polymyositis. inherited, metabolic, endocrine, and toxic myopathies, myasthenia gravis, periodic paralysis, mental disorders including mood, anxiety, and schizophrenic disorders, seasonal affective disorder (SAD), akathesia, amnesia, catatonia, diabetic neuropathy, tardive dyskinesia, dystonias, paranoid psychoses. postherpetic neuralgia, Tourette's disorder, progressive supranuclear palsy, corticobasal degeneration, and familial frontotemporal dementia; and a developmental disorder such as renal tubular acidosis, anemia, Cushing's syndrome, achondroplastic dwarfism, Duchenne and Becker muscular dystrophy. epilepsy, gonadal dysgenesis, WAGR syndrome (Wilms' tumor, aniridia, genitourinary abnormalities, and mental retardation), Smith-Magenis syndrome, myelodysplastic syndrome, hereditary

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mucoepithelial dysplasia, hereditary keratodermas, hereditary neuropathies such as Charcot-Marie-Tooth disease and neurofibromatosis, hypothyroidism, hydrocephalus, seizure disorders such as Syndenham's chorea and cerebral palsy, spina bifida, anencephaly, craniorachischisis, congenital glaucoma, cataract, and sensorineural hearing loss.

In another embodiment, a vector capable of expressing SECP or a fragment or derivative thereof may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of SECP including, but not limited to, those described above.

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In a further embodiment, a composition comprising a substantially purified SECP in conjunction with a suitable pharmaceutical carrier may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of SECP including, but not limited to, those provided above.

In still another embodiment, an agonist which modulates the activity of SECP may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of SECP including, but not limited to, those listed above.

In a further embodiment, an antagonist of SECP may be administered to a subject to treat or prevent a disorder associated with increased expression or activity of SECP. Examples of such disorders include, but are not limited to, those cell proliferative, autoimmune/inflammatory, cardiovascular, neurological, and developmental disorders described above. In one aspect, an antibody which specifically binds SECP may be used directly as an antagonist or indirectly as a targeting or delivery mechanism for bringing a pharmaceutical agent to cells or tissues which express SECP.

In an additional embodiment, a vector expressing the complement of the polynucleotide encoding SECP may be administered to a subject to treat or prevent a disorder associated with increased expression or activity of SECP including, but not limited to, those described above.

In other embodiments, any of the proteins, antagonists, antibodies, agonists, complementary sequences, or vectors of the invention may be administered in combination with other appropriate therapeutic agents. Selection of the appropriate agents for use in combination therapy may be made by one of ordinary skill in the art, according to conventional pharmaceutical principles. The combination of therapeutic agents may act synergistically to effect the treatment or prevention of the various disorders described above. Using this approach, one may be able to achieve therapeutic efficacy with lower dosages of each agent, thus reducing the potential for adverse side effects.

An antagonist of SECP may be produced using methods which are generally known in the art. In particular, purified SECP may be used to produce antibodies or to screen libraries of pharmaceutical agents to identify those which specifically bind SECP. Antibodies to SECP may also be generated using methods that are well known in the art. Such antibodies may include, but are not

limited to, polyclonal, monoclonal, chimeric, and single chain antibodies, Fab fragments, and fragments produced by a Fab expression library. Neutralizing antibodies (i.e., those which inhibit dimer formation) are generally preferred for therapeutic use.

For the production of antibodies, various hosts including goats, rabbits, rats, mice, humans, and others may be immunized by injection with SECP or with any fragment or oligopeptide thereof which has immunogenic properties. Depending on the host species, various adjuvants may be used to increase immunological response. Such adjuvants include, but are not limited to, Freund's, mineral gels such as aluminum hydroxide, and surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, KLH, and dinitrophenol. Among adjuvants used in humans, BCG (bacilli Calmette-Guerin) and Corynebacterium parvum are especially preferable.

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It is preferred that the oligopeptides, peptides, or fragments used to induce antibodies to SECP have an amino acid sequence consisting of at least about 5 amino acids, and generally will consist of at least about 10 amino acids. It is also preferable that these oligopeptides, peptides, or fragments are identical to a portion of the amino acid sequence of the natural protein. Short stretches of SECP amino acids may be fused with those of another protein, such as KLH, and antibodies to the chimeric molecule may be produced.

Monoclonal antibodies to SECP may be prepared using any technique which provides for the production of antibody molecules by continuous cell lines in culture. These include, but are not limited to, the hybridoma technique, the human B-cell hybridoma technique, and the EBV-hybridoma technique. (See, e.g., Kohler, G. et al. (1975) Nature 256:495-497; Kozbor, D. et al. (1985) J. Immunol. Methods 81:31-42; Cote, R.J. et al. (1983) Proc. Natl. Acad. Sci. USA 80:2026-2030; and Cole, S.P. et al. (1984) Mol. Cell Biol. 62:109-120.)

In addition, techniques developed for the production of "chimeric antibodies," such as the splicing of mouse antibody genes to human antibody genes to obtain a molecule with appropriate antigen specificity and biological activity, can be used. (See, e.g., Morrison, S.L. et al. (1984) Proc. Natl. Acad. Sci. USA 81:6851-6855; Neuberger, M.S. et al. (1984) Nature 312:604-608; and Takeda, S. et al. (1985) Nature 314:452-454.) Alternatively, techniques described for the production of single chain antibodies may be adapted, using methods known in the art, to produce SECP-specific single chain antibodies. Antibodies with related specificity, but of distinct idiotypic composition, may be generated by chain shuffling from random combinatorial immunoglobulin libraries. (See, e.g., Burton, D.R. (1991) Proc. Natl. Acad. Sci. USA 88:10134-10137.)

Antibodies may also be produced by inducing <u>in vivo</u> production in the lymphocyte population or by screening immunoglobulin libraries or panels of highly specific binding reagents as disclosed in the literature. (See, e.g., Orlandi, R. et al. (1989) Proc. Natl. Acad. Sci. USA 86:3833-3837; Winter, G. et al. (1991) Nature 349:293-299.)

Antibody fragments which contain specific binding sites for SECP may also be generated. For example, such fragments include, but are not limited to, $F(ab)_2$ fragments produced by pepsin digestion of the antibody molecule and Fab fragments generated by reducing the disulfide bridges of the $F(ab)_2$ fragments. Alternatively, Fab expression libraries may be constructed to allow rapid and easy identification of monoclonal Fab fragments with the desired specificity. (See, e.g., Huse, W.D. et al. (1989) Science 246:1275-1281.)

Various immunoassays may be used for screening to identify antibodies having the desired specificity. Numerous protocols for competitive binding or immunoradiometric assays using either polyclonal or monoclonal antibodies with established specificities are well known in the art. Such immunoassays typically involve the measurement of complex formation between SECP and its specific antibody. A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering SECP epitopes is generally used, but a competitive binding assay may also be employed (Pound, supra).

Various methods such as Scatchard analysis in conjunction with radioimmunoassay techniques may be used to assess the affinity of antibodies for SECP. Affinity is expressed as an association constant, K_a , which is defined as the molar concentration of SECP-antibody complex divided by the molar concentrations of free antigen and free antibody under equilibrium conditions. The K_a determined for a preparation of polyclonal antibodies, which are heterogeneous in their affinities for multiple SECP epitopes, represents the average affinity, or avidity, of the antibodies for SECP. The K_a determined for a preparation of monoclonal antibodies, which are monospecific for a particular SECP epitope, represents a true measure of affinity. High-affinity antibody preparations with K_a ranging from about 10^9 to 10^{12} L/mole are preferred for use in immunoassays in which the SECP-antibody complex must withstand rigorous manipulations. Low-affinity antibody preparations with K_a ranging from about 10^6 to 10^7 L/mole are preferred for use in immunopurification and similar procedures which ultimately require dissociation of SECP, preferably in active form, from the antibody (Catty, D. (1988) Antibodies, Volume I: A Practical Approach, IRL Press, Washington DC; Liddell, J.E. and A. Cryer (1991) A Practical Guide to Monoclonal Antibodies, John Wiley & Sons, New York NY).

The titer and avidity of polyclonal antibody preparations may be further evaluated to determine the quality and suitability of such preparations for certain downstream applications. For example, a polyclonal antibody preparation containing at least 1-2 mg specific antibody/ml, preferably 5-10 mg specific antibody/ml, is generally employed in procedures requiring precipitation of SECP-antibody complexes. Procedures for evaluating antibody specificity, titer, and avidity, and guidelines for antibody quality and usage in various applications, are generally available. (See, e.g., Catty, supra, and Coligan et al. supra.)

In another embodiment of the invention, the polynucleotides encoding SECP, or any fragment or complement thereof, may be used for therapeutic purposes. In one aspect, modifications of gene expression can be achieved by designing complementary sequences or antisense molecules (DNA, RNA, PNA, or modified oligonucleotides) to the coding or regulatory regions of the gene encoding SECP. Such technology is well known in the art, and antisense oligonucleotides or larger fragments can be designed from various locations along the coding or control regions of sequences encoding SECP. (See, e.g., Agrawal, S., ed. (1996) Antisense Therapeutics, Humana Press Inc., Totawa NJ.)

In therapeutic use, any gene delivery system suitable for introduction of the antisense sequences into appropriate target cells can be used. Antisense sequences can be delivered intracellularly in the form of an expression plasmid which, upon transcription, produces a sequence complementary to at least a portion of the cellular sequence encoding the target protein. (See, e.g., Slater, J.E. et al. (1998) J. Allergy Clin. Immunol. 102(3):469-475; and Scanlon, K.J. et al. (1995) 9(13):1288-1296.) Antisense sequences can also be introduced intracellularly through the use of viral vectors, such as retrovirus and adeno-associated virus vectors. (See, e.g., Miller, A.D. (1990) Blood 76:271; Ausubel, supra; Uckert, W. and W. Walther (1994) Pharmacol. Ther. 63(3):323-347.) Other gene delivery mechanisms include liposome-derived systems, artificial viral envelopes, and other systems known in the art. (See, e.g., Rossi, J.J. (1995) Br. Med. Bull. 51(1):217-225; Boado, R.J. et al. (1998) J. Pharm. Sci. 87(11):1308-1315; and Morris, M.C. et al. (1997) Nucleic Acids Res. 25(14):2730-2736.)

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In another embodiment of the invention, polynucleotides encoding SECP may be used for somatic or germline gene therapy. Gene therapy may be performed to (i) correct a genetic deficiency (e.g., in the cases of severe combined immunodeficiency (SCID)-X1 disease characterized by Xlinked inheritance (Cavazzana-Calvo, M. et al. (2000) Science 288:669-672), severe combined immunodeficiency syndrome associated with an inherited adenosine deaminase (ADA) deficiency (Blaese, R.M. et al. (1995) Science 270:475-480; Bordignon, C. et al. (1995) Science 270:470-475), cystic fibrosis (Zabner, J. et al. (1993) Cell 75:207-216; Crystal, R.G. et al. (1995) Hum. Gene Therapy 6:643-666; Crystal, R.G. et al. (1995) Hum. Gene Therapy 6:667-703), thalassamias, familial hypercholesterolemia, and hemophilia resulting from Factor VIII or Factor IX deficiencies (Crystal, R.G. (1995) Science 270:404-410; Verma, I.M. and N. Somia (1997) Nature 389:239-242)), (ii) express a conditionally lethal gene product (e.g., in the case of cancers which result from unregulated cell proliferation), or (iii) express a protein which affords protection against intracellular parasites (e.g., against human retroviruses, such as human immunodeficiency virus (HIV) (Baltimore, D. (1988) Nature 335:395-396; Poeschla, E. et al. (1996) Proc. Natl. Acad. Sci. USA 93:11395-11399). hepatitis B or C virus (HBV, HCV); fungal parasites, such as Candida albicans and Paracoccidioides brasiliensis; and protozoan parasites such as Plasmodium falciparum and Trypanosoma cruzi). In the

case where a genetic deficiency in SECP expression or regulation causes disease, the expression of SECP from an appropriate population of transduced cells may alleviate the clinical manifestations caused by the genetic deficiency.

In a further embodiment of the invention, diseases or disorders caused by deficiencies in SECP are treated by constructing mammalian expression vectors encoding SECP and introducing these vectors by mechanical means into SECP-deficient cells. Mechanical transfer technologies for use with cells in vivo or ex vitro include (i) direct DNA microinjection into individual cells, (ii) ballistic gold particle delivery, (iii) liposome-mediated transfection, (iv) receptor-mediated gene transfer, and (v) the use of DNA transposons (Morgan, R.A. and W.F. Anderson (1993) Annu. Rev. Biochem. 62:191-217; Ivics, Z. (1997) Cell 91:501-510; Boulay, J-L. and H. Récipon (1998) Curr. Opin. Biotechnol. 9:445-450).

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Expression vectors that may be effective for the expression of SECP include, but are not limited to, the PCDNA 3.1, EPITAG, PRCCMV2, PREP, PVAX, PCR2-TOPOTA vectors (Invitrogen, Carlsbad CA), PCMV-SCRIPT, PCMV-TAG, PEGSH/PERV (Stratagene, La Jolla CA), and PTET-OFF, PTET-ON, PTRE2, PTRE2-LUC, PTK-HYG (Clontech, Palo Alto CA). SECP may be expressed using (i) a constitutively active promoter, (e.g., from cytomegalovirus (CMV), Rous sarcoma virus (RSV), SV40 virus, thymidine kinase (TK), or β-actin genes), (ii) an inducible promoter (e.g., the tetracycline-regulated promoter (Gossen, M. and H. Bujard (1992) Proc. Natl. Acad. Sci. USA 89:5547-5551; Gossen, M. et al. (1995) Science 268:1766-1769; Rossi, F.M.V. and H.M. Blau (1998) Curr. Opin. Biotechnol. 9:451-456), commercially available in the T-REX plasmid (Invitrogen)); the ecdysone-inducible promoter (available in the plasmids PVGRXR and PIND; Invitrogen); the FK506/rapamycin inducible promoter; or the RU486/mifepristone inducible promoter (Rossi, F.M.V. and H.M. Blau, supra)), or (iii) a tissue-specific promoter or the native promoter of the endogenous gene encoding SECP from a normal individual.

Commercially available liposome transformation kits (e.g., the PERFECT LIPID TRANSFECTION KIT, available from Invitrogen) allow one with ordinary skill in the art to deliver polynucleotides to target cells in culture and require minimal effort to optimize experimental parameters. In the alternative, transformation is performed using the calcium phosphate method (Graham, F.L. and A.J. Eb (1973) Virology 52:456-467), or by electroporation (Neumann, E. et al. (1982) EMBO J. 1:841-845). The introduction of DNA to primary cells requires modification of these standardized mammalian transfection protocols.

In another embodiment of the invention, diseases or disorders caused by genetic defects with respect to SECP expression are treated by constructing a retrovirus vector consisting of (i) the polynucleotide encoding SECP under the control of an independent promoter or the retrovirus long terminal repeat (LTR) promoter, (ii) appropriate RNA packaging signals, and (iii) a Rev-responsive

element (RRE) along with additional retrovirus cis-acting RNA sequences and coding sequences required for efficient vector propagation. Retrovirus vectors (e.g., PFB and PFBNEO) are commercially available (Stratagene) and are based on published data (Riviere, I. et al. (1995) Proc. Natl. Acad. Sci. USA 92:6733-6737), incorporated by reference herein. The vector is propagated in an appropriate vector producing cell line (VPCL) that expresses an envelope gene with a tropism for receptors on the target cells or a promiscuous envelope protein such as VSVg (Armentano, D. et al. (1987) J. Virol. 61:1647-1650; Bender, M.A. et al. (1987) J. Virol. 61:1639-1646; Adam, M.A. and A.D. Miller (1988) J. Virol. 62:3802-3806; Dull, T. et al. (1998) J. Virol. 72:8463-8471; Zufferey, R. et al. (1998) J. Virol. 72:9873-9880). U.S. Patent No. 5,910,434 to Rigg ("Method for obtaining retrovirus packaging cell lines producing high transducing efficiency retroviral supernatant") discloses a method for obtaining retrovirus packaging cell lines and is hereby incorporated by reference. Propagation of retrovirus vectors, transduction of a population of cells (e.g., CD4+ Tcells), and the return of transduced cells to a patient are procedures well known to persons skilled in the art of gene therapy and have been well documented (Ranga, U. et al. (1997) J. Virol. 71:7020-7029; Bauer, G. et al. (1997) Blood 89:2259-2267; Bonyhadi, M.L. (1997) J. Virol. 71:4707-4716; 15 Ranga, U. et al. (1998) Proc. Natl. Acad. Sci. USA 95:1201-1206; Su, L. (1997) Blood 89:2283-2290).

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In the alternative, an adenovirus-based gene therapy delivery system is used to deliver polynucleotides encoding SECP to cells which have one or more genetic abnormalities with respect to the expression of SECP. The construction and packaging of adenovirus-based vectors are well known to those with ordinary skill in the art. Replication defective adenovirus vectors have proven to be versatile for importing genes encoding immunoregulatory proteins into intact islets in the pancreas (Csete, M.E. et al. (1995) Transplantation 27:263-268). Potentially useful adenoviral vectors are described in U.S. Patent No. 5,707,618 to Armentano ("Adenovirus vectors for gene therapy"), hereby incorporated by reference. For adenoviral vectors, see also Antinozzi, P.A. et al. (1999) Annu. Rev. Nutr. 19:511-544 and Verma, I.M. and N. Somia (1997) Nature 18:389:239-242, both incorporated by reference herein.

In another alternative, a herpes-based, gene therapy delivery system is used to deliver polynucleotides encoding SECP to target cells which have one or more genetic abnormalities with respect to the expression of SECP. The use of herpes simplex virus (HSV)-based vectors may be especially valuable for introducing SECP to cells of the central nervous system, for which HSV has a tropism. The construction and packaging of herpes-based vectors are well known to those with ordinary skill in the art. A replication-competent herpes simplex virus (HSV) type 1-based vector has been used to deliver a reporter gene to the eyes of primates (Liu, X. et al. (1999) Exp. Eye Res. 169:385-395). The construction of a HSV-1 virus vector has also been disclosed in detail in U.S.

Patent No. 5,804,413 to DeLuca ("Herpes simplex virus strains for gene transfer"), which is hereby incorporated by reference. U.S. Patent No. 5,804,413 teaches the use of recombinant HSV d92 which consists of a genome containing at least one exogenous gene to be transferred to a cell under the control of the appropriate promoter for purposes including human gene therapy. Also taught by this patent are the construction and use of recombinant HSV strains deleted for ICP4, ICP27 and ICP22. For HSV vectors, see also Goins, W.F. et al. (1999) J. Virol. 73:519-532 and Xu, H. et al. (1994) Dev. Biol. 163:152-161, hereby incorporated by reference. The manipulation of cloned herpesvirus sequences, the generation of recombinant virus following the transfection of multiple plasmids containing different segments of the large herpesvirus genomes, the growth and propagation of herpesvirus, and the infection of cells with herpesvirus are techniques well known to those of ordinary skill in the art.

In another alternative, an alphavirus (positive, single-stranded RNA virus) vector is used to deliver polynucleotides encoding SECP to target cells. The biology of the prototypic alphavirus, Semliki Forest Virus (SFV), has been studied extensively and gene transfer vectors have been based on the SFV genome (Garoff, H. and K.-J. Li (1998) Curr. Opin. Biotechnol. 9:464-469). During alphavirus RNA replication, a subgenomic RNA is generated that normally encodes the viral capsid proteins. This subgenomic RNA replicates to higher levels than the full length genomic RNA. resulting in the overproduction of capsid proteins relative to the viral proteins with enzymatic activity (e.g., protease and polymerase). Similarly, inserting the coding sequence for SECP into the alphavirus genome in place of the capsid-coding region results in the production of a large number of SECP-coding RNAs and the synthesis of high levels of SECP in vector transduced cells. While alphavirus infection is typically associated with cell lysis within a few days, the ability to establish a persistent infection in hamster normal kidney cells (BHK-21) with a variant of Sindbis virus (SIN) indicates that the lytic replication of alphaviruses can be altered to suit the needs of the gene therapy application (Dryga, S.A. et al. (1997) Virology 228:74-83). The wide host range of alphaviruses will allow the introduction of SECP into a variety of cell types. The specific transduction of a subset of cells in a population may require the sorting of cells prior to transduction. The methods of manipulating infectious cDNA clones of alphaviruses, performing alphavirus cDNA and RNA transfections, and performing alphavirus infections, are well known to those with ordinary skill in the art.

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Oligonucleotides derived from the transcription initiation site, e.g., between about positions -10 and +10 from the start site, may also be employed to inhibit gene expression. Similarly, inhibition can be achieved using triple helix base-pairing methodology. Triple helix pairing is useful because it causes inhibition of the ability of the double helix to open sufficiently for the binding of polymerases, transcription factors, or regulatory molecules. Recent therapeutic advances using

triplex DNA have been described in the literature. (See, e.g., Gee, J.E. et al. (1994) in Huber, B.E. and B.I. Carr, Molecular and Immunologic Approaches, Futura Publishing, Mt. Kisco NY, pp. 163-177.) A complementary sequence or antisense molecule may also be designed to block translation of mRNA by preventing the transcript from binding to ribosomes.

Ribozymes, enzymatic RNA molecules, may also be used to catalyze the specific cleavage of RNA. The mechanism of ribozyme action involves sequence-specific hybridization of the ribozyme molecule to complementary target RNA, followed by endonucleolytic cleavage. For example, engineered hammerhead motif ribozyme molecules may specifically and efficiently catalyze endonucleolytic cleavage of sequences encoding SECP.

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Specific ribozyme cleavage sites within any potential RNA target are initially identified by scanning the target molecule for ribozyme cleavage sites, including the following sequences: GUA, GUU, and GUC. Once identified, short RNA sequences of between 15 and 20 ribonucleotides, corresponding to the region of the target gene containing the cleavage site, may be evaluated for secondary structural features which may render the oligonucleotide inoperable. The suitability of candidate targets may also be evaluated by testing accessibility to hybridization with complementary oligonucleotides using ribonuclease protection assays.

Complementary ribonucleic acid molecules and ribozymes of the invention may be prepared by any method known in the art for the synthesis of nucleic acid molecules. These include techniques for chemically synthesizing oligonucleotides such as solid phase phosphoramidite chemical synthesis. Alternatively, RNA molecules may be generated by in vitro and in vivo transcription of DNA sequences encoding SECP. Such DNA sequences may be incorporated into a wide variety of vectors with suitable RNA polymerase promoters such as T7 or SP6. Alternatively, these cDNA constructs that synthesize complementary RNA, constitutively or inducibly, can be introduced into cell lines, cells, or tissues.

RNA molecules may be modified to increase intracellular stability and half-life. Possible modifications include, but are not limited to, the addition of flanking sequences at the 5' and/or 3' ends of the molecule, or the use of phosphorothioate or 2' O-methyl rather than phosphodiesterase linkages within the backbone of the molecule. This concept is inherent in the production of PNAs and can be extended in all of these molecules by the inclusion of nontraditional bases such as inosine, queosine, and wybutosine, as well as acetyl-, methyl-, thio-, and similarly modified forms of adenine, cytidine, guanine, thymine, and uridine which are not as easily recognized by endogenous endonucleases.

An additional embodiment of the invention encompasses a method for screening for a compound which is effective in altering expression of a polynucleotide encoding SECP. Compounds which may be effective in altering expression of a specific polynucleotide may include, but are not

limited to, oligonucleotides, antisense oligonucleotides, triple helix-forming oligonucleotides, transcription factors and other polypeptide transcriptional regulators, and non-macromolecular chemical entities which are capable of interacting with specific polynucleotide sequences. Effective compounds may alter polynucleotide expression by acting as either inhibitors or promoters of polynucleotide expression. Thus, in the treatment of disorders associated with increased SECP expression or activity, a compound which specifically inhibits expression of the polynucleotide encoding SECP may be therapeutically useful, and in the treatment of disorders associated with decreased SECP expression or activity, a compound which specifically promotes expression of the polynucleotide encoding SECP may be therapeutically useful.

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At least one, and up to a plurality, of test compounds may be screened for effectiveness in altering expression of a specific polynucleotide. A test compound may be obtained by any method commonly known in the art, including chemical modification of a compound known to be effective in altering polynucleotide expression; selection from an existing, commercially-available or proprietary library of naturally-occurring or non-natural chemical compounds; rational design of a compound based on chemical and/or structural properties of the target polynucleotide; and selection from a library of chemical compounds created combinatorially or randomly. A sample comprising a polynucleotide encoding SECP is exposed to at least one test compound thus obtained. The sample may comprise, for example, an intact or permeabilized cell, or an in vitro cell-free or reconstituted biochemical system. Alterations in the expression of a polynucleotide encoding SECP are assayed by any method commonly known in the art. Typically, the expression of a specific nucleotide is detected by hybridization with a probe having a nucleotide sequence complementary to the sequence of the polynucleotide encoding SECP. The amount of hybridization may be quantified, thus forming the basis for a comparison of the expression of the polynucleotide both with and without exposure to one or more test compounds. Detection of a change in the expression of a polynucleotide exposed to a test compound indicates that the test compound is effective in altering the expression of the polynucleotide. A screen for a compound effective in altering expression of a specific polynucleotide can be carried out, for example, using a Schizosaccharomyces pombe gene expression system (Atkins, D. et al. (1999) U.S. Patent No. 5,932,435; Arndt, G.M. et al. (2000) Nucleic Acids Res. 28:E15) or a human cell line such as HeLa cell (Clarke, M.L. et al. (2000) Biochem. Biophys. Res. Commun. 268:8-13). A particular embodiment of the present invention involves screening a combinatorial library of oligonucleotides (such as deoxyribonucleotides, ribonucleotides, peptide nucleic acids, and modified oligonucleotides) for antisense activity against a specific polynucleotide sequence (Bruice, T.W. et al. (1997) U.S. Patent No. 5,686,242; Bruice, T.W. et al. (2000) U.S. Patent No. 6,022,691).

Many methods for introducing vectors into cells or tissues are available and equally suitable

for use in vivo, in vitro, and ex vivo. For ex vivo therapy, vectors may be introduced into stem cells taken from the patient and clonally propagated for autologous transplant back into that same patient. Delivery by transfection, by liposome injections, or by polycationic amino polymers may be achieved using methods which are well known in the art. (See, e.g., Goldman, C.K. et al. (1997) Nat. Biotechnol. 15:462-466.)

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Any of the therapeutic methods described above may be applied to any subject in need of such therapy, including, for example, mammals such as humans, dogs, cats, cows, horses, rabbits, and monkeys.

An additional embodiment of the invention relates to the administration of a composition which generally comprises an active ingredient formulated with a pharmaceutically acceptable excipient. Excipients may include, for example, sugars, starches, celluloses, gums, and proteins. Various formulations are commonly known and are thoroughly discussed in the latest edition of Remington's Pharmaceutical Sciences (Maack Publishing, Easton PA). Such compositions may consist of SECP, antibodies to SECP, and mimetics, agonists, antagonists, or inhibitors of SECP.

The compositions utilized in this invention may be administered by any number of routes including, but not limited to, oral, intravenous, intramuscular, intra-arterial, intramedullary, intrathecal, intraventricular, pulmonary, transdermal, subcutaneous, intraperitoneal, intranasal, enteral, topical, sublingual, or rectal means.

Compositions for pulmonary administration may be prepared in liquid or dry powder form. These compositions are generally aerosolized immediately prior to inhalation by the patient. In the case of small molecules (e.g. traditional low molecular weight organic drugs), aerosol delivery of fast-acting formulations is well-known in the art. In the case of macromolecules (e.g. larger peptides and proteins), recent developments in the field of pulmonary delivery via the alveolar region of the lung have enabled the practical delivery of drugs such as insulin to blood circulation (see, e.g., Patton, J.S. et al., U.S. Patent No. 5,997,848). Pulmonary delivery has the advantage of administration without needle injection, and obviates the need for potentially toxic penetration enhancers.

Compositions suitable for use in the invention include compositions wherein the active ingredients are contained in an effective amount to achieve the intended purpose. The determination of an effective dose is well within the capability of those skilled in the art.

Specialized forms of compositions may be prepared for direct intracellular delivery of macromolecules comprising SECP or fragments thereof. For example, liposome preparations containing a cell-impermeable macromolecule may promote cell fusion and intracellular delivery of the macromolecule. Alternatively, SECP or a fragment thereof may be joined to a short cationic N-terminal portion from the HIV Tat-1 protein. Fusion proteins thus generated have been found to transduce into the cells of all tissues, including the brain, in a mouse model system (Schwarze, S.R. et

al. (1999) Science 285:1569-1572).

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For any compound, the therapeutically effective dose can be estimated initially either in cell culture assays, e.g., of neoplastic cells, or in animal models such as mice, rats, rabbits, dogs, monkeys, or pigs. An animal model may also be used to determine the appropriate concentration range and route of administration. Such information can then be used to determine useful doses and routes for administration in humans.

A therapeutically effective dose refers to that amount of active ingredient, for example SECP or fragments thereof, antibodies of SECP, and agonists, antagonists or inhibitors of SECP, which ameliorates the symptoms or condition. Therapeutic efficacy and toxicity may be determined by standard pharmaceutical procedures in cell cultures or with experimental animals, such as by calculating the ED₅₀ (the dose therapeutically effective in 50% of the population) or LD₅₀ (the dose lethal to 50% of the population) statistics. The dose ratio of toxic to therapeutic effects is the therapeutic index, which can be expressed as the LD₅₀/ED₅₀ ratio. Compositions which exhibit large therapeutic indices are preferred. The data obtained from cell culture assays and animal studies are used to formulate a range of dosage for human use. The dosage contained in such compositions is preferably within a range of circulating concentrations that includes the ED₅₀ with little or no toxicity. The dosage varies within this range depending upon the dosage form employed, the sensitivity of the patient, and the route of administration.

The exact dosage will be determined by the practitioner, in light of factors related to the subject requiring treatment. Dosage and administration are adjusted to provide sufficient levels of the active moiety or to maintain the desired effect. Factors which may be taken into account include the severity of the disease state, the general health of the subject, the age, weight, and gender of the subject, time and frequency of administration, drug combination(s), reaction sensitivities, and response to therapy. Long-acting compositions may be administered every 3 to 4 days, every week, or biweekly depending on the half-life and clearance rate of the particular formulation.

Normal dosage amounts may vary from about 0.1 μ g to 100,000 μ g, up to a total dose of about 1 gram, depending upon the route of administration. Guidance as to particular dosages and methods of delivery is provided in the literature and generally available to practitioners in the art. Those skilled in the art will employ different formulations for nucleotides than for proteins or their inhibitors. Similarly, delivery of polynucleotides or polypeptides will be specific to particular cells, conditions, locations, etc.

DIAGNOSTICS

In another embodiment, antibodies which specifically bind SECP may be used for the diagnosis of disorders characterized by expression of SECP, or in assays to monitor patients being treated with SECP or agonists, antagonists, or inhibitors of SECP. Antibodies useful for diagnostic

purposes may be prepared in the same manner as described above for therapeutics. Diagnostic assays for SECP include methods which utilize the antibody and a label to detect SECP in human body fluids or in extracts of cells or tissues. The antibodies may be used with or without modification, and may be labeled by covalent or non-covalent attachment of a reporter molecule. A wide variety of reporter molecules, several of which are described above, are known in the art and may be used.

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A variety of protocols for measuring SECP, including ELISAs, RIAs, and FACS, are known in the art and provide a basis for diagnosing altered or abnormal levels of SECP expression. Normal or standard values for SECP expression are established by combining body fluids or cell extracts taken from normal mammalian subjects, for example, human subjects, with antibodies to SECP under conditions suitable for complex formation. The amount of standard complex formation may be quantitated by various methods, such as photometric means. Quantities of SECP expressed in subject, control, and disease samples from biopsied tissues are compared with the standard values. Deviation between standard and subject values establishes the parameters for diagnosing disease.

In another embodiment of the invention, the polynucleotides encoding SECP may be used for diagnostic purposes. The polynucleotides which may be used include oligonucleotide sequences, complementary RNA and DNA molecules, and PNAs. The polynucleotides may be used to detect and quantify gene expression in biopsied tissues in which expression of SECP may be correlated with disease. The diagnostic assay may be used to determine absence, presence, and excess expression of SECP, and to monitor regulation of SECP levels during therapeutic intervention.

In one aspect, hybridization with PCR probes which are capable of detecting polynucleotide sequences, including genomic sequences, encoding SECP or closely related molecules may be used to identify nucleic acid sequences which encode SECP. The specificity of the probe, whether it is made from a highly specific region, e.g., the 5' regulatory region, or from a less specific region, e.g., a conserved motif, and the stringency of the hybridization or amplification will determine whether the probe identifies only naturally occurring sequences encoding SECP, allelic variants, or related sequences.

Probes may also be used for the detection of related sequences, and may have at least 50% sequence identity to any of the SECP encoding sequences. The hybridization probes of the subject invention may be DNA or RNA and may be derived from the sequence of SEQ ID NO:55-108 or from genomic sequences including promoters, enhancers, and introns of the SECP gene.

Means for producing specific hybridization probes for DNAs encoding SECP include the cloning of polynucleotide sequences encoding SECP or SECP derivatives into vectors for the production of mRNA probes. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes in vitro by means of the addition of the appropriate RNA polymerases and the appropriate labeled nucleotides. Hybridization probes may be labeled by a

variety of reporter groups, for example, by radionuclides such as ³²P or ³⁵S, or by enzymatic labels, such as alkaline phosphatase coupled to the probe via avidin/biotin coupling systems, and the like.

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Polynucleotide sequences encoding SECP may be used for the diagnosis of disorders associated with expression of SECP. Examples of such disorders include, but are not limited to, a cell proliferative disorder such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease (MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia, and cancers including adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, a cancer of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus; an autoimmune/inflammatory disorder such as acquired immunodeficiency syndrome (AIDS), Addison's disease, adult respiratory distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, autoimmune polyendocrinopathy-candidiasis-ectodermal dystrophy (APECED), bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, episodic lymphopenia with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis, glomerulonephritis, Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, psoriasis, Reiter's syndrome, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome, complications of cancer, hemodialysis, and extracorporeal circulation, viral, bacterial, fungal, parasitic, protozoal, and helminthic infections, and trauma; a cardiovascular disorder such as congestive heart failure, ischemic heart disease, angina pectoris, myocardial infarction, hypertensive heart disease, degenerative valvular heart disease, calcific aortic valve stenosis, congenitally bicuspid aortic valve, mitral annular calcification, mitral valve prolapse, rheumatic fever and rheumatic heart disease, infective endocarditis, nonbacterial thrombotic endocarditis, endocarditis of systemic lupus erythematosus, carcinoid heart disease, cardiomyopathy, myocarditis, pericarditis, neoplastic heart disease, congenital heart disease, complications of cardiac transplantation, arteriovenous fistula, atherosclerosis, hypertension, vasculitis, Raynaud's disease, aneurysms, arterial dissections, varicose veins, thrombophlebitis and phlebothrombosis, vascular tumors, and complications of thrombolysis, balloon angioplasty, vascular replacement, and coronary artery bypass graft surgery; a neurological disorder such as epilepsy, ischemic cerebrovascular disease, stroke, cerebral neoplasms, Alzheimer's disease, Pick's disease, Huntington's disease,

dementia, Parkinson's disease and other extrapyramidal disorders, amyotrophic lateral sclerosis and other motor neuron disorders, progressive neural muscular atrophy, retinitis pigmentosa, hereditary ataxias, multiple sclerosis and other demyelinating diseases, bacterial and viral meningitis, brain abscess, subdural empyema, epidural abscess, suppurative intracranial thrombophlebitis, myelitis and radiculitis, viral central nervous system disease, prion diseases including kuru, Creutzfeldt-Jakob disease, and Gerstmann-Straussler-Scheinker syndrome, fatal familial insomnia, nutritional and metabolic diseases of the nervous system, neurofibromatosis, tuberous sclerosis, cerebelloretinal hemangioblastomatosis, encephalotrigeminal syndrome, mental retardation and other developmental disorders of the central nervous system including Down syndrome, cerebral palsy, neuroskeletal disorders, autonomic nervous system disorders, cranial nerve disorders, spinal cord diseases, muscular dystrophy and other neuromuscular disorders, peripheral nervous system disorders, dermatomyositis and polymyositis, inherited, metabolic, endocrine, and toxic myopathies, myasthenia gravis, periodic paralysis, mental disorders including mood, anxiety, and schizophrenic disorders, seasonal affective disorder (SAD), akathesia, amnesia, catatonia, diabetic neuropathy, tardive dyskinesia, dystonias, paranoid psychoses, postherpetic neuralgia, Tourette's disorder, progressive supranuclear palsy, corticobasal degeneration, and familial frontotemporal dementia; and a developmental disorder such as renal tubular acidosis, anemia, Cushing's syndrome, achondroplastic dwarfism, Duchenne and Becker muscular dystrophy, epilepsy, gonadal dysgenesis, WAGR syndrome (Wilms' tumor, aniridia, genitourinary abnormalities, and mental retardation), Smith-Magenis syndrome, myelodysplastic syndrome, hereditary mucoepithelial dysplasia, hereditary keratodermas, hereditary neuropathies such as Charcot-Marie-Tooth disease and neurofibromatosis. hypothyroidism, hydrocephalus, seizure disorders such as Syndenham's chorea and cerebral palsy, spina bifida, anencephaly, craniorachischisis, congenital glaucoma, cataract, and sensorineural hearing loss. The polynucleotide sequences encoding SECP may be used in Southern or northern analysis, dot blot, or other membrane-based technologies; in PCR technologies; in dipstick, pin, and multiformat ELISA-like assays; and in microarrays utilizing fluids or tissues from patients to detect altered SECP expression. Such qualitative or quantitative methods are well known in the art.

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In a particular aspect, the nucleotide sequences encoding SECP may be useful in assays that detect the presence of associated disorders, particularly those mentioned above. The nucleotide sequences encoding SECP may be labeled by standard methods and added to a fluid or tissue sample from a patient under conditions suitable for the formation of hybridization complexes. After a suitable incubation period, the sample is washed and the signal is quantified and compared with a standard value. If the amount of signal in the patient sample is significantly altered in comparison to a control sample then the presence of altered levels of nucleotide sequences encoding SECP in the sample indicates the presence of the associated disorder. Such assays may also be used to evaluate

the efficacy of a particular therapeutic treatment regimen in animal studies, in clinical trials, or to monitor the treatment of an individual patient.

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In order to provide a basis for the diagnosis of a disorder associated with expression of SECP, a normal or standard profile for expression is established. This may be accomplished by combining body fluids or cell extracts taken from normal subjects, either animal or human, with a sequence, or a fragment thereof, encoding SECP, under conditions suitable for hybridization or amplification. Standard hybridization may be quantified by comparing the values obtained from normal subjects with values from an experiment in which a known amount of a substantially purified polynucleotide is used. Standard values obtained in this manner may be compared with values obtained from samples from patients who are symptomatic for a disorder. Deviation from standard values is used to establish the presence of a disorder.

Once the presence of a disorder is established and a treatment protocol is initiated, hybridization assays may be repeated on a regular basis to determine if the level of expression in the patient begins to approximate that which is observed in the normal subject. The results obtained from successive assays may be used to show the efficacy of treatment over a period ranging from several days to months.

With respect to cancer, the presence of an abnormal amount of transcript (either under- or overexpressed) in biopsied tissue from an individual may indicate a predisposition for the development of the disease, or may provide a means for detecting the disease prior to the appearance of actual clinical symptoms. A more definitive diagnosis of this type may allow health professionals to employ preventative measures or aggressive treatment earlier thereby preventing the development or further progression of the cancer.

Additional diagnostic uses for oligonucleotides designed from the sequences encoding SECP may involve the use of PCR. These oligomers may be chemically synthesized, generated enzymatically, or produced in vitro. Oligomers will preferably contain a fragment of a polynucleotide encoding SECP, or a fragment of a polynucleotide complementary to the polynucleotide encoding SECP, and will be employed under optimized conditions for identification of a specific gene or condition. Oligomers may also be employed under less stringent conditions for detection or quantification of closely related DNA or RNA sequences.

In a particular aspect, oligonucleotide primers derived from the polynucleotide sequences encoding SECP may be used to detect single nucleotide polymorphisms (SNPs). SNPs are substitutions, insertions and deletions that are a frequent cause of inherited or acquired genetic disease in humans. Methods of SNP detection include, but are not limited to, single-stranded conformation polymorphism (SSCP) and fluorescent SSCP (fSSCP) methods. In SSCP, oligonucleotide primers derived from the polynucleotide sequences encoding SECP are used to

amplify DNA using the polymerase chain reaction (PCR). The DNA may be derived, for example, from diseased or normal tissue, biopsy samples, bodily fluids, and the like. SNPs in the DNA cause differences in the secondary and tertiary structures of PCR products in single-stranded form, and these differences are detectable using gel electrophoresis in non-denaturing gels. In fSCCP, the oligonucleotide primers are fluorescently labeled, which allows detection of the amplimers in high-throughput equipment such as DNA sequencing machines. Additionally, sequence database analysis methods, termed in silico SNP (isSNP), are capable of identifying polymorphisms by comparing the sequence of individual overlapping DNA fragments which assemble into a common consensus sequence. These computer-based methods filter out sequence variations due to laboratory preparation of DNA and sequencing errors using statistical models and automated analyses of DNA sequence chromatograms. In the alternative, SNPs may be detected and characterized by mass spectrometry using, for example, the high throughput MASSARRAY system (Sequenom, Inc., San Diego CA).

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Methods which may also be used to quantify the expression of SECP include radiolabeling or biotinylating nucleotides, coamplification of a control nucleic acid, and interpolating results from standard curves. (See, e.g., Melby, P.C. et al. (1993) J. Immunol. Methods 159:235-244; Duplaa, C. et al. (1993) Anal. Biochem. 212:229-236.) The speed of quantitation of multiple samples may be accelerated by running the assay in a high-throughput format where the oligomer or polynucleotide of interest is presented in various dilutions and a spectrophotometric or colorimetric response gives rapid quantitation.

In further embodiments, oligonucleotides or longer fragments derived from any of the polynucleotide sequences described herein may be used as elements on a microarray. The microarray can be used in transcript imaging techniques which monitor the relative expression levels of large numbers of genes simultaneously as described below. The microarray may also be used to identify genetic variants, mutations, and polymorphisms. This information may be used to determine gene function, to understand the genetic basis of a disorder, to diagnose a disorder, to monitor progression/regression of disease as a function of gene expression, and to develop and monitor the activities of therapeutic agents in the treatment of disease. In particular, this information may be used to develop a pharmacogenomic profile of a patient in order to select the most appropriate and effective treatment regimen for that patient. For example, therapeutic agents which are highly effective and display the fewest side effects may be selected for a patient based on his/her pharmacogenomic profile.

In another embodiment, SECP, fragments of SECP, or antibodies specific for SECP may be used as elements on a microarray. The microarray may be used to monitor or measure protein-protein interactions, drug-target interactions, and gene expression profiles, as described above.

A particular embodiment relates to the use of the polynucleotides of the present invention to

generate a transcript image of a tissue or cell type. A transcript image represents the global pattern of gene expression by a particular tissue or cell type. Global gene expression patterns are analyzed by quantifying the number of expressed genes and their relative abundance under given conditions and at a given time. (See Seilhamer et al., "Comparative Gene Transcript Analysis," U.S. Patent No. 5,840,484, expressly incorporated by reference herein.) Thus a transcript image may be generated by hybridizing the polynucleotides of the present invention or their complements to the totality of transcripts or reverse transcripts of a particular tissue or cell type. In one embodiment, the hybridization takes place in high-throughput format, wherein the polynucleotides of the present invention or their complements comprise a subset of a plurality of elements on a microarray. The resultant transcript image would provide a profile of gene activity.

Transcript images may be generated using transcripts isolated from tissues, cell lines, biopsies, or other biological samples. The transcript image may thus reflect gene expression in vivo, as in the case of a tissue or biopsy sample, or in vitro, as in the case of a cell line.

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Transcript images which profile the expression of the polynucleotides of the present invention may also be used in conjunction with in vitro model systems and preclinical evaluation of pharmaceuticals, as well as toxicological testing of industrial and naturally-occurring environmental compounds. All compounds induce characteristic gene expression patterns, frequently termed molecular fingerprints or toxicant signatures, which are indicative of mechanisms of action and toxicity (Nuwaysir, E.F. et al. (1999) Mol. Carcinog. 24:153-159; Steiner, S. and N.L. Anderson (2000) Toxicol. Lett. 112-113:467-471, expressly incorporated by reference herein). If a test compound has a signature similar to that of a compound with known toxicity, it is likely to share those toxic properties. These fingerprints or signatures are most useful and refined when they contain expression information from a large number of genes and gene families. Ideally, a genome-wide measurement of expression provides the highest quality signature. Even genes whose expression is not altered by any tested compounds are important as well, as the levels of expression of these genes are used to normalize the rest of the expression data. The normalization procedure is useful for comparison of expression data after treatment with different compounds. While the assignment of gene function to elements of a toxicant signature aids in interpretation of toxicity mechanisms. knowledge of gene function is not necessary for the statistical matching of signatures which leads to prediction of toxicity. (See, for example, Press Release 00-02 from the National Institute of Environmental Health Sciences, released February 29, 2000, available at http://www.niehs.nih.gov/oc/news/toxchip.htm.) Therefore, it is important and desirable in toxicological screening using toxicant signatures to include all expressed gene sequences.

In one embodiment, the toxicity of a test compound is assessed by treating a biological sample containing nucleic acids with the test compound. Nucleic acids that are expressed in the

treated biological sample are hybridized with one or more probes specific to the polynucleotides of the present invention, so that transcript levels corresponding to the polynucleotides of the present invention may be quantified. The transcript levels in the treated biological sample are compared with levels in an untreated biological sample. Differences in the transcript levels between the two samples are indicative of a toxic response caused by the test compound in the treated sample.

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Another particular embodiment relates to the use of the polypeptide sequences of the present invention to analyze the proteome of a tissue or cell type. The term proteome refers to the global pattern of protein expression in a particular tissue or cell type. Each protein component of a proteome can be subjected individually to further analysis. Proteome expression patterns, or profiles, are analyzed by quantifying the number of expressed proteins and their relative abundance under given conditions and at a given time. A profile of a cell's proteome may thus be generated by separating and analyzing the polypeptides of a particular tissue or cell type. In one embodiment, the separation is achieved using two-dimensional gel electrophoresis, in which proteins from a sample are separated by isoelectric focusing in the first dimension, and then according to molecular weight by sodium dodecyl sulfate slab gel electrophoresis in the second dimension (Steiner and Anderson, supra). The proteins are visualized in the gel as discrete and uniquely positioned spots, typically by staining the gel with an agent such as Coomassie Blue or silver or fluorescent stains. The optical density of each protein spot is generally proportional to the level of the protein in the sample. The optical densities of equivalently positioned protein spots from different samples, for example, from biological samples either treated or untreated with a test compound or therapeutic agent, are compared to identify any changes in protein spot density related to the treatment. The proteins in the spots are partially sequenced using, for example, standard methods employing chemical or enzymatic cleavage followed by mass spectrometry. The identity of the protein in a spot may be determined by comparing its partial sequence, preferably of at least 5 contiguous amino acid residues, to the polypeptide sequences of the present invention. In some cases, further sequence data may be obtained for definitive protein identification.

A proteomic profile may also be generated using antibodies specific for SECP to quantify the levels of SECP expression. In one embodiment, the antibodies are used as elements on a microarray, and protein expression levels are quantified by exposing the microarray to the sample and detecting the levels of protein bound to each array element (Lucking, A. et al. (1999) Anal. Biochem. 270:103-111; Mendoze, L.G. et al. (1999) Biotechniques 27:778-788). Detection may be performed by a variety of methods known in the art, for example, by reacting the proteins in the sample with a thiolor amino-reactive fluorescent compound and detecting the amount of fluorescence bound at each array element.

Toxicant signatures at the proteome level are also useful for toxicological screening, and

should be analyzed in parallel with toxicant signatures at the transcript level. There is a poor correlation between transcript and protein abundances for some proteins in some tissues (Anderson, N.L. and J. Seilhamer (1997) Electrophoresis 18:533-537), so proteome toxicant signatures may be useful in the analysis of compounds which do not significantly affect the transcript image, but which alter the proteomic profile. In addition, the analysis of transcripts in body fluids is difficult, due to rapid degradation of mRNA, so proteomic profiling may be more reliable and informative in such cases.

In another embodiment, the toxicity of a test compound is assessed by treating a biological sample containing proteins with the test compound. Proteins that are expressed in the treated biological sample are separated so that the amount of each protein can be quantified. The amount of each protein is compared to the amount of the corresponding protein in an untreated biological sample. A difference in the amount of protein between the two samples is indicative of a toxic response to the test compound in the treated sample. Individual proteins are identified by sequencing the amino acid residues of the individual proteins and comparing these partial sequences to the polypeptides of the present invention.

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In another embodiment, the toxicity of a test compound is assessed by treating a biological sample containing proteins with the test compound. Proteins from the biological sample are incubated with antibodies specific to the polypeptides of the present invention. The amount of protein recognized by the antibodies is quantified. The amount of protein in the treated biological sample is compared with the amount in an untreated biological sample. A difference in the amount of protein between the two samples is indicative of a toxic response to the test compound in the treated sample.

Microarrays may be prepared, used, and analyzed using methods known in the art. (See, e.g., Brennan, T.M. et al. (1995) U.S. Patent No. 5,474,796; Schena, M. et al. (1996) Proc. Natl. Acad. Sci. USA 93:10614-10619; Baldeschweiler et al. (1995) PCT application WO95/251116; Shalon, D. et al. (1995) PCT application WO95/35505; Heller, R.A. et al. (1997) Proc. Natl. Acad. Sci. USA 94:2150-2155; and Heller, M.J. et al. (1997) U.S. Patent No. 5,605,662.) Various types of microarrays are well known and thoroughly described in <u>DNA Microarrays: A Practical Approach</u>, M. Schena, ed. (1999) Oxford University Press, London, hereby expressly incorporated by reference.

In another embodiment of the invention, nucleic acid sequences encoding SECP may be used to generate hybridization probes useful in mapping the naturally occurring genomic sequence. Either coding or noncoding sequences may be used, and in some instances, noncoding sequences may be preferable over coding sequences. For example, conservation of a coding sequence among members of a multi-gene family may potentially cause undesired cross hybridization during chromosomal mapping. The sequences may be mapped to a particular chromosome, to a specific region of a

chromosome, or to artificial chromosome constructions, e.g., human artificial chromosomes (HACs), yeast artificial chromosomes (YACs), bacterial artificial chromosomes (BACs), bacterial P1 constructions, or single chromosome cDNA libraries. (See, e.g., Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355; Price, C.M. (1993) Blood Rev. 7:127-134; and Trask, B.J. (1991) Trends Genet. 7:149-154.) Once mapped, the nucleic acid sequences of the invention may be used to develop genetic linkage maps, for example, which correlate the inheritance of a disease state with the inheritance of a particular chromosome region or restriction fragment length polymorphism (RFLP). (See, for example, Lander, E.S. and D. Botstein (1986) Proc. Natl. Acad. Sci. USA 83:7353-7357.)

Fluorescent in situ hybridization (FISH) may be correlated with other physical and genetic map data. (See, e.g., Heinz-Ulrich, et al. (1995) in Meyers, supra, pp. 965-968.) Examples of genetic map data can be found in various scientific journals or at the Online Mendelian Inheritance in Man (OMIM) World Wide Web site. Correlation between the location of the gene encoding SECP on a physical map and a specific disorder, or a predisposition to a specific disorder, may help define the region of DNA associated with that disorder and thus may further positional cloning efforts.

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In situ hybridization of chromosomal preparations and physical mapping techniques, such as linkage analysis using established chromosomal markers, may be used for extending genetic maps. Often the placement of a gene on the chromosome of another mammalian species, such as mouse, may reveal associated markers even if the exact chromosomal locus is not known. This information is valuable to investigators searching for disease genes using positional cloning or other gene discovery techniques. Once the gene or genes responsible for a disease or syndrome have been crudely localized by genetic linkage to a particular genomic region, e.g., ataxia-telangiectasia to 11q22-23, any sequences mapping to that area may represent associated or regulatory genes for further investigation. (See, e.g., Gatti, R.A. et al. (1988) Nature 336:577-580.) The nucleotide sequence of the instant invention may also be used to detect differences in the chromosomal location due to translocation, inversion, etc., among normal, carrier, or affected individuals.

In another embodiment of the invention, SECP, its catalytic or immunogenic fragments, or oligopeptides thereof can be used for screening libraries of compounds in any of a variety of drug screening techniques. The fragment employed in such screening may be free in solution, affixed to a solid support, borne on a cell surface, or located intracellularly. The formation of binding complexes between SECP and the agent being tested may be measured.

Another technique for drug screening provides for high throughput screening of compounds having suitable binding affinity to the protein of interest. (See, e.g., Geysen, et al. (1984) PCT application WO84/03564.) In this method, large numbers of different small test compounds are synthesized on a solid substrate. The test compounds are reacted with SECP, or fragments thereof, and washed. Bound SECP is then detected by methods well known in the art. Purified SECP can

also be coated directly onto plates for use in the aforementioned drug screening techniques.

Alternatively, non-neutralizing antibodies can be used to capture the peptide and immobilize it on a solid support.

In another embodiment, one may use competitive drug screening assays in which neutralizing antibodies capable of binding SECP specifically compete with a test compound for binding SECP. In this manner, antibodies can be used to detect the presence of any peptide which shares one or more antigenic determinants with SECP.

In additional embodiments, the nucleotide sequences which encode SECP may be used in any molecular biology techniques that have yet to be developed, provided the new techniques rely on properties of nucleotide sequences that are currently known, including, but not limited to, such properties as the triplet genetic code and specific base pair interactions.

Without further elaboration, it is believed that one skilled in the art can, using the preceding description, utilize the present invention to its fullest extent. The following embodiments are, therefore, to be construed as merely illustrative, and not limitative of the remainder of the disclosure in any way whatsoever.

The disclosures of all patents, applications and publications, mentioned above and below, including U.S. Ser. No. 60/262,932, U.S. Ser. No. 60/265,926, U.S. Ser. No. 60/255,639, U.S. Ser. No. 60/257,852, U.S. Ser. No. 60/260,105, U.S. Ser. No. 60/263,090 and U.S. Ser. No. 60/263,096 are expressly incorporated by reference herein.

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EXAMPLES

I. Construction of cDNA Libraries

Incyte cDNAs were derived from cDNA libraries described in the LIFESEQ GOLD database (Incyte Genomics, Palo Alto CA). Some tissues were homogenized and lysed in guanidinium isothiocyanate, while others were homogenized and lysed in phenol or in a suitable mixture of denaturants, such as TRIZOL (Life Technologies), a monophasic solution of phenol and guanidine isothiocyanate. The resulting lysates were centrifuged over CsCl cushions or extracted with chloroform. RNA was precipitated from the lysates with either isopropanol or sodium acetate and ethanol, or by other routine methods.

Phenol extraction and precipitation of RNA were repeated as necessary to increase RNA purity. In some cases, RNA was treated with DNase. For most libraries, poly(A)+RNA was isolated using oligo d(T)-coupled paramagnetic particles (Promega), OLIGOTEX latex particles (QIAGEN, Chatsworth CA), or an OLIGOTEX mRNA purification kit (QIAGEN). Alternatively, RNA was isolated directly from tissue lysates using other RNA isolation kits, e.g., the POLY(A)PURE mRNA purification kit (Ambion, Austin TX).

In some cases, Stratagene was provided with RNA and constructed the corresponding cDNA libraries. Otherwise, cDNA was synthesized and cDNA libraries were constructed with the UNIZAP vector system (Stratagene) or SUPERSCRIPT plasmid system (Life Technologies), using the recommended procedures or similar methods known in the art. (See, e.g., Ausubel, 1997, supra, units 5.1-6.6.) Reverse transcription was initiated using oligo d(T) or random primers. Synthetic oligonucleotide adapters were ligated to double stranded cDNA, and the cDNA was digested with the appropriate restriction enzyme or enzymes. For most libraries, the cDNA was size-selected (300-1000 bp) using SEPHACRYL S1000, SEPHAROSE CL2B, or SEPHAROSE CL4B column chromatography (Amersham Pharmacia Biotech) or preparative agarose gel electrophoresis. cDNAs were ligated into compatible restriction enzyme sites of the polylinker of a suitable plasmid, e.g., PBLUESCRIPT plasmid (Stratagene), PSPORT1 plasmid (Life Technologies), PCDNA2.1 plasmid (Invitrogen, Carlsbad CA), PBK-CMV plasmid (Stratagene), PCR2-TOPOTA plasmid (Invitrogen), PCMV-ICIS plasmid (Stratagene), pIGEN (Incyte Genomics, Palo Alto CA), pRARE (Incyte Genomics), or pINCY (Incyte Genomics), or derivatives thereof. Recombinant plasmids were transformed into competent E. coli cells including XL1-Blue, XL1-BlueMRF, or SOLR from Stratagene or DH5a, DH10B, or ElectroMAX DH10B from Life Technologies.

II. Isolation of cDNA Clones

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Plasmids obtained as described in Example I were recovered from host cells by <u>in vivo</u> excision using the UNIZAP vector system (Stratagene) or by cell lysis. Plasmids were purified using at least one of the following: a Magic or WIZARD Minipreps DNA purification system (Promega); an AGTC Miniprep purification kit (Edge Biosystems, Gaithersburg MD); and QIAWELL 8 Plasmid, QIAWELL 8 Plasmid, QIAWELL 8 Ultra Plasmid purification systems or the R.E.A.L. PREP 96 plasmid purification kit from QIAGEN. Following precipitation, plasmids were resuspended in 0.1 ml of distilled water and stored, with or without lyophilization, at 4°C.

Alternatively, plasmid DNA was amplified from host cell lysates using direct link PCR in a high-throughput format (Rao, V.B. (1994) Anal. Biochem. 216:1-14). Host cell lysis and thermal cycling steps were carried out in a single reaction mixture. Samples were processed and stored in 384-well plates, and the concentration of amplified plasmid DNA was quantified fluorometrically using PICOGREEN dye (Molecular Probes, Eugene OR) and a FLUOROSKAN II fluorescence scanner (Labsystems Oy, Helsinki, Finland).

III. Sequencing and Analysis

Incyte cDNA recovered in plasmids as described in Example II were sequenced as follows. Sequencing reactions were processed using standard methods or high-throughput instrumentation such as the ABI CATALYST 800 (Applied Biosystems) thermal cycler or the PTC-200 thermal cycler (MJ Research) in conjunction with the HYDRA microdispenser (Robbins Scientific) or the

MICROLAB 2200 (Hamilton) liquid transfer system. cDNA sequencing reactions were prepared using reagents provided by Amersham Pharmacia Biotech or supplied in ABI sequencing kits such as the ABI PRISM BIGDYE Terminator cycle sequencing ready reaction kit (Applied Biosystems). Electrophoretic separation of cDNA sequencing reactions and detection of labeled polynucleotides were carried out using the MEGABACE 1000 DNA sequencing system (Molecular Dynamics); the ABI PRISM 373 or 377 sequencing system (Applied Biosystems) in conjunction with standard ABI protocols and base calling software; or other sequence analysis systems known in the art. Reading frames within the cDNA sequences were identified using standard methods (reviewed in Ausubel, 1997, supra, unit 7.7). Some of the cDNA sequences were selected for extension using the techniques disclosed in Example VIII.

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The polynucleotide sequences derived from Incyte cDNAs were validated by removing vector, linker, and poly(A) sequences and by masking ambiguous bases, using algorithms and programs based on BLAST, dynamic programming, and dinucleotide nearest neighbor analysis. The Incyte cDNA sequences or translations thereof were then queried against a selection of public databases such as the GenBank primate, rodent, mammalian, vertebrate, and eukaryote databases, and BLOCKS, PRINTS, DOMO, PRODOM; PROTEOME databases with sequences from Homo sapiens, Rattus norvegicus, Mus musculus, Caenorhabditis elegans, Saccharomyces cerevisiae, Schizosaccharomyces pombe, and Candida albicans (Incyte Genomics, Palo Alto CA); and hidden Markov model (HMM)-based protein family databases such as PFAM. (HMM is a probabilistic approach which analyzes consensus primary structures of gene families. See, for example, Eddy, S.R. (1996) Curr. Opin. Struct. Biol. 6:361-365.) The queries were performed using programs based on BLAST, FASTA, BLIMPS, and HMMER. The Incyte cDNA sequences were assembled to produce full length polynucleotide sequences. Alternatively, GenBank cDNAs, GenBank ESTs, stitched sequences, stretched sequences, or Genscan-predicted coding sequences (see Examples IV and V) were used to extend Incyte cDNA assemblages to full length. Assembly was performed using programs based on Phred, Phrap, and Consed, and cDNA assemblages were screened for open reading frames using programs based on GeneMark, BLAST, and FASTA. The full length polynucleotide sequences were translated to derive the corresponding full length polypeptide sequences. Alternatively, a polypeptide of the invention may begin at any of the methionine residues of the full length translated polypeptide. Full length polypeptide sequences were subsequently analyzed by querying against databases such as the GenBank protein databases (genpept), SwissProt, the PROTEOME databases, BLOCKS, PRINTS, DOMO, PRODOM, Prosite, and hidden Markov model (HMM)-based protein family databases such as PFAM. Full length polynucleotide sequences are also analyzed using MACDNASIS PRO software (Hitachi Software Engineering, South San Francisco CA) and LASERGENE software (DNASTAR). Polynucleotide and polypeptide sequence

alignments are generated using default parameters specified by the CLUSTAL algorithm as incorporated into the MEGALIGN multisequence alignment program (DNASTAR), which also calculates the percent identity between aligned sequences.

Table 7 summarizes the tools, programs, and algorithms used for the analysis and assembly of Incyte cDNA and full length sequences and provides applicable descriptions, references, and threshold parameters. The first column of Table 7 shows the tools, programs, and algorithms used, the second column provides brief descriptions thereof, the third column presents appropriate references, all of which are incorporated by reference herein in their entirety, and the fourth column presents, where applicable, the scores, probability values, and other parameters used to evaluate the strength of a match between two sequences (the higher the score or the lower the probability value, the greater the identity between two sequences).

The programs described above for the assembly and analysis of full length polynucleotide and polypeptide sequences were also used to identify polynucleotide sequence fragments from SEQ ID NO:55-108. Fragments from about 20 to about 4000 nucleotides which are useful in hybridization and amplification technologies are described in Table 4, column 2.

IV. Identification and Editing of Coding Sequences from Genomic DNA

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Putative secreted proteins were initially identified by running the Genscan gene identification program against public genomic sequence databases (e.g., gbpri and gbhtg). Genscan is a generalpurpose gene identification program which analyzes genomic DNA sequences from a variety of organisms (See Burge, C. and S. Karlin (1997) J. Mol. Biol. 268:78-94, and Burge, C. and S. Karlin (1998) Curr. Opin. Struct. Biol. 8:346-354). The program concatenates predicted exons to form an assembled cDNA sequence extending from a methionine to a stop codon. The output of Genscan is a FASTA database of polynucleotide and polypeptide sequences. The maximum range of sequence for Genscan to analyze at once was set to 30 kb. To determine which of these Genscan predicted cDNA sequences encode secreted proteins, the encoded polypeptides were analyzed by querying against PFAM models for secreted proteins. Potential secreted proteins were also identified by homology to Incyte cDNA sequences that had been annotated as secreted proteins. These selected Genscanpredicted sequences were then compared by BLAST analysis to the genpept and gbpri public databases. Where necessary, the Genscan-predicted sequences were then edited by comparison to the top BLAST hit from genpept to correct errors in the sequence predicted by Genscan, such as extra or omitted exons. BLAST analysis was also used to find any Incyte cDNA or public cDNA coverage of the Genscan-predicted sequences, thus providing evidence for transcription. When Incyte cDNA coverage was available, this information was used to correct or confirm the Genscan predicted sequence. Full length polynucleotide sequences were obtained by assembling Genscan-predicted coding sequences with Incyte cDNA sequences and/or public cDNA sequences using the assembly

process described in Example III. Alternatively, full length polynucleotide sequences were derived entirely from edited or unedited Genscan-predicted coding sequences.

V. Assembly of Genomic Sequence Data with cDNA Sequence Data "Stitched" Sequences

Partial cDNA sequences were extended with exons predicted by the Genscan gene identification program described in Example IV. Partial cDNAs assembled as described in Example III were mapped to genomic DNA and parsed into clusters containing related cDNAs and Genscan exon predictions from one or more genomic sequences. Each cluster was analyzed using an algorithm based on graph theory and dynamic programming to integrate cDNA and genomic information, generating possible splice variants that were subsequently confirmed, edited, or extended to create a full length sequence. Sequence intervals in which the entire length of the interval was present on more than one sequence in the cluster were identified, and intervals thus identified were considered to be equivalent by transitivity. For example, if an interval was present on a cDNA and two genomic sequences, then all three intervals were considered to be equivalent. This process allows unrelated but consecutive genomic sequences to be brought together, bridged by cDNA sequence. Intervals thus identified were then "stitched" together by the stitching algorithm in the order that they appear along their parent sequences to generate the longest possible sequence, as well as sequence variants. Linkages between intervals which proceed along one type of parent sequence (cDNA to cDNA or genomic sequence to genomic sequence) were given preference over linkages which change parent type (cDNA to genomic sequence). The resultant stitched sequences were translated and compared by BLAST analysis to the genpept and gbpri public databases. Incorrect exons predicted by Genscan were corrected by comparison to the top BLAST hit from genpept. Sequences were further extended with additional cDNA sequences, or by inspection of genomic DNA, when necessary.

"Stretched" Sequences

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Partial DNA sequences were extended to full length with an algorithm based on BLAST analysis. First, partial cDNAs assembled as described in Example III were queried against public databases such as the GenBank primate, rodent, mammalian, vertebrate, and eukaryote databases using the BLAST program. The nearest GenBank protein homolog was then compared by BLAST analysis to either Incyte cDNA sequences or GenScan exon predicted sequences described in Example IV. A chimeric protein was generated by using the resultant high-scoring segment pairs (HSPs) to map the translated sequences onto the GenBank protein homolog. Insertions or deletions may occur in the chimeric protein with respect to the original GenBank protein homolog. The GenBank protein homolog, the chimeric protein, or both were used as probes to search for homologous genomic sequences from the public human genome databases. Partial DNA sequences were therefore "stretched" or extended by the addition of homologous genomic sequences. The

resultant stretched sequences were examined to determine whether it contained a complete gene.

VI. Chromosomal Mapping of SECP Encoding Polynucleotides

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The sequences which were used to assemble SEQ ID NO:55-108 were compared with sequences from the Incyte LIFESEQ database and public domain databases using BLAST and other implementations of the Smith-Waterman algorithm. Sequences from these databases that matched SEQ ID NO:55-108 were assembled into clusters of contiguous and overlapping sequences using assembly algorithms such as Phrap (Table 7). Radiation hybrid and genetic mapping data available from public resources such as the Stanford Human Genome Center (SHGC), Whitehead Institute for Genome Research (WIGR), and Généthon were used to determine if any of the clustered sequences had been previously mapped. Inclusion of a mapped sequence in a cluster resulted in the assignment of all sequences of that cluster, including its particular SEQ ID NO:, to that map location.

Map locations are represented by ranges, or intervals, of human chromosomes. The map position of an interval, in centiMorgans, is measured relative to the terminus of the chromosome's parm. (The centiMorgan (cM) is a unit of measurement based on recombination frequencies between chromosomal markers. On average, 1 cM is roughly equivalent to 1 megabase (Mb) of DNA in humans, although this can vary widely due to hot and cold spots of recombination.) The cM distances are based on genetic markers mapped by Généthon which provide boundaries for radiation hybrid markers whose sequences were included in each of the clusters. Human genome maps and other resources available to the public, such as the NCBI "GeneMap'99" World Wide Web site (http://www.ncbi.nlm.nih.gov/genemap/), can be employed to determine if previously identified disease genes map within or in proximity to the intervals indicated above.

In this manner, SEQ ID NO:58 was mapped to chromosome 3 within the interval from 160.0 to 187.1 centiMorgans. SEQ ID NO:59 was mapped to chromosome 15 within the interval from 59.3 centiMorgans to the q-terminus. SEQ ID NO:60 was mapped to chromosome 15 within the interval from 39.5 to 59.3 centiMorgans. SEQ ID NO:61 was mapped to chromosome 3 within the interval from 67.9 to 77.4 centiMorgans. SEQ ID NO:62 was mapped to chromosome 16 at 473.44 centiMorgans. SEQ ID NO:63 was mapped to chromosome 9 within the interval from 75.8 to 136.7 centiMorgans. SEQ ID NO:64 was mapped to chromosome 19. SEQ ID NO:65 was mapped to chromosome 1 within the interval from 196.5 to 205.1 centiMorgans. SEQ ID NO:66 was mapped to chromosome 5 within the interval from 138.7 to 141.4 centiMorgans. SEQ ID NO:67 was mapped to chromosome 2 within the interval from 223.1 to 231.8 centiMorgans. SEQ ID NO:68 was mapped to chromosome 2 within the interval from 223.1 to 231.8 centiMorgans. SEQ ID NO:69 was mapped to chromosome 17 within the interval from 62.2 centiMorgans to the q-terminus. SEQ ID NO:75 was mapped to chromosome 15 within the interval from 59.3 centiMorgans to the q terminus. SEQ ID NO:76 was mapped to chromosome 15 within the interval from 59.3 centiMorgans to the q-terminus SEQ ID NO:76 was mapped to chromosome 13 within the interval from the p-terminus to 36.6 centiMorgans.

SEQ ID NO:77 was mapped to the short arm of chromosome 8 within the cytogenetic band 23.3. SEQ ID NO:78 was mapped to chromosome 11 within the interval from 102.6 to 131.7 centiMorgans. SEQ ID NO:79 was mapped to chromosome 3 within the interval from 49.5 to 64.4 centiMorgans. SEQ ID NO:80 was mapped to chromosome 5 within the interval from 104.5 to 121.4 centiMorgans.

VII. Analysis of Polynucleotide Expression

Northern analysis is a laboratory technique used to detect the presence of a transcript of a gene and involves the hybridization of a labeled nucleotide sequence to a membrane on which RNAs from a particular cell type or tissue have been bound. (See, e.g., Sambrook, <u>supra</u>, ch. 7; Ausubel (1995) <u>supra</u>, ch. 4 and 16.)

Analogous computer techniques applying BLAST were used to search for identical or related molecules in cDNA databases such as GenBank or LIFESEQ (Incyte Genomics). This analysis is much faster than multiple membrane-based hybridizations. In addition, the sensitivity of the computer search can be modified to determine whether any particular match is categorized as exact or similar. The basis of the search is the product score, which is defined as:

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BLAST Score x Percent Identity 5 x minimum {length(Seq. 1), length(Seq. 2)}

The product score takes into account both the degree of similarity between two sequences and the length of the sequence match. The product score is a normalized value between 0 and 100, and is calculated as follows: the BLAST score is multiplied by the percent nucleotide identity and the product is divided by (5 times the length of the shorter of the two sequences). The BLAST score is calculated by assigning a score of +5 for every base that matches in a high-scoring segment pair (HSP), and -4 for every mismatch. Two sequences may share more than one HSP (separated by gaps). If there is more than one HSP, then the pair with the highest BLAST score is used to calculate the product score. The product score represents a balance between fractional overlap and quality in a BLAST alignment. For example, a product score of 100 is produced only for 100% identity over the entire length of the shorter of the two sequences being compared. A product score of 70 is produced either by 100% identity and 70% overlap at one end, or by 88% identity and 100% overlap at the other. A product score of 50 is produced either by 100% identity and 50% overlap at one end, or 79% identity and 100% overlap.

Alternatively, polynucleotide sequences encoding SECP are analyzed with respect to the tissue sources from which they were derived. For example, some full length sequences are assembled, at least in part, with overlapping Incyte cDNA sequences (see Example III). Each cDNA sequence is derived from a cDNA library constructed from a human tissue. Each human tissue is

classified into one of the following organ/tissue categories: cardiovascular system; connective tissue; digestive system; embryonic structures; endocrine system; exocrine glands; genitalia, female; genitalia, male; germ cells; hemic and immune system; liver; musculoskeletal system; nervous system; pancreas; respiratory system; sense organs; skin; stomatognathic system; unclassified/mixed; or urinary tract. The number of libraries in each category is counted and divided by the total number of libraries across all categories. Similarly, each human tissue is classified into one of the following disease/condition categories: cancer, cell line, developmental, inflammation, neurological, trauma, cardiovascular, pooled, and other, and the number of libraries in each category is counted and divided by the total number of libraries across all categories. The resulting percentages reflect the tissue- and disease-specific expression of cDNA encoding SECP. cDNA sequences and cDNA library/tissue information are found in the LIFESEQ GOLD database (Incyte Genomics, Palo Alto CA).

VIII. Extension of SECP Encoding Polynucleotides

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Full length polynucleotide sequences were also produced by extension of an appropriate fragment of the full length molecule using oligonucleotide primers designed from this fragment. One primer was synthesized to initiate 5' extension of the known fragment, and the other primer was synthesized to initiate 3' extension of the known fragment. The initial primers were designed using OLIGO 4.06 software (National Biosciences), or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the target sequence at temperatures of about 68°C to about 72°C. Any stretch of nucleotides which would result in hairpin structures and primer-primer dimerizations was avoided.

Selected human cDNA libraries were used to extend the sequence. If more than one extension was necessary or desired, additional or nested sets of primers were designed.

High fidelity amplification was obtained by PCR using methods well known in the art. PCR was performed in 96-well plates using the PTC-200 thermal cycler (MJ Research, Inc.). The reaction mix contained DNA template, 200 nmol of each primer, reaction buffer containing Mg²⁺, (NH₄)₂SO₄, and 2-mercaptoethanol, Taq DNA polymerase (Amersham Pharmacia Biotech), ELONGASE enzyme (Life Technologies), and Pfu DNA polymerase (Stratagene), with the following parameters for primer pair PCI A and PCI B: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 60°C, 1 min; Step 4: 68°C, 2 min; Step 5: Steps 2, 3, and 4 repeated 20 times; Step 6: 68°C, 5 min; Step 7: storage at 4°C. In the alternative, the parameters for primer pair T7 and SK+ were as follows: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 57°C, 1 min; Step 4: 68°C, 2 min; Step 5: Steps 2, 3, and 4 repeated 20 times; Step 6: 68°C, 5 min; Step 7: storage at 4°C.

The concentration of DNA in each well was determined by dispensing 100 μ l PICOGREEN quantitation reagent (0.25% (v/v) PICOGREEN; Molecular Probes, Eugene OR) dissolved in 1X TE and 0.5 μ l of undiluted PCR product into each well of an opaque fluorimeter plate (Corning Costar,

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Acton MA), allowing the DNA to bind to the reagent. The plate was scanned in a Fluoroskan II (Labsystems Oy, Helsinki, Finland) to measure the fluorescence of the sample and to quantify the concentration of DNA. A 5 μ l to 10 μ l aliquot of the reaction mixture was analyzed by electrophoresis on a 1 % agarose gel to determine which reactions were successful in extending the sequence.

The extended nucleotides were desalted and concentrated, transferred to 384-well plates, digested with CviJI cholera virus endonuclease (Molecular Biology Research, Madison WI), and sonicated or sheared prior to religation into pUC 18 vector (Amersham Pharmacia Biotech). For shotgun sequencing, the digested nucleotides were separated on low concentration (0.6 to 0.8%) 10 agarose gels, fragments were excised, and agar digested with Agar ACE (Promega). Extended clones were religated using T4 ligase (New England Biolabs, Beverly MA) into pUC 18 vector (Amersham Pharmacia Biotech), treated with Pfu DNA polymerase (Stratagene) to fill-in restriction site overhangs, and transfected into competent E. coli cells. Transformed cells were selected on antibiotic-containing media, and individual colonies were picked and cultured overnight at 37°C in 384-well plates in LB/2x carb liquid media.

The cells were lysed, and DNA was amplified by PCR using Taq DNA polymerase (Amersham Pharmacia Biotech) and Pfu DNA polymerase (Stratagene) with the following parameters: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 60°C, 1 min; Step 4: 72°C, 2 min; Step 5: steps 2, 3, and 4 repeated 29 times; Step 6: 72 °C, 5 min; Step 7: storage at 4 °C. DNA was quantified by PICOGREEN reagent (Molecular Probes) as described above. Samples with low DNA recoveries were reamplified using the same conditions as described above. Samples were diluted with 20% dimethysulfoxide (1:2, v/v), and sequenced using DYENAMIC energy transfer sequencing primers and the DYENAMIC DIRECT kit (Amersham Pharmacia Biotech) or the ABI PRISM BIGDYE Terminator cycle sequencing ready reaction kit (Applied Biosystems).

In like manner, full length polynucleotide sequences are verified using the above procedure or are used to obtain 5' regulatory sequences using the above procedure along with oligonucleotides designed for such extension, and an appropriate genomic library.

IX. Labeling and Use of Individual Hybridization Probes

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Hybridization probes derived from SEQ ID NO:55-108 are employed to screen cDNAs, genomic DNAs, or mRNAs. Although the labeling of oligonucleotides, consisting of about 20 base pairs, is specifically described, essentially the same procedure is used with larger nucleotide fragments. Oligonucleotides are designed using state-of-the-art software such as OLIGO 4.06 software (National Biosciences) and labeled by combining 50 pmol of each oligomer, 250 μ Ci of [y-32P] adenosine triphosphate (Amersham Pharmacia Biotech), and T4 polynucleotide kinase (DuPont NEN, Boston MA). The labeled oligonucleotides are substantially purified using a

SEPHADEX G-25 superfine size exclusion dextran bead column (Amersham Pharmacia Biotech). An aliquot containing 10⁷ counts per minute of the labeled probe is used in a typical membrane-based hybridization analysis of human genomic DNA digested with one of the following endonucleases: Ase I, Bgl II, Eco RI, Pst I, Xba I, or Pvu II (DuPont NEN).

The DNA from each digest is fractionated on a 0.7% agarose gel and transferred to nylon membranes (Nytran Plus, Schleicher & Schuell, Durham NH). Hybridization is carried out for 16 hours at 40°C. To remove nonspecific signals, blots are sequentially washed at room temperature under conditions of up to, for example, 0.1 x saline sodium citrate and 0.5% sodium dodecyl sulfate. Hybridization patterns are visualized using autoradiography or an alternative imaging means and compared.

X. Microarrays

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The linkage or synthesis of array elements upon a microarray can be achieved utilizing photolithography, piezoelectric printing (ink-jet printing, See, e.g., Baldeschweiler, supra.), mechanical microspotting technologies, and derivatives thereof. The substrate in each of the aforementioned technologies should be uniform and solid with a non-porous surface (Schena (1999), supra). Suggested substrates include silicon, silica, glass slides, glass chips, and silicon wafers. Alternatively, a procedure analogous to a dot or slot blot may also be used to arrange and link elements to the surface of a substrate using thermal, UV, chemical, or mechanical bonding procedures. A typical array may be produced using available methods and machines well known to those of ordinary skill in the art and may contain any appropriate number of elements. (See, e.g., Schena, M. et al. (1995) Science 270:467-470; Shalon, D. et al. (1996) Genome Res. 6:639-645; Marshall, A. and J. Hodgson (1998) Nat. Biotechnol. 16:27-31.)

Full length cDNAs, Expressed Sequence Tags (ESTs), or fragments or oligomers thereof may comprise the elements of the microarray. Fragments or oligomers suitable for hybridization can be 25 selected using software well known in the art such as LASERGENE software (DNASTAR). The array elements are hybridized with polynucleotides in a biological sample. The polynucleotides in the biological sample are conjugated to a fluorescent label or other molecular tag for ease of detection. After hybridization, nonhybridized nucleotides from the biological sample are removed, and a fluorescence scanner is used to detect hybridization at each array element. Alternatively, laser desorbtion and mass spectrometry may be used for detection of hybridization. The degree of complementarity and the relative abundance of each polynucleotide which hybridizes to an element on the microarray may be assessed. In one embodiment, microarray preparation and usage is described in detail below.

Tissue or Cell Sample Preparation

Total RNA is isolated from tissue samples using the guanidinium thiocyanate method and

poly(A)⁺ RNA is purified using the oligo-(dT) cellulose method. Each poly(A)⁺ RNA sample is reverse transcribed using MMLV reverse-transcriptase, 0.05 pg/μl oligo-(dT) primer (21mer), 1X first strand buffer, 0.03 units/μl RNase inhibitor, 500 μM dATP, 500 μM dGTP, 500 μM dTTP, 40 μM dCTP, 40 μM dCTP-Cy3 (BDS) or dCTP-Cy5 (Amersham Pharmacia Biotech). The reverse transcription reaction is performed in a 25 ml volume containing 200 ng poly(A)⁺ RNA with GEMBRIGHT kits (Incyte). Specific control poly(A)⁺ RNAs are synthesized by in vitro transcription from non-coding yeast genomic DNA. After incubation at 37°C for 2 hr, each reaction sample (one with Cy3 and another with Cy5 labeling) is treated with 2.5 ml of 0.5M sodium hydroxide and incubated for 20 minutes at 85°C to the stop the reaction and degrade the RNA. Samples are purified using two successive CHROMA SPIN 30 gel filtration spin columns (CLONTECH Laboratories, Inc. (CLONTECH), Palo Alto CA) and after combining, both reaction samples are ethanol precipitated using 1 ml of glycogen (1 mg/ml), 60 ml sodium acetate, and 300 ml of 100% ethanol. The sample is then dried to completion using a SpeedVAC (Savant Instruments Inc., Holbrook NY) and resuspended in 14 μl 5X SSC/0.2% SDS.

15 Microarray Preparation

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Sequences of the present invention are used to generate array elements. Each array element is amplified from bacterial cells containing vectors with cloned cDNA inserts. PCR amplification uses primers complementary to the vector sequences flanking the cDNA insert. Array elements are amplified in thirty cycles of PCR from an initial quantity of 1-2 ng to a final quantity greater than 5 μ g. Amplified array elements are then purified using SEPHACRYL-400 (Amersham Pharmacia Biotech).

Purified array elements are immobilized on polymer-coated glass slides. Glass microscope slides (Corning) are cleaned by ultrasound in 0.1% SDS and acetone, with extensive distilled water washes between and after treatments. Glass slides are etched in 4% hydrofluoric acid (VWR Scientific Products Corporation (VWR), West Chester PA), washed extensively in distilled water, and coated with 0.05% aminopropyl silane (Sigma) in 95% ethanol. Coated slides are cured in a 110°C oven.

Array elements are applied to the coated glass substrate using a procedure described in U.S. Patent No. 5,807,522, incorporated herein by reference. 1 μ l of the array element DNA, at an average concentration of 100 ng/ μ l, is loaded into the open capillary printing element by a high-speed robotic apparatus. The apparatus then deposits about 5 nl of array element sample per slide.

Microarrays are UV-crosslinked using a STRATALINKER UV-crosslinker (Stratagene). Microarrays are washed at room temperature once in 0.2% SDS and three times in distilled water. Non-specific binding sites are blocked by incubation of microarrays in 0.2% casein in phosphate buffered saline (PBS) (Tropix, Inc., Bedford MA) for 30 minutes at 60°C followed by washes in

0.2% SDS and distilled water as before.

Hybridization

Hybridization reactions contain 9 μ l of sample mixture consisting of 0.2 μ g each of Cy3 and Cy5 labeled cDNA synthesis products in 5X SSC, 0.2% SDS hybridization buffer. The sample mixture is heated to 65°C for 5 minutes and is aliquoted onto the microarray surface and covered with an 1.8 cm² coverslip. The arrays are transferred to a waterproof chamber having a cavity just slightly larger than a microscope slide. The chamber is kept at 100% humidity internally by the addition of 140 μ l of 5X SSC in a corner of the chamber. The chamber containing the arrays is incubated for about 6.5 hours at 60°C. The arrays are washed for 10 min at 45°C in a first wash buffer (1X SSC, 0.1% SDS), three times for 10 minutes each at 45°C in a second wash buffer (0.1X SSC), and dried.

Detection

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Reporter-labeled hybridization complexes are detected with a microscope equipped with an Innova 70 mixed gas 10 W laser (Coherent, Inc., Santa Clara CA) capable of generating spectral lines at 488 nm for excitation of Cy3 and at 632 nm for excitation of Cy5. The excitation laser light is focused on the array using a 20X microscope objective (Nikon, Inc., Melville NY). The slide containing the array is placed on a computer-controlled X-Y stage on the microscope and raster-scanned past the objective. The 1.8 cm x 1.8 cm array used in the present example is scanned with a resolution of 20 micrometers.

In two separate scans, a mixed gas multiline laser excites the two fluorophores sequentially. Emitted light is split, based on wavelength, into two photomultiplier tube detectors (PMT R1477, Hamamatsu Photonics Systems, Bridgewater NJ) corresponding to the two fluorophores. Appropriate filters positioned between the array and the photomultiplier tubes are used to filter the signals. The emission maxima of the fluorophores used are 565 nm for Cy3 and 650 nm for Cy5. Each array is typically scanned twice, one scan per fluorophore using the appropriate filters at the laser source, although the apparatus is capable of recording the spectra from both fluorophores simultaneously.

The sensitivity of the scans is typically calibrated using the signal intensity generated by a cDNA control species added to the sample mixture at a known concentration. A specific location on the array contains a complementary DNA sequence, allowing the intensity of the signal at that location to be correlated with a weight ratio of hybridizing species of 1:100,000. When two samples from different sources (e.g., representing test and control cells), each labeled with a different fluorophore, are hybridized to a single array for the purpose of identifying genes that are differentially expressed, the calibration is done by labeling samples of the calibrating cDNA with the two fluorophores and adding identical amounts of each to the hybridization mixture.

The output of the photomultiplier tube is digitized using a 12-bit RTI-835H analog-to-digital

(A/D) conversion board (Analog Devices, Inc., Norwood MA) installed in an IBM-compatible PC computer. The digitized data are displayed as an image where the signal intensity is mapped using a linear 20-color transformation to a pseudocolor scale ranging from blue (low signal) to red (high signal). The data is also analyzed quantitatively. Where two different fluorophores are excited and measured simultaneously, the data are first corrected for optical crosstalk (due to overlapping emission spectra) between the fluorophores using each fluorophore's emission spectrum.

A grid is superimposed over the fluorescence signal image such that the signal from each spot is centered in each element of the grid. The fluorescence signal within each element is then integrated to obtain a numerical value corresponding to the average intensity of the signal. The software used for signal analysis is the GEMTOOLS gene expression analysis program (Incyte).

XI. Complementary Polynucleotides

Sequences complementary to the SECP-encoding sequences, or any parts thereof, are used to detect, decrease, or inhibit expression of naturally occurring SECP. Although use of oligonucleotides comprising from about 15 to 30 base pairs is described, essentially the same procedure is used with smaller or with larger sequence fragments. Appropriate oligonucleotides are designed using OLIGO 4.06 software (National Biosciences) and the coding sequence of SECP. To inhibit transcription, a complementary oligonucleotide is designed from the most unique 5' sequence and used to prevent promoter binding to the coding sequence. To inhibit translation, a complementary oligonucleotide is designed to prevent ribosomal binding to the SECP-encoding transcript.

20 XII. Expression of SECP

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Expression and purification of SECP is achieved using bacterial or virus-based expression systems. For expression of SECP in bacteria, cDNA is subcloned into an appropriate vector containing an antibiotic resistance gene and an inducible promoter that directs high levels of cDNA transcription. Examples of such promoters include, but are not limited to, the trp-lac (tac) hybrid promoter and the T5 or T7 bacteriophage promoter in conjunction with the lac operator regulatory element. Recombinant vectors are transformed into suitable bacterial hosts, e.g., BL21(DE3). Antibiotic resistant bacteria express SECP upon induction with isopropyl beta-D-thiogalactopyranoside (IPTG). Expression of SECP in eukaryotic cells is achieved by infecting insect or mammalian cell lines with recombinant Autographica californica nuclear polyhedrosis virus (AcMNPV), commonly known as baculovirus. The nonessential polyhedrin gene of baculovirus is replaced with cDNA encoding SECP by either homologous recombination or bacterial-mediated transposition involving transfer plasmid intermediates. Viral infectivity is maintained and the strong polyhedrin promoter drives high levels of cDNA transcription. Recombinant baculovirus is used to infect Spodoptera frugiperda (Sf9) insect cells in most cases, or human hepatocytes, in some cases. Infection of the latter requires additional genetic modifications to baculovirus. (See Engelhard, E.K.

et al. (1994) Proc. Natl. Acad. Sci. USA 91:3224-3227; Sandig, V. et al. (1996) Hum. Gene Ther. 7:1937-1945.)

In most expression systems, SECP is synthesized as a fusion protein with, e.g., glutathione Stransferase (GST) or a peptide epitope tag, such as FLAG or 6-His, permitting rapid, single-step, affinity-based purification of recombinant fusion protein from crude cell lysates. GST, a 26-kilodalton enzyme from Schistosoma japonicum, enables the purification of fusion proteins on immobilized glutathione under conditions that maintain protein activity and antigenicity (Amersham Pharmacia Biotech). Following purification, the GST moiety can be proteolytically cleaved from SECP at specifically engineered sites. FLAG, an 8-amino acid peptide, enables immunoaffinity purification using commercially available monoclonal and polyclonal anti-FLAG antibodies (Eastman Kodak). 6-His, a stretch of six consecutive histidine residues, enables purification on metal-chelate resins (QIAGEN). Methods for protein expression and purification are discussed in Ausubel (1995, supra, ch. 10 and 16). Purified SECP obtained by these methods can be used directly in the assays shown in Examples XVI, XVII, and XVIII, where applicable.

15 XIII. Functional Assays

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SECP function is assessed by expressing the sequences encoding SECP at physiologically elevated levels in mammalian cell culture systems. cDNA is subcloned into a mammalian expression vector containing a strong promoter that drives high levels of cDNA expression. Vectors of choice include PCMV SPORT (Life Technologies) and PCR3.1 (Invitrogen, Carlsbad CA), both of which contain the cytomegalovirus promoter. 5-10 μ g of recombinant vector are transiently transfected into a human cell line, for example, an endothelial or hematopoietic cell line, using either liposome formulations or electroporation. 1-2 μ g of an additional plasmid containing sequences encoding a marker protein are co-transfected. Expression of a marker protein provides a means to distinguish transfected cells from nontransfected cells and is a reliable predictor of cDNA expression from the recombinant vector. Marker proteins of choice include, e.g., Green Fluorescent Protein (GFP; Clontech), CD64, or a CD64-GFP fusion protein. Flow cytometry (FCM), an automated, laser opticsbased technique, is used to identify transfected cells expressing GFP or CD64-GFP and to evaluate the apoptotic state of the cells and other cellular properties. FCM detects and quantifies the uptake of fluorescent molecules that diagnose events preceding or coincident with cell death. These events include changes in nuclear DNA content as measured by staining of DNA with propidium iodide; changes in cell size and granularity as measured by forward light scatter and 90 degree side light scatter; down-regulation of DNA synthesis as measured by decrease in bromodeoxyuridine uptake; alterations in expression of cell surface and intracellular proteins as measured by reactivity with specific antibodies; and alterations in plasma membrane composition as measured by the binding of fluorescein-conjugated Annexin V protein to the cell surface. Methods in flow cytometry are

discussed in Ormerod, M.G. (1994) Flow Cytometry, Oxford, New York NY.

The influence of SECP on gene expression can be assessed using highly purified populations of cells transfected with sequences encoding SECP and either CD64 or CD64-GFP. CD64 and CD64-GFP are expressed on the surface of transfected cells and bind to conserved regions of human immunoglobulin G (IgG). Transfected cells are efficiently separated from nontransfected cells using magnetic beads coated with either human IgG or antibody against CD64 (DYNAL, Lake Success NY). mRNA can be purified from the cells using methods well known by those of skill in the art. Expression of mRNA encoding SECP and other genes of interest can be analyzed by northern analysis or microarray techniques.

10 XIV. Production of SECP Specific Antibodies

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SECP substantially purified using polyacrylamide gel electrophoresis (PAGE; see, e.g., Harrington, M.G. (1990) Methods Enzymol. 182:488-495), or other purification techniques, is used to immunize rabbits and to produce antibodies using standard protocols.

Alternatively, the SECP amino acid sequence is analyzed using LASERGENE software (DNASTAR) to determine regions of high immunogenicity, and a corresponding oligopeptide is synthesized and used to raise antibodies by means known to those of skill in the art. Methods for selection of appropriate epitopes, such as those near the C-terminus or in hydrophilic regions are well described in the art. (See, e.g., Ausubel, 1995, supra, ch. 11.)

Typically, oligopeptides of about 15 residues in length are synthesized using an ABI 431A

20 peptide synthesizer (Applied Biosystems) using FMOC chemistry and coupled to KLH (Sigma-Aldrich, St. Louis MO) by reaction with N-maleimidobenzoyl-N-hydroxysuccinimide ester (MBS) to increase immunogenicity. (See, e.g., Ausubel, 1995, supra.) Rabbits are immunized with the oligopeptide-KLH complex in complete Freund's adjuvant. Resulting antisera are tested for antipeptide and anti-SECP activity by, for example, binding the peptide or SECP to a substrate, blocking with 1% BSA, reacting with rabbit antisera, washing, and reacting with radio-iodinated goat anti-rabbit IgG.

XV. Purification of Naturally Occurring SECP Using Specific Antibodies

Naturally occurring or recombinant SECP is substantially purified by immunoaffinity chromatography using antibodies specific for SECP. An immunoaffinity column is constructed by covalently coupling anti-SECP antibody to an activated chromatographic resin, such as CNBr-activated SEPHAROSE (Amersham Pharmacia Biotech). After the coupling, the resin is blocked and washed according to the manufacturer's instructions.

Media containing SECP are passed over the immunoaffinity column, and the column is washed under conditions that allow the preferential absorbance of SECP (e.g., high ionic strength buffers in the presence of detergent). The column is eluted under conditions that disrupt

antibody/SECP binding (e.g., a buffer of pH 2 to pH 3, or a high concentration of a chaotrope, such as urea or thiocyanate ion), and SECP is collected.

XVI. Identification of Molecules Which Interact with SECP

SECP, or biologically active fragments thereof, are labeled with ¹²⁵I Bolton-Hunter reagent. (See, e.g., Bolton, A.E. and W.M. Hunter (1973) Biochem. J. 133:529-539.) Candidate molecules previously arrayed in the wells of a multi-well plate are incubated with the labeled SECP, washed, and any wells with labeled SECP complex are assayed. Data obtained using different concentrations of SECP are used to calculate values for the number, affinity, and association of SECP with the candidate molecules.

Alternatively, molecules interacting with SECP are analyzed using the yeast two-hybrid system as described in Fields, S. and O. Song (1989) Nature 340:245-246, or using commercially available kits based on the two-hybrid system, such as the MATCHMAKER system (Clontech).

SECP may also be used in the PATHCALLING process (CuraGen Corp., New Haven CT) which employs the yeast two-hybrid system in a high-throughput manner to determine all interactions between the proteins encoded by two large libraries of genes (Nandabalan, K. et al. (2000) U.S. Patent No. 6,057,101).

XVII. Demonstration of SECP Activity

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An assay for growth stimulating or inhibiting activity of SECP measures the amount of DNA synthesis in Swiss mouse 3T3 cells (McKay, I. and Leigh, I., eds. (1993) Growth Factors: A Practical Approach, Oxford University Press, New York, NY). In this assay, varying amounts of SECP are added to quiescent 3T3 cultured cells in the presence of [3H]thymidine, a radioactive DNA precursor. SECP for this assay can be obtained by recombinant means or from biochemical preparations. Incorporation of [3H]thymidine into acid-precipitable DNA is measured over an appropriate time interval, and the amount incorporated is directly proportional to the amount of newly synthesized DNA. A linear dose-response curve over at least a hundred-fold SECP concentration range is indicative of growth modulating activity. One unit of activity per milliliter is defined as the concentration of SECP producing a 50% response level, where 100% represents maximal incorporation of [3H]thymidine into acid-precipitable DNA.

Alternatively, an assay for SECP activity measures the stimulation or inhibition of neurotransmission in cultured cells. Cultured CHO fibroblasts are exposed to SECP. Following endocytic uptake of SECP, the cells are washed with fresh culture medium, and a whole cell voltage-clamped Xenopus myocyte is manipulated into contact with one of the fibroblasts in SECP-free medium. Membrane currents are recorded from the myocyte. Increased or decreased current relative to control values are indicative of neuromodulatory effects of SECP (Morimoto, T. et al. (1995) Neuron 15:689-696).

Alternatively, an assay for SECP activity measures the amount of SECP in secretory, membrane-bound organelles. Transfected cells as described above are harvested and lysed. The lysate is fractionated using methods known to those of skill in the art, for example, sucrose gradient ultracentrifugation. Such methods allow the isolation of subcellular components such as the Golgi apparatus, ER, small membrane-bound vesicles, and other secretory organelles.

Immunoprecipitations from fractionated and total cell lysates are performed using SECP-specific antibodies, and immunoprecipitated samples are analyzed using SDS-PAGE and immunoblotting techniques. The concentration of SECP in secretory organelles relative to SECP in total cell lysate is proportional to the amount of SECP in transit through the secretory pathway.

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Alternatively, AMP binding activity is measured by combining SECP with ³²P-labeled AMP. The reaction is incubated at 37°C and terminated by addition of trichloroacetic acid. The acid extract is neutralized and subjected to gel electrophoresis to remove unbound label. The radioactivity retained in the gel is proportional to SECP activity.

Alternatively, the activity of purified SECP can be tested by introducing the molecule into an in vitro production system for tissue plasminogen activator (tPA). Any statistically significant improvement of correctly folded tPA in the presence as compared to the absence of SECP would indicate that SECP is active and functioning correctly.

Alternatively, SECP activity may be measured by the enzymatic activity they possess. For SEQ ID NO:4, for example, SECP activity is measured as ferroxidase activity at pH 6 in 0.3 M acetate buffer. The appearance of ferric ions is monitored at 315 nm (Bonomi, F. et al. (1996) J. Biol. Inorg. Chem. 1:67-72). For SEQ ID NO:6, for example, SECP activity is measured by the phosphorylation of galactose. SECP is incubated for 5 minutes in a 100 μ l reaction containing 200 μM ³H-galactose (30,000 cpm), 5 mM ATP, 5 mM MgCl₂, 5 mM NaF, 100 mM Tris-HCl buffer, pH 8.5. The reaction is stopped by heating at 100 °C for 1 min, and the incubation mixture applied to a DE52 column. The column is washed with at least 5 column volumes of 10 mM (NH₄)HCO₃ to remove unbound material. Galactose-P is eluted with 500 mM (NH₄)HCO₃ and assayed for radioactive content by scintillation counting (Pastuszak, I. et al. (1996) J. Biol. Chem. 271:23653-23656). For SEQ ID NO:9, for example, SECP activity is measured by the amount of cobalamin bound using the isotope dilution method of Nexø, and Gimsing, employing human IF as the binding protein (1981, Scand. J. Clin. Lab. Invest. 41:465-468). For SEQ ID NO:10, for example, SECP activity is measured by the hydrolysis of appropriate synthetic peptide substrates conjugated with various chromogenic molecules in which the degree of hydrolysis is quantified by spectrophotometric (or fluorometric) absorption of the released chromophore (Beynon, R.J. and J.S. Bond (1994) Proteolytic Enzymes: A Practical Approach, Oxford University Press, New York, NY, pp.25-55). Peptide substrates are designed according to the category of protease activity as

endopeptidase (serine, cysteine, aspartic proteases, or metalloproteases), aminopeptidase (leucine aminopeptidase), or carboxypeptidase (carboxypeptidases A and B, procollagen C-proteinase). Commonly used chromogens are 2-naphthylamine, 4-nitroaniline, and furylacrylic acid. Assays are performed at ambient temperature and contain an aliquot of the enzyme and the appropriate substrate in a suitable buffer. Reactions are carried out in an optical cuvette, and the increase/decrease in absorbance of the chromogen released during hydrolysis of the peptide substrate is measured. The change in absorbance is proportional to the enzyme activity in the assay.

Alternatively, SECP activity can be measured as enzyme activity. For SEQ ID NO:20, for example, activity is proportional to the hydrolysis of glucosamine-6-sulfate by SECP which can be measured by the method of Robertson et al. (1992, Biochem. J. 288:539-544).

In another alternative, SECP can be assayed by its interaction with the insulin-like growth factor complex. For SEQ ID NO:17, for example, ¹²⁵I-labeled SECP is incubated for 2 h with 10 ng of IGF-I or-II and a range from 0 to 10 ng of IGFBP-3 in 50 mM sodium phosphate buffer, pH 6.5, at 22 °C (final volume 0.3 ml). SECP complexed to IGFBP-3 is precipitated using IGFBP-3 antiserum and radioactivity in each tube measured (Janosi, J.B.M. et al. (1999) J. Biol. Chem. 274:5292-5298).

XVIII. Demonstration of Immunoglobulin Activity

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An assay for SECP activity measures the ability of SECP to recognize and precipitate antigens from serum. This activity can be measured by the quantitative precipitin reaction. (Golub, E.S. et al. (1987) Immunology: A Synthesis, Sinauer Associates, Sunderland, MA, pages 113-115.) SECP is isotopically labeled using methods known in the art. Various serum concentrations are added to constant amounts of labeled SECP. SECP-antigen complexes precipitate out of solution and are collected by centrifugation. The amount of precipitable SECP-antigen complex is proportional to the amount of radioisotope detected in the precipitate. The amount of precipitable SECP-antigen complex is plotted against the serum concentration. For various serum concentrations, a characteristic precipitin curve is obtained, in which the amount of precipitable SECP-antigen complex initially increases proportionately with increasing serum concentration, peaks at the equivalence point, and then decreases proportionately with further increases in serum concentration. Thus, the amount of precipitable SECP-antigen complex is a measure of SECP activity which is characterized by sensitivity to both limiting and excess quantities of antigen.

Alternatively, an assay for SECP activity measures the expression of SECP on the cell surface. cDNA encoding SECP is transfected into a non-leukocytic cell line. Cell surface proteins are labeled with biotin (de la Fuente, M.A. et al. (1997) Blood 90:2398-2405). Immunoprecipitations are performed using SECP-specific antibodies, and immunoprecipitated samples are analyzed using SDS-PAGE and immunoblotting techniques. The ratio of labeled immunoprecipitant to unlabeled immunoprecipitant is proportional to the amount of SECP expressed on the cell surface.

Alternatively, an assay for SECP activity measures the amount of cell aggregation induced by overexpression of SECP. In this assay, cultured cells such as NIH3T3 are transfected with cDNA encoding SECP contained within a suitable mammalian expression vector under control of a strong promoter. Cotransfection with cDNA encoding a fluorescent marker protein, such as Green Fluorescent Protein (CLONTECH), is useful for identifying stable transfectants. The amount of cell agglutination, or clumping, associated with transfected cells is compared with that associated with untransfected cells. The amount of cell agglutination is a direct measure of SECP activity.

Various modifications and variations of the described methods and systems of the invention will be apparent to those skilled in the art without departing from the scope and spirit of the invention. Although the invention has been described in connection with certain embodiments, it should be understood that the invention as claimed should not be unduly limited to such specific embodiments. Indeed, various modifications of the described modes for carrying out the invention which are obvious to those skilled in molecular biology or related fields are intended to be within the scope of the following claims.

10

Table 1

Incyte Project ID	Polypeptide SEO ID NO:	Incyte Polynentide ID	Polynucleotide SEO ID NO:	Incyte Polymicleotide
	200	and bolome		ID.
95765	1	095765CD1	55	095765CB1
6399886	2	6399886CD1	56	6399886CB1
6024420	3	6024420CD1	57	6024420CB1
7481067	4	7481067CD1	58	7481067CB1
3378720	5	3378720CD1	59	3378720CB1
938824	9	938824CD1	09	938824CB1
1683721	7	1683721CD1	61	1683721CB1
1694122	8	1694122CD1	62	1694122CB1
1970615	6	1970615CD1	63	1970615CB1
2314152	10	2314152CD1	64	2314152CB1
2886225	11	2886225CD1	65	2886225CB1
6144418	12	6144418CD1	99	6144418CB1
6834184	13	6834184CD1	<i>L9</i>	6834184CB1
6951005	14	6951005CD1	89	6951005CB1
7250331	15	7250331CD1	69	7250331CB1
1758413	16	1758413CD1	70	1758413CB1
7011042	17	7011042CD1	71	7011042CB1
7427362	18	7427362CD1	72	7427362CB1
7485304	19	7485304CD1	73	7485304CB1
1422394	20	1422394CD1	74	1422394CB1
1336022	21	1336022CD1	75	1336022CB1
7473674	22	7473674CD1	9/	7473674CB1
7475846	23	7475846CD1	77	7475846CB1
7475860	24	7475860CD1	78	7475860CB1
7950941	25	7950941CD1	62	7950941CB1
7485334	26	7485334CD1	80	7485334CB1
7220001	27	7220001CD1	81	7220001CB1
5956275	28	5956275CD1	82	5956275CB1
346472	29	346472CD1	83	346472CB1

Table 1

Incyte Project ID	Polypeptide	Incyte	Polynucleotide	Incyte
•	SEQ ID NO:	Polypeptide ID	SEQ ID NO:	Polynucleotide
				Д
643526	30	643526CD1	84	643526CB1
1483418	31	1483418CD1	85	1483418CB1
2683477	32	2683477CD1	98	2683477CB1
5580991	33	5580991CD1	87	5580991CB1
5605931	34	5605931CD1	88	5605931CB1
6975241	35	6975241CD1	68	6975241CB1
6988529	36	6988529CD1	06	6988529CB1
8089669	37	6996808CD1	91	6996808CB1
7472689	38	7472689CD1	92	7472689CB1
876751	39	876751CD1	93	876751CB1
2512510	40	2512510CD1	94	2512510CB1
7486326	41	7486326CD1	95	7486326CB1
1221545	42	1221545CD1	96	1221545CB1
124737	43	124737CD1	26	124737CB1
1510784	4	1510784CD1	86	1510784CB1
1901257	45	1901257CD1	99	1901257CB1
2044370	46	2044370CD1	100	2044370CB1
2820933	47	2820933CD1	101	2820933CB1
2902793	48	2902793CD1	102	2902793CB1
7486536	49	7486536CD1	103	7486536CB1
8137305	50	8137305CD1	104	8137305CB1
3793128	51	3793128CD1	105	3793128CB1
4001243	52	4001243CD1	106	4001243CB1
6986717	53	6986717CD1	107	6986717CB1
7503512	54	7503512CD1	108	7503512CB1

Table 2

Polypeptide SEO Incyte	Incyte	GenBank ID NO: Probability	Γ	Annotation
D NO:	ptide ID	or PROTEOME ID NO:		
1	95765CD1	g190183	2.6E-76	[Homo sapiens] opiomelanocortin Krude, H. et al. (1998) Nature Genet. 19:155-157
2	6399886CD1	g6996429	5.8E-262	dJ568C11.3 (novel AMP-binding enzyme similar to acetyl-coenzyme A synthethase (acetate-coA ligase)) [Homo sapiens] Luong, A. et al., (2000) J. Biol. Chem. 275:26458-26466
93	6024420CD1	g163497	1.1E-70	Bos taurus] PDI (E.C.5.3.4.1) (protein disulfide isomerase) Yamauchi, K., et al. (1987) Biochem. Biophys. Res. Commun. 146, 1485-1492
4	7481067CD1	g180256	0	[Homo sapiens] preceruloplasmin (EC 1.16.3.1) Waggoner, D.J. et al. (1999) Neurobiol. Dis. 9:221-230; Hellman, N.E. et al. (2000) Gut 47:858-860
5	3378720CD1	£006586g	9.1E-65	[Homo sapiens] tumor metastasis-suppressor
16	1758413CD1	g183178	5.6E-106	[Homo sapiens] hGH-V2 Cooke, N.E. et al. (1988) J. Biol. Chem. 263:9001-9006
17	7011042CD1	g184808	3.0E-11	[Homo sapiens] insulin-like growth factor binding protein complex, acid-labile subunit Leone, S.R. et al. (1992) Mol. Endocrinol. 6:870-876
18	7427362CD1	g14530679	7.0E-70	WNT3A [Homo sapiens] Saitoh,T., et al (2001) Biochem. Biophys. Res. Commun. 284:1168-1175
19	7485304CD1	g2623871	2.1E-115	[Gallus gallus] Wnt-14 protein Bergstein, I. et al. (1997) Genomics 46:450-458
20	1422394CD1	g15430244	0	N-acetylglucosamine-6-sulfatase [Coturnix coturnix] Dhoot, G.K., et al (2001) Science 293:1663-6.
22	7473674CD1	g13620917	9.0E-46	mitochondrial ribosomal protein bMRP63 [Mus musculus] Suzuki,T., et al (2001) J. Biol. Chem. 276:33181-33195
26	7485334CD1	g10566471	1.2E-73	[Mus musculus] Gliacolin Koide, T. et al. (2000) J. Biol. Chem. 275:27957-27963

Table 2

Polypeptide SEQ Incyte ID NO: Polype	ptide ID	Incyte GenBank ID NO: Probability Polypeptide ID or PROTEOME Score ID NO:	Probability Score	Annotation
27	7220001CD1	g11071950	2.2E-115	[Mus musculus] (AB048834) Fca/m receptor Shibuya,A., et al (2000) Nat. Immunol. 1:441-446
28	5956275CD1	g7259265	4.4E-129	[Mus musculus] contains transmembrane (TM) region Inoue, S., et al. (2000) Biochem. Biophys. Res. Commun. 268, 553-561
39	876751CD1	g15430246	0	nephronectin short isoform [Mus musculus] Brandenberger, R., et al (2001) J. Cell Biol. 154:447-458
40	2512510CD1	g14423349	0	membrane glycoprotein LIG-1 [Homo sapiens]
41	7486326CD1	g3822218	0	[Homo sapiens] chordin Pappano, W. N. et al. (1998) Genomics 52:236-239
. 48	2902793CD1	g15026974	2.0E-36	obscurin [Homo sapiens] Young,P., et al (2001) J. Cell Biol. 154: 123-136
54	7503512CD1	g14423349	0	membrane glycoprotein LIG-1 [Homo sapiens]

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Table 3

Analytical Methods	and Databases	SPSCAN	HMMER	HMMER_PFAM		TIMAP		BLIMPS_PRINTS		BLAST_PRODOM				BLAST_PRODOM			BLAST_PRODOM				BLAST_PRODOM				BLAST_DOMO
Signature Sequences, Domains and Motifs		signal_cleavage: M1-E23	Signal Peptide: M1-E23, M1-G26, M1-V24, M1-C28, M1-E30	Corticotropin ACTH domain:	S106-F144, P188-T220	Transmembrane Domain: C6-W27	N-terminus is non-cytosolic	Pro-opiomelanocortin signature	PR00383: Y107-P117, V118-E133, D134-E143, A181-W196, W196-M209	ROPIN LIPOTROPIN	田	SIGNAL ENDORPHIN CLEAVAGE ON PAIR	PD004218:L12-Q68	PRO-OPIOMELANOTROPIN POM PRECURSOR	SIGNAL	PD116389: R101-N229, C28-W84		LIPOTROPIN PRO-OPIOMELANOCORTIN POMC	CLEAVAGE ON PAIR OF BASIC	PD003250: S106-F144	CORTICOTROPIN LIPOTROPIN PRECURSOR	PRO-OPIOMELANOCORTIN POMC ENDORPHIN	HORMONE CLEAVAGE ON PAIR OF	PD029102: D164-N229	CORTICOTROPIN-LIPOTROPIN DAMO1703IB01100[1-83: M1-W84
Potential	Glycosylation Sites	16N																							
Acid Potential	Phosphorylation Sites	S92 S211 Y189																							
Amino Acid	Residues	235									•														
Incyté	Polypeptide ID	095765CD1																							
SEQ ID Incyte	NO:																								

Table 3

SEQ ID	Incyte	Amino Acid	Acid Potential	Potential	Signature Sequences, Domains and Motifs	Analytical Methods
NO:	NO: Polypeptide ID	Residues	Phosphorylation Sites	Glycosylation Sites		and Databases
1 (cont.)					CORTICOTROPIN-LIPOTROPIN DM00964[P01190[171-264: K145-E235	BLAST_DOMO
					CORTICOTROPIN-LIPOTROPIN DM01793[p19402]1-79: M1-V78	BLAST_DOMO
					1 3235	BLAST_DOMO
2	6399886CD1	689	S16 S409 S415 S486 S601 S640 T89 T97 T142 T150 T156 T318 T438 T546 T663 T664	N109 N492 N554		HMMER
					AMP-binding enzyme: T142-V580	HIMMER_PFAM
					ıs:	TMAP
					P173-A201, B463-P478 N-terminus is cytosolic	
					Putative AMP-binding domain signature: A272-V322 PROFIL ESCAN	PROFILESCAN
					Putative AMP-binding domain BL00455:Y293-Q308	BLIMPS_BLOCKS
					AMP-binding signature PR00154:E286-S297, T298-H306	BLIMPS_PRINTS
					LIGASE SYNTHETASE PROTEIN ENZYME BIOSYNTHESIS ANTIBIOTIC	BLAST_PRODOM
					PHOSPHOPANTETHEINE MULTIFUNCTIONAL	
					REFEAT ACTL-COA PD000070: T142-T438, D482-I581	

Table 3

SEQ ID	SEQID Incyte	Amino Acid Potential		Potential	Signature Sequences, Domains and Motifs	Analytical Methods
Ö	Polypeptide ID	Residues	Phosphorylation Sites	Glycosylation Sites		and Databases
2 (cont.)					SYNTHETASE LIGASE ACETYL COENZYME A BLAST_PRODOM	BLAST_PRODOM
	<u> </u>				ENZYME ACETATE-COA ACETYL-COA	
					PROTEIN ACYL ACTIVATING	
					PD009307: Y58-L146	
					PUTATIVE AMP-BINDING DOMAIN	BLAST_DOMO
					DM00073	
					S46276 96-631: L115-R649	
					P27550[82-615: L115-R649	
					S52154 15-546: 1141-L648	
					P16928 102-634: L115-R649	
					Putative AMP-binding domain signature	MOTIFS
					M291-K302	
3	6024420CD1	584	S20 S71 S134 S153 N58 N160 N238	N58 N160 N238	signal_cleavage: M1-S20	SPSCAN
			S358 S376 S417	N340 N370 N436		
			S546 S558 T128	N540		
			T241 T329 T448			
			T481 T495 T557			
			Y356			
					Signal Peptide: M1-S20; M1-E23, M1-A26	HMMER
					Thioredoxin domain: Q386-D451	HMMER_PFAM
					Transmembrane Domain: P199-H216	TMAP
					N-terminal is non-cytosolic	•
					Thioredoxin family active site: L392-1440	PROFILESCAN
					Thioredoxin family proteins signature	BLIMPS_BLOCKS
					BL00194: F409-K421	

Table 3

	Slycosylation	phorylation
aa aa vaa day	ites	Sites
ISOMEKASE PRECUKSOR PROTEIN SIGNAL REDOX ACTIVE CENTER DISULFIDE ENDOPLASMIC RETICULUM REPEAT		
PD001708: Q55-L367		
REDOX ACTIVE CENTER PROTEIN ISOMERASE PRECURSOR THIOREDOXIN		
SIGNAL DISULFIDE ENDOPLASMIC ELECTRON		
PD000175: V389-K496		
PROTEIN DISULFIDE-ISOMERASE		
DM00799 P09102 112-347: L148-D385		
DM00799[P54399]129-364: E167-D385		-
THIOREDOXIN FAMILY		
DM00054P09102 349-452:		
1169 signal_cleavage: M1-A19	4111 N137 N	S38 S180 S221 N111 N137 N169
	4245 N270 I	
1549	1460 N475 N	
6281	4575 N781 N	
	4830 N908	T184 T198 T203 N830 N908
		T208 T222 T311
		T389 T414 T430
		T452 T515 T570
		T589 T661 T688
		T783 T855 T1007
		T1037 Y134 Y483
		Y566 Y715
OL 134 0171 137 131 134 - 153 - 1710 141 4 10		

Table 3

SEQ ID	Incyte	Amino Acid Potential	Potential	Potential	Signature Sequences, Domains and Motifs	Analytical Methods
NO:	NO: Polypeptide ID	Residues	Phosphorylation Sites	Glycosylation Sites		and Databases
4 (cont.)					Transmembrane Domain: 1794-1809	TIMAP
					Multicopper oxidase: L900-R1041, L567-C705, L218-HMMER_PFAM C352	HMMER_PFAM
					Multicopper oxidases signatures	PROFILESCAN
					multicopper_oxidase1.prf: D310-V366, D660-E742, D996-L1050	
					multicopper_oxidase2.prf: P1001-N1048	
					Multicopper oxidases signature 1: G680-Y700,	MOTIFS
					G1016-Y1036	
					Multicopper oxidases signature	BLIMPS_BLOCKS
					BL00079A: G93-N110, R817-T828, L975-F985,	
					D1014-M1032	
					FERROXIDASE REPEAT	BLAST_DOMO
					DM00956 P00450 90-336: L89-L334, L438-L684,	
					P838-L1019	
					DM00956 P00450 445-697: H437-L684, L89-S318,	
					Y819-W1018,	
					H791-S831 DM00956 P00450 804-1038: H791-	
					Y1021, H437-G680, L89-L334	
-					DM00956P12259 83-308: G90-P249, G439-S577,	
					1794-V992,	
					E252-L334, D595-T675, S538-L552	
					FACTOR PRECURSOR GLYCOPROTEIN	BLAST_PRODOM
					PLASMA REPEAT SIGNAL COAGULATION	
					BLOOD VIII CALCIUM	
					PD002090: H368-K559, R22-N205,	
					K707-P893	

Table 3

Analytical Methods	aDasca.	8	7.			BLAST_PRODOM			7	~			PROFILESCAN	7	_	7	7	~			
Analytical Me	מוום המווי	MOTIFS	SPSCAN	TMAP		BLAST			SPSCAN	HMMER	TMAP		PROFIL	SPSCAN	HIMIMER	SPSCAN	SPSCAN	HIMMER		TMAP	
Signature Sequences, Domains and Motifs		ATP/GTP-binding site motif A (P-loop): G335-S342	signal_cleavage: MI-E57	Transmembrane Domains: H38-A61, C136-Y160, Y181-F198, K201-R229, N261-F281, H294-L314	N terminus is non-cytosolic	PROTEIN TRANSMEMBRANE LONGEVITY ASSURANCE FACTOR UOGI SIMILAR S	CEREVISIAE	PD006418: S119-L369	signal_cleavage: M1-A63	Signal Peptide: M1-G29	Transmembrane Domain: P9-A36	N terminus is non-cytosolic	Galactokinase signature galactokinase.prf: S17-A67	signal_cleavage: M1-P23	Signal Peptide: M1-P23	signal_cleavage: M1-R62	signal_cleavage: M1-A26	Signal Peptide:	M1-A26, M1-R27, M1-V28, M1-C30	Transmembrane Domain: Q4-C20	N-terminus is non-cytosolic
Potential	Grycosyranon Sites		N107																!		
Acid Potential	Fnospnorylanon Sites		S23 S90 S119 S247 N107 S334 S340 T4 T19 T73 T78 T81 T109 Y345			,								S41 T34 T53		T19	S55				
Amino Acid	Kesidues		383						72					16		160	95				
Incyte	roiypepuae in		3378720CD1						938824CD1					1683721CD1		1694122CD1	1970615CD1				
SEQ ID Incyte	Con	4 (cont.)	5						9					7		8	6				

Table 3

OEO II	Increte	Aming Acid Dotantial	Detentiol	Detentiof	Circulation Commence Demoins and Maritia	A male strain 1 Marke de
))	and any		1 Occupat	T Oreninai	orginature ocquerices, Domains and Monts	Analytical Mellions
:ON	Polypeptide ID	Residues	Phosphorylation Sites	Glycosylation Sites		and Databases
9 (cont.)					Eukaryotic cobalamin-binding proteins signature cobalamin_binding.prf: D25-R72	PROFILESCAN
10	2314152CD1	92	S11 S23 S33 S52 T65 Y43		signal_cleavage: M1-S19	SPSCAN
					Signal Peptide: M1-R25, M1-S24	HMMER
					Endopeptidase Clp active sites clpp_protease_ser.prf: PROFILESCAN L32-G80	PROFILESCAN
11	2886225CD1	71	S41 T37 Y67		signal_cleavage: M1-L29	SPSCAN
					Signal Peptide: M1-L29	HIMIMER
					15-C33	TMAP
					N-terminus is non-cytosolic	
					Vitamin K-dependent carboxylation domain	PROFILESCAN
					glu_carboxylation.prf: M1-G70	
12	6144418CD1	100	S46 S69 T50 T60 T84	N67 N85	signal_cleavage: M1-G36	SPSCAN
					Signal Peptide: M1-A29, M1-V31	HMMER
_						TMAP
13	6834184CD1	122		N108	signal cleavage: MI-E38	SPSCAN
					Signal Peptide: M1-G25	HMMER
					: G8-T36, C81-I105	TMAP
	•				N-terminus is non-cytosolic	
14	6951005CD1	113	S25 T35 Y81		signal_cleavage: M1-D28	SPSCAN
					Signal Peptide: M1-Q29, M1-D28	HMMER
					Transmembrane Domain: S50-S72	TMAP
					N-terminus is non-cytosolic	
15	7250331CD1	85			signal_cleavage: MI-L27	SPSCAN

Table 3

SEQ ID Incyte Amino Acid Potential NO: Polymentide ID Residues Phosphor	cid	Phosnhor Phosnhor	vlation	Potential Glycosylation	Signature Sequences, Domains and Motifs	Analytical Methods
Sites	Sites		Site	cosyianon :S		ally Databases
			1		Signal Peptide: M1-P23, M1-V25, M1-R26, M1-L27, M1-C35	HMMER
					Immunoglobulins and major histocompatibility complex proteins signature ig_mhc.prf: L14-L64	PROFILESCAN
1758413CD1 256 S88 S97 S111 S132 S181 S194 T53		S88 S97 S111 S132 S181 S194 T53			signal_cleavage: M1-A26	SPSCAN
					Signal Peptide: M1-A26	HIMMER
			_		7-L35	TMAP
		•			N-terminus is non-cytosolic	
					Somatotropin hormone family: L9-M151	HIMMER_PFAM
					Somatotropin, prolactin and related hormones somatotropin 1.prf: S88-H138	PROFILESCAN
					Somatotropin, prolactin and related hormones signature 1: C79-W112	MOTIFS
					Somatotropin, prolactin and related hormones BI 00266: 135-V61 C79-V116 D135-M151 P201-	BLIMPS_BLOCKS
					P224	
					Somatotropin hormone family signature PR00836: C79-E92, L101-L119	BLIMPS_PRINTS
			L		GROWTH HORMONE VARIANT II PRECURSOR BLAST_PRODOM	BLAST_PRODOM
					GHV2 PLACENTA SIGNAL ALTERNATIVE	
					SPLICING	
			[PD084405: P165-V256	
					HORMONE PRECURSOR SIGNAL PITUITARY	BLAST_PRODOM
					GROWTH SOMATOTROPIN PROLACTIN	
					GLYCOPROTEIN PRL	
					PD000259: T7-M151	

Table 3

Analytical Methods and Databases	BLAST_DOMO	SPSCAN	HMMER	HMMER_PFAM	अमारावव अवस्था रव	MOTIFS	SPSCAN		HMMER	TMAP	HMMER_PFAM		PROFILESCAN	MOTIFS	BLIMPS_BLOCKS	
Signature Sequences, Domains and Motifs	SOMATOTROPIN, PROLACTIN AND RELATED HORMONES DM00125 P09587 17-227: C17-P228 DM00125 P01243 17-212: C17-G163 DM00125 F01242 17-212: C17-G163 DM00125 P01242 17-212: C17-G163	signal_cleavage: M1-A18	Signal Peptide: M1-A18, M1-W17	Leucine Rich Repeat: N254-P278, Q100-A122, A171-HMMER_PFAM D199, T205-P228, G123-S146, T76-E99, S147-P170,	K229-P253	Leucine Kich Kepeat r K00015A; L124-L137 Leucine zioner pattern: L59-L80, L66-L87			Signal Peptide: M1-A29	5-L33	wnt family of developmental signaling proteins: A58- HMIMER_PFAM	K365	Wnt-1 family signature wnt1.prf: M196-K245	s: C216-C225	Wnt-1 family proteins	BL00246: M196-Y248, N319-C364, A85-C104, G118-D152, W163-E187
Potential Glycosylation Sites		N42 N176					N103									
Potential Phosphorylation Sites		S16 S46 S91 S147 176 T105 T214 T242					S89 T34 T81 T150 N103	T356 T363								
Amino Acid Potential Residues Phosphor Sites		287					366									
Incyte Polypeptide ID		7011042CD1					7427362CD1									
SEQ ID Incyte NO: Polype	16 (cont.)	17					18									

Table 3

Analytical Methods	and Databases	BLAST_PRODOM	BLAST_DOMO	MMER_PFAM	PROFILESCAN	MOTIFS	BLIMPS_BLOCKS	BLAST_PRODOM:	BLAST_DOMO
Signature Sequences, Domains and Motifs	a	DEVELOPMENTAL GLYCOPROTEIN PRECURSOR SIGNAL WNT1 WNT5A WNT2 EXTRACELLULAR MATRIX PD000810: C59-A195, E153-K365	55 74 55 55	wnt family of developmental signaling proteins: Q113-HMMER_PFAM K415	Wnt-1 family signature wntl.prf: D250-K298	Wnt-1 family signature: C269-C278	W216-R240,	PROTEIN DEVELOPMENTAL GLYCOPROTEIN B PRECURSOR SIGNAL WNT1 WNT5A WNT2 EXTRACELLULAR MATRIX PD000810: L119-A251, D206-K415	WNT-1 FAMILY DM00403 P49340 30-391: L119-S365, A334-C414, S69-Q106 DM00403 P21551 32-368: L119-C414 DM00403 P47793 24-351: C114-K415 DM00403 P22727 24-363: E211-L410, K118-G209
Potential	Glycosylation Sites			N158					
Potential	Phosphorylation Sites			<i>S72</i> S120 S228 S238 T87 T136 T203 T268 T413 Y342					
Amino Acid Potential	Residues			416					
Incyte	Polypeptide ID			7485304CD1					
A	NO:	18 (cont.)		19					

Table 3

Analytical Methods and Databases	SPSCAN	HIMMER	TMAP		HMMER PFAM	MOTIFS	BLIMPS_BLOCKS			BLAST_PRODOM		BLAST_PRODOM	
Signature Sequences, Domains and Motifs	signal_cleavage: M1-C22	Signal Peptide: M1-C22	Transmembrane Domain: C5-L21, S353-D370	IN-terminus is cytosolic	Sulfatase: P43-T467	Sulfatases signature 1: P85-G97	Sulfatases proteins	BL00523: P43-S59, C87-K98, G134-L144,	P214-H225, V289-G318, D363-G373, Y800-Q809	ARYLSULFATASE SIGNAL GLYCOPROTEIN LYSOSOME SULPHOHYDROLASE	MUCOPOL YSACCHARIDOSIS PD001700: P43-I200, Q279-P378, K330-E392	N-ACETYLGLUCOSAMINE-6-SULFATASE PRECURSOR GLUCOSAMINE-6-SULFATASE HYDROLASE LYSOSOME SIGNAL	GLYCOFKO LEIN MUCUPUL YSAUCHAKIDUSIN PD022780: C766-G837
Potential Glycosylation Sites	N64 N111 N131 N148 N170 N197 N240 N623 N773 N783												
Potential Phosphorylation Sites	S27 S206 S288 S425 S468 S488 S505 S516 S520 S635 T24 T66 T96 T107 T367 T376 T400 T452 T484 T530 T611 T615												
Amino Acid Potential Residues Phosphor Sites	871												
SEQ ID Incyte NO: Polypeptide ID	1422394CD1												
SEQ ID NO:	20												

Table 3

Analytical Methods and Databases	BLAST_PRODOM	BLAST_DOMO	SPSCAN	HMMER	TMAP	SPSCAN	SPSCAN	SPSCAN	HMMER	SPSCAN	HMMER	HIMIMER_PFAM	MOTIFS
Signature Sequences, Domains and Motifs	SIMILAR TO SULFATASE PD122645: V385-Q487, H700-F765	ARYLSULFATASE; SULFATASE; PLANT; DM08669 Q10723 23-520: R280-R490, R42-Q279, N773-P843 DM08669 P14217 24-519: L278-G373, R42-P269, S770-P843, E294-Y310 DM01026 P50842 24-548: V289-D390, R42-Y143 DM01026 P50473 63-522: E273-L389, R42-P153	signal_cleavage: M1-V24		: V4-P23 blic	signal_cleavage: M1-G19	signal_cleavage: M1-C32		Signal Peptide: M1-G24	signal_cleavage: M1-A15	Signal Peptide: M1-A20, M1-A21	C1g domain: A160-L284	Clq domain signature: F177-Y207
Potential Glycosylation Sites			NS			96N		N46 N66					
Potential Phosphorylation Sites		,	S3 S72 S93			S28 S102 T79 T98 N96		S65 T58 T99 T113 N46 N66 T138 Y143	S10 S34 S39 S49	S111 S167 S255 T28 T142 T192			
Amino Acid Potential Residues Phosphoi Sites			100			102	117	150	68	287			
Incyte Polypeptide ID			1336022CD1			7473674CD1	7475846CD1	7475860CD1	7950941CD1	7485334CD1			
SEQ ID Incyte NO: Polype	20 (cont.)		21			22	23	24	25	26			

Table 3

						_									
Analytical Methods and Databases	BLIMPS_BLOCKS	BLIMPS_PRINTS	HMMER_PFAM	PROFILESCAN	BLAST_PRODOM		BLAST_PRODOM		BLAST_PRODOM		BLAST_PRODOM	BLAST_DOMO			
Signature Sequences, Domains and Motifs	C1q domain proteins BL01113: V174-I209, D243-K262, S277-P286, G85- S111	Complement C1Q domain signature PR00007: P168-K194, F195-G214, D243-D264, K275-Y285	Collagen triple helix repeat (20 copies): P71-V129	Lysosome-associated membrane glycoproteins lamp_2.prf: E145-L175	COLLAGEN REPEAT LYCOPROTEIN CHAIN LULAR MATRIX	PD002992: P168-L284	COLLAGEN ALPHA PRECURSOR REPEAT SIGNAL CONNECTIVE TISSUE	EXTRACELLULAR MATRIX PD000007: G43-G118	PROCOLLAGEN TYPE XVII ALPHA 1 BULLOUS BLAST_PRODOM PEMPHIGOID AUTO-ANTIGEN CELL	ADHESION PD071338: G43-G112	PRECOLLAGEN P PRECURSOR SIGNAL PD072959: G43-G128	CIQ DOMAIN	DM00777P23206/477-673: G88-P286	DM00777 P98085 222-418: G85-D287	DM00777P27658 551-743: G88-P286
Potential Glycosylation Sites													•		
Acid Potential es Phosphorylation Sites								; ;							
Amino Acid Residues								-							
SEQ ID Incyte NO: Polypeptide ID															
SEQ ID NO:	26 (cont.)														

Table 3

Signature Sequences, Domains and Motifs Analytical Methon and Databases signal_cleavage: M1-A61 SPSCAN Signal Peptide: M46-A61, M46-P63, M46-Q64 HMMER Immunoglobulin domain: G120-1200 HMMER Transmembrane Domain: S39-P67 R496-R518 TMAP N-terminus is cytosolic TMAP POL YMERIC IMMUNOGLOBULIN RECEPTOR BLAST_PRODO PRECURSOR PLGR CONTAINS: SECRETORY BLAST_PRODO REPEAT PRO03917: G120-E203 (P-value = 5.4e-09) BLAST_DOMO DM00001P01833[41-120: H128-G201 BLAST_DOMO DM00001[P01832]28-125: G120-G201 BM00001[P01832]28-125: G120-G201 DM000001[P01834[41-120: H128-G201 BM00001[A98841[41-120: H128-G201 DM000001[S48841[41-120: H128-G201 BM00001[S48841[41-120: H128-G201 DM000001[S48841[41-120: H128-G201 BM00001[S48841[41-120: H128-G201	şş						¥									_	_
Signature Sequences, Domains and Motifs signal_cleavage: M1-A61 Signal Peptide: M46-A61, M46-P63, M46-Q64 Immunoglobulin domain: G120-1200 Transmembrane Domain: S39-P67 R496-R518 N-terminus is cytosolic POLYMERIC IMMUNOGLOBULIN RECEPTOR PRECURSOR PLGR CONTAINS: SECRETORY COMPONENT IMMUNOGLOBULIN FOLD REPEAT PD003917: G120-E203 (P-value = 5.4e-09) IMMUNOGLOBULIN DM00001 P15083 41-120: H128-G201 DM00001 P15083 41-120: H128-F208 DM00001 S1832 28-125: G120-G201 DM00001 S183841 41-120: H128-G201 Signal_cleavage: M1-A28	Analytical Methods	and Databases	SPSCAN	HMMER	HIMIMER_PFAM	TMAP	BLAST_PRODOM				:	BLAST_DOMO			SPSCAN		
	Signature Sequences, Domains and Motifs		signal_cleavage: M1-A61	Signal Peptide: M46-A61, M46-P63, M46-Q64	Immunoglobulin domain: G120-I200	Transmembrane Domain: \$39-P67 R496-R518 N-terminus is cytosolic	POLYMERIC IMMUNOGLOBULIN RECEPTOR	PRECURSOR PLGR CONTAINS: SECRETORY	COMPONENT IMMUNOGLOBULIN FOLD DEBEAT	PD003917: G120-E203	(P-value = 5.4e-09)	IMMUNOGLOBULIN DM00001P01833 41-120: H128-G201	DM00001 P15083 41-120: H128-F208	DM00001P01832 28-125; G120-G201 DM00001 S48841 41-120; H128-G201	signal_cleavage: M1-A28		
	Potential Potential	Phosphorylation Glycosylation Sites Sites	S39 \$108 \$189 S252 \$227 \$302 \$406 \$483 \$494 \$526 T6 T38 T88 T234 T272 T336 T350 T351 T438 T487 T525 T570 Y24												S109 S133 S256	T38 T91 T100	T191 Y125
S S 189 3 S 189 37 S 302 33 S 494 T 38 T 88 72 T 336 51 T 438 25 T 570	Amino Acid	Residues	<i>578</i>												285		
ino Acid Potential Fhosphorylation Sites S39 S108 S189 S252 S297 S302 S406 S483 S494 S526 T6 T38 T88 T234 T272 T336 T350 T351 T438 T487 T525 T570 Y24 S258 S109 S133 S256 T38 T88 T272 T336 T38	Incyte	ptide ID													5956275CD1		
Ptide ID Residues Phosphorylation Sites IICD1 578 S39 S108 S189 S252 S297 S302 S406 S483 S494 S526 T6 T38 T88 T234 T272 T336 T350 T351 T438 T487 T525 T570 Y24 S252 S297 S302 S406 S483 S494 S526 T6 T38 T88 T487 T525 T570 T236 T356 T351 T438 T487 T525 T570 T236 T356 T351 T438 T487 T525 T570 T236 T336 T336 T338 T3556 T35D1 285 S109 S133 S256 T38 T91 T100	SEQ ID Incyte	ÖN	27												28		

Table 3

۲	SEO ID Incyte	Amino Acid Potential	Potential	Potential	Signature Sequences Domains and Motifs	Anslytical Methode
0	Polypeptide ID	Residues	Phosphorylation Sites	Glycosylation Sites		and Databases
f 1					Transmembrane Domain: Q3-T31 N-terminus is non-cytosolic	TMAP
1 1					Cell attachment sequence: R197-D199	MOTIFS
121	346472CD1	72	\$25		signal_cleavage: M1-C14	SPSCAN
					Signal Peptide: M1-S16, M1-S20, M1-N21, M1-F22, HMMER M1-H24 M1-S25 M1-K27	HMMER
1					Aminotransferases class-V pyridoxal-phosphate	PROFILESCAN
- 1					attachment site: S25-S72	
141	643526CD1	72			signal_cleavage: MI-C18	SPSCAN
					Signal Peptide: M1-C18, M1-S19, M1-E20, M1-S21, HMMER	HIMIMER
- 1					M1-G23, M1-S24, M1-P26	
- 1					S24-V41	TMAP
41	1483418CD1	149	S65 S70			SPSCAN
1					M1-V34, M1-S35	HMMER
			-			TMAP
JI.					N-terminus is non-cytosolic	
9	2683477CD1	100	S71 T54 T81	69N	Signal Peptide: M7-L35	HMMER
					13-R38	TMAP
- li					N-terminus is non-cytosolic	
ו נאו	5580991CD1	78	S50		signal_cleavage: M1-C24	SPSCAN
					Signal Peptide: M4-S22, M4-C24, M4-P25, M4-S26, HMMER	HMMER
- 1					M4-S29	
					Transmembrane Domain: F13-F41	TMAP
- 1					N-terminus is non-cytosolic	
- 1					CBF/NF-Y subunits signatures: M4-S73	PROFILESCAN
9	5605931CD1	75	S65		Signal Peptide: M29-A43	HMMER
_ /					Signal Peptide: M1-Q28	HMMER

Table 3

SEQ ID Incyte	Incyte	Amino Acid	Acid Potential	Potential	Signature Sequences, Domains and Motifs	Analytical Methods
NO:	Polypeptide ID	Residues	Phosphorylation Sites	Glycosylation Sites		and Databases
34 (cont.)					Transmembrane Domain: A5-F25 N-terminus is non-cytosolic	TMAP
35	6975241CD1	111	S6 S51 S75			SPSCAN
					Signal Peptide: M1-E22, M1-K24, M1-K24, M1-G28, HMMER	HMMER
36	 6088529CD1	77	C30 T35		Jeanage: M1-A15	SPSCAN
3					M1-T17, M1-A20	HMMER
37	6996808CD1	06	S34 S42 S68			SPSCAN
					M17-L38, M17-S41	HMMER
					212-F40, R44-I72	TMAP
					N-terminus is non-cytosolic	
38	7472689CD1	283	S66 S140 S197	N131	Signal Peptide: M21-G38	HMMER
			T259			
39	876751CD1	299	S111 S238 S407	N274	Signal_Peptide: M1-A19	SPSCAN
			S485 S556 T167			
			T176 T308 T312			
			T316 T320 T346			
			T527 Y206			
					Signal Peptide: M1-A18, M1-E20	HMMER
					EGF-like domain: C94-C128, C174-C213, C219-	HMMER_PFAM
					C254, C134-C168, C61-C87	
	!				MAM domain: C423-E564	HIMMER_PFAM
					Anaphylatoxin domain proteins	BLIMPS_BLOCKS
					BL01177: R99-L117, P163-S180	
					Calcium-binding BGF-like domain	BLIMPS_BLOCKS
					BL01187: C105-Y120, C168-A179	

Table 3

otifs Analytical Methods		NAL BLAST_PRODOM PROTEIN				BLAST_DOMO		MOTIFS	ution site: C105-MOTIFS		MOTIFS	C114-C128 MOTIFS		tern signature: MOTIFS		SPSCAN											
Signature Sequences, Domains and Motifs		PRECURSOR GLYCOPROTEIN SIGNAL TRANSMEMBRANE HYDROLASE PROTEIN	REPEAT RECEPTOR PHOSPHATASE	NEUROPILIN	PD001482: W432-E564	PROLINE-RICH PROTEIN 3	DM00215 P41479 30-111: P297-1369	Cell attachment sequence: R383-D385	Aspartic acid and asparagine hydroxylation site: C105 MOTIFS	C116 C187-C198 C232-C243	EGF-like domain signature 1: C76-C87	EGF-like domain signature 2: C76-C87 C114-C128	C241-C254	Calcium-binding EGF-like domain pattern signature:	D90-C114 D170-C196 D215-C241	N74 N150 N246 Signal_Peptide: M1-A33											N
Potential Glycosylation	Sites															N74 N150 N246	N292 N318 N684							•			
Potential Phosphorylation	Sites															S81 S270 S326	S356 S379 S403	S473 S588 S705	S734 S822 S850	866S 786S 076S	T192 T214 T320	T370 T499 T538	T604 T609 T688	T736 T771 T818	T828 T848 T936	V670	
Amino Acid Potential Residues Phosphor																1093											
Incyte Polypeptide ID				-												2512510CD1											
SEQ ID Incyte		39 (cont.)														9											

Table 3

Г									Γ		_
Analytical Methods	and Databases	HMMER_PFAM	HMMER_PFAM	HMMER_PFAM	TMAP	BLIMPS_PRINTS	BLIMPS_PRINTS	BLAST_PRODOM	BLAST_PRODOM	BLAST_DOMO	MOTIFS
Signature Sequences, Domains and Motifs		Leucine Rich Repeat: S332-R355, S236-S259, K308- HMMER_PFAM S331, R212-N235, G407-K430, S356-D382, N93-S114, S189-P211, K260-T283, S383-E406, P164-S187, S140-P163, W69-P92, A284-Q307, H116-K136	Leucine rich repeat C-terminal domain: D440-D490	Immunoglobulin domain: G707-M765, T613-A674, G509-I579	Transmembrane Domain: R13-A33 N terminus non-cytosolic	Bee Venom Hyaluronidase PR00847F; R738-V754	Leucine-rich repeat signature PR00019 L91-L104, L141-V154	MEMBRANE GLYCOPROTEIN PD129774: M765-S1093 PD172109: D491-F583 PD165826: E29-T70	SIMILARITY MULTIPLE LEUCINE RICH PD037237: L432-1610	IMMUNOGLOBULIN DM00001[P35918[668-745: L699-A774 DM00001[A46065]668-745: L699-A774	Leucine zipper pattern: L52-L73 L59-L80
Potential	Glycosylation Sites										
Acid Potential	Phosphorylation Sites										
Amino Acid	Residues										
SEQ ID Incyte	Polypeptide ID										
SEQ ID	NO:	40 (cont.)									

Table 3

SEQ ID Incyte	Incyte	Amino Acid Potential		Potential	Signature Sequences, Domains and Motifs	Analytical Methods
NO:	Polypeptide ID	Residues	Phosphorylation Sites	Glycosylation Sites		and Databases
41	7486326CD1	915	S38 S98 S130 S132 S151 S153 S195 S201 S245 S306 S353 S488 S596 S751 S882 T304 T367 T592 T775 T859 T902 T904 Y152 Y745	N217 N351 N365 N434	S38 S98 S130 S132 N217 N351 N365 Signal_Peptide: M1-P23 S151 S153 S195 N434 S201 S245 S306 S353 S488 S596 S751 S882 T304 T367 T592 T775 T859 T902 T904 Y152 Y745	SPSCAN
					Signal Peptide: M1-P23, M1-G26	HMMER
					von Willebrand factor (growth regulator) type C domain: C51-C125, C832-C892, C744-C810, C665- C722	HMMER_PFAM
					Transmembrane Domain: P5-R25 N terminus non-cytosolic	TMAP
					CHORDIN PD018424: Q440-L618, G232-Y439, F141-P292, A413-V605 PD069130: P811-C892PD015143: P662-D727	BLAST_PRODOM
					VON WILLEBRAND FACTOR TYPE C REPEAT DM00551 A55195 22-117: P31-P126 DM00551 A55195 752-835: E724-K808, P656-C719 DM00551 A55195 637-751: R660-C722, R738-C810, T592-L618 DM00551 A55195 836-920: Q809-C892	BLAST_DOMO
					Cell attachment sequence R165-D167	MOTIFS
					VWFC domain signature: C686-C722 C771-C810	MOTIFS

Table 3

SEO ID	Incyte	Amino Acid	Acid Potential	Potential	Signature Sequences, Domains and Motifs	Analytical Methods
ÖN	Polypeptide ID	Residues	Phosphorylation Sites	Glycosylation Sites		and Databases
41 (cont.)					Leucine zipper pattern: L315-L336	MOTIFS
42	1221545CD1	113	S66 S71 T44 T100 N88	N88	Signal_Peptide: M1-T45	SPSCAN
					Signal Peptide: M1-D18, M1-G20	HMMER
					Transmembrane Domain: T4-R24	TMAP
					N terminus cytosolic	
43	124737CD1	16	S30 S43	*		SPSCAN
						HMMER
4	1510784CD1	83	S33 S61 S71		Signal_Peptide: M1-S61	SPSCAN
						HMMER
				•	58-C67	BLIMPS_PRINTS
45	1901257CD1	128	S28 T31 T38		Signal_Peptide: M1-S27	SPSCAN
					M46-S74	HIMMER
						TMAP
					N terminus non-cytosolic	
94	2044370CDI	84	S31 S49 T22	N38		SPSCAN
					Signal Peptide: M1-G21	HMMER
					n: L3-Y23	TMAP
					nent) domain:	BLIMPS_PFAM
					PF00084A:C69-P73	
47	2820933CD1	109	S33 S52 S97			SPSCAN
					Signal Peptide: M1-S21, M1-E31	HMIMER
48	2902793CD1	159	S85 S154 T122		Signal_Peptide: M1-A23	SPSCAN
						HMMER
					ı: G54-C112	HIMIMER PFAIM
				,	Transmembrane Domain: V4-S21	TMAP
					N terminus non-cytosolic	

Table 3

SEQ ID Incyte	Incyte	Amino Acid Potential		Potential	Signature Sequences, Domains and Motifs	Analytical Methods
Ö	Polypeptide ID	Residues	Phosphorylation Sites	Glycosylation Sites		and Databases
49	7486536CD1	242	3100 S104	N76	Signal Peptide: M1-L19, M1-N21	HMMER
			S115 S139 S229			
			T30 T134 Y176			
					Transmembrane Domain: T6-L29	TMAP
					N terminus non-cytosolic	
50	8137305CD1	542	S74, S90, S95,	N41, N333	Signal cleavage: M1-F28	SPSCAN
			S111, S126, T225,			
			Y245, T231, S320,		-	
			S338, S379, S424,			
			T536			
					Signal Peptide: M1-C34	HMMER
					Transmembrane Domain: I7-L32;	TIMAP
					N-terminus is non-cytoplasmic	
51	3793128CD1	105	S30, T37	N55	Signal cleavage: M1-R22	SPSCAN
					Signal Peptide: M1-S19	HMMER
					Transmembrane Domain: P7-Y34;	TMAP
					N-terminus is non-cytoplasmic	
52	4001243CD1	102			Signal Peptide: M76-A95, M1-A27	HMMER
53	6986717CD1	129	S31, S77, T125	N83	Signal Peptide: M25-R48, M25-A53, M25-Q51	HMMER
					Transmembrane Domain: P4-R19;	TMAP
					N-terminus is non-cytoplasmic	

Table 3

Analytical Methods	and Databases	SPSCAN	HMMER	HIMMER PFAM				HMMER_PFAM	HMMER_PFAM	HMMER_PFAM	TIMHIMIMER			BLIMPS_PRODOM	
Signature Sequences, Domains and Motifs		signal_cleavage: M1-A33	Signal Peptide: M1-A33, M1-A34	S213-S236, K285-	S308, S333-S356, G384-K407, N93-S114, W69-P92,	K237-T260, S360-E383, P164-S187, S140-P163,	S189-N212, A261-Q284, H116-K136	Leucine rich repeat C-terminal domain: D417-D467	Leucine rich repeat N-terminal domain: P40-P67	globulin domain: G684-M742, T590-A651, 56	Cytosolic domain: Y793-S1070	Transmembrane domain: V770-I792	Non-cytosolic domain: M1-T769	RECEPTOR INTERLEUKIN-1 P	PD02870: F634-V666, L725-P759
Potential	Glycosylation Sites	N74 N150 N223 N269 N295 N661													
Potential	Phosphorylation Sites	S81 S247 S303 S333 S356 S380 S450 S565 S682 S711 S799 S827 S947 S964 S975 T192 T297 T347 T476 T515 T581 T586 T665 T713 T748 T795 T805 T825 T913 Y647													
Amino Acid Potential	Residues	1070													
Incyte	NO: Polypeptide ID	7503512CD1					-								
SEQ ID	ÄÖ:	54													

Table 3

SEQ ID	Incyte	Amino Acid	Acid Potential	Potential	Signature Sequences, Domains and Motifs	Analytical Methods
NO:	NO: Polypeptide ID Residues		Phosphorylation	Glycosylation		and Databases
			Sites	Sites		
54	!				MEMBRANE GLYCOPROTEIN	BLAST_PRODOM
(cont.)					PD129774: M742-S1070	
					PD172109: D468-F560	
					PD165826: E29-T70	
					KEK1 PRECURSOR T21D12.9 SPLICING SIGNAL BLAST_PRODOM	BLAST_PRODOM
					ALTERNATIVE KEK2	
					PD037237: L409-1587	
					IMMUNOGLOBULIN	BLAST_DOMO
					DM00001 P35918 668-745: L676-A751	
					DM00001 A46065 668-745; L676-A751	
					Leucine zipper pattern: L52-L73, L59-L80	MOTIFS

Polynucleotide SEO ID NO:/	Sequence Fragments
Incyte D/ Sequence Length	
55/095765CB1/1315	1-68, 1-341, 1-346, 1-359, 1-365, 1-367, 1-435, 1-566, 1-589, 1-625, 1-647, 2-367, 3-365, 3-367, 4-367, 5-367, 6-367, 7-68, 7-367, 9-367, 11-367, 67-367, 124-367, 244-367, 290-358, 361-547, 361-484, 363-484, 363-384, 363-394, 363-399, 363-400, 363-401, 363-411, 363-411, 363-411, 363-418, 363-424, 363-424, 363-424, 363-429, 363-460, 363-461, 363-411, 363-411, 363-417, 363-418, 363-428, 363-429, 363-460, 363-461, 363-411, 363-417, 363-418, 363-428, 363-429, 363-429, 363-490, 363-491, 363-492, 363-496, 363-497, 363-498, 363-401, 363-417, 363-418, 363-510, 363-510, 363-510, 363-501, 363-501, 363-501, 363-501, 363-501, 363-510, 363-510, 363-511, 363-512, 363-517, 363-518, 363-521, 363-521, 363-521, 363-527, 363-527, 363-528, 363-520, 363-521, 363-521, 363-521, 363-527, 363-527, 363-528, 363-520, 363-521, 363-521, 363-521, 363-527, 363-528, 363-520, 363-521, 36
	805-957, 813-957, 823-957, 836-957, 850-957, 854-957, 881-954, 911-956
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Table 4

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	634-1223, 644-926, 730-1013, 742-1003, 742-1013, 742-1055, 742-1300, 836-1135, 908-1128, 908-1377, 950-1192, 984-1199, 1055-
	1297, 1110-1390, 1117-1738, 1149-1753, 1171-1754, 1177-1663, 1181-1732, 1222-1733, 1302-1533, 1302-1754, 1302-1777, 1310-
	1758, 1324-1764, 1327-1622, 1333-1568, 1348-1755, 1454-1756, 1469-1696, 1511-1769, 1587-1765, 1590-1777, 1603-1765

Table 4

Folynucleonde	Sequence Fragments
Incyte ID/ Sequence Length	
104/8137305CB1/2587	1-308, 1-590, 1-899, 2-529, 5-481, 5-706, 27-611, 29-786, 30-611, 40-710, 41-276, 63-603, 87-873, 88-735, 89-688, 175-965, 309-609,
	410-925, 481-1028, 550-806, 642-912, 676-1032, 693-936, 693-1024, 813-1102, 875-1451, 1042-1605, 1069-1228, 1170-1692, 1185-
	1403, 1290-1512, 1317-1947, 1346-1907, 1346-1998, 1381-1968, 1384-2097, 1427-2080, 1431-1686, 1463-1947, 1471-1955, 1548-
	1845, 1548-1991, 1559-2178, 1560-2068, 1566-1851, 1585-2176, 1592-1909, 1592-2009, 1604-1834, 1611-2217, 1674-2067, 1683-
	2151, 1689-2119, 1725-2276, 1734-1978, 1734-2119, 1734-2257, 1734-2301, 1734-2307, 1734-2587, 1740-2195, 1761-2201, 1775-
	134, 1800-2211, 1802-2200
105/3793128CB1/1490	105/3793128CB1/1490 [1-278, 1-452, 1-531, 1-534, 1-547, 1-583, 1-625, 1-707, 7-752, 91-728, 103-724, 110-524, 110-554, 110-575, 110-651, 110-677, 110-
	.95, 110-739, 134-796, 193-576, 319-942, 435-1187, 436-970, 437-1067, 454-1190, 488-1163, 541-1134, 595-1270, 642-1173, 647-
	162, 661-1280, 668-1271, 685-1271, 703-1376, 743-1435, 753-1286, 753-1454, 840-1490, 887-1349, 915-1374, 959-1484, 962-1484,
	968-1437, 1005-1484, 1057-1484, 1073-1484, 1149-1484, 1177-1484, 1332-1484
106/4001243CB1/1174	106/4001243CB1/1174 1-292, 1-406, 1-442, 1-503, 1-570, 1-648, 52-303, 52-723, 86-644, 101-373, 101-583, 101-672, 113-321, 116-663, 285-854, 291-744,
	291-898, 305-810, 415-1174, 427-710, 430-945, 436-723, 488-963, 504-720, 695-963
107/6986717CB1/818	1-556, 1-611, 13-546, 83-818
108/7503512CB1/4717	1-4679, 1-4690, 124-403, 124-474, 124-511, 124-523, 124-529, 124-547, 124-576, 124-595, 124-599, 126-398, 132-583, 132-599, 142-
	594, 142-599, 155-583, 171-360, 197-433, 242-482, 252-357, 252-360, 257-561, 366-478, 396-946, 397-946, 620-1172, 628-1180, 744-
	1413, 764-1400, 848-1500, 876-1270, 877-1423, 904-1260, 967-1454, 1032-1696, 1038-1658, 1045-1432, 1079-1357, 1107-1361,
	1116-1615, 1146-1396, 1147-1749, 1148-1397, 1152-1814, 1181-1742, 1197-1745, 1204-1439, 1224-1806, 1347-1811, 1399-1968,
	1418-1492, 1425-1950, 1425-1977, 1427-2000, 1443-1622, 1461-1742, 1496-2145, 1678-2162, 1769-2318, 1809-2230, 2031-2303,
	:773-3462, 2797-3170, 2799-3170, 2807-3484, 2820-3351, 2824-3468, 2935-3513, 2955-3204, 2968-3205, 2982-3216, 2986-3262,
	2999-3194, 3008-3212, 3011-3293, 3019-3399, 3020-3261, 3022-3553, 3045-3499, 3057-3500, 3059-3500, 3072-3371, 3077-3291,
	3084-3494, 3086-3372, 3093-3572, 3096-3532, 3110-3434, 3112-3373, 3120-3468, 3139-3605, 3152-3217, 3166-3290, 3216-3525,
	3252-3499, 3276-3508, 3276-3880, 3279-3861, 3326-3572, 3331-3592, 3331-3596, 3340-3536, 3340-3594, 3340-3601, 3355-3620,
•	3357-3609, 3362-3693, 3379-4037, 3382-4151, 3383-3644, 3383-3669, 3384-3658, 3391-3660, 3396-3913, 3403-3651, 3404-3662,

Polynucleotide	Sequence Fragments
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Incyte ID/ Sequence Length	
108 (cont.)	3404-3680, 3412-3676, 3433-3706, 3434-3728, 3434-3886, 3435-3649, 3443-3683, 3444-3742, 3456-3855, 3458-3694, 3458-3706,
	3458-4047, 3459-3683, 3465-3750, 3477-3717, 3479-3708, 3491-4005, 3499-4117, 3501-3784, 3501-4125, 3513-3732, 3513-3737,
	3515-3843, 3516-3806, 3528-3606, 3539-3830, 3542-3627, 3542-3807, 3542-3824, 3542-3882, 3542-4149, 3548-3744, 3548-3782,
	3548-3808, 3568-3922, 3568-3978, 3571-4320, 3582-4060, 3586-3868, 3586-4184, 3588-4068, 3590-4295, 3600-3808, 3600-4261,
	3603-3884, 3603-4075, 3603-4133, 3614-3807, 3621-3829, 3637-4192, 3638-3878, 3638-3895, 3640-3852, 3650-3933, 3654-4176,
	3656-4212, 3659-3909, 3660-3950, 3667-3940, 3669-3925, 3669-4234, 3671-4053, 3671-4065, 3681-3901, 3707-4013, 3713-4166,
	3714-3975, 3715-3971, 3717-3974, 3720-3873, 3721-3965, 3737-4296, 3738-4125, 3747-4032, 3756-4373, 3770-4118, 3772-4050,
	3773-4212, 3775-4038, 3802-4095, 3808-4032, 3809-4390, 3813-4315, 3815-4059, 3815-4496, 3825-4084, 3825-4119, 3825-4120,
	3828-4318, 3828-4462, 3829-4016, 3829-4043, 3831-4101, 3832-4343, 3838-4100, 3841-4091, 3847-4480, 3858-4082, 3860-4236,
	3860-4303, 3866-4134, 3871-4132, 3874-4634, 3876-4187, 3877-4132, 3878-4164, 3892-4110, 3896-4608, 3901-4188, 3904-4118,
	3904-4121, 3904-4122, 3904-4140, 3904-4305, 3904-4320, 3907-4157, 3908-4200, 3915-4171, 3920-4122, 3920-4345, 3922-4167,
	3930-4225, 3931-4147, 3931-4186, 3931-4195, 3931-4201, 3931-4202, 3933-4165, 3934-4025, 3938-4147, 3938-4374, 3940-4201,
	3942-4245, 3944-4238, 3947-4218, 3953-4561, 3956-4331, 3963-4628, 3967-4223, 3981-4265, 3982-4247, 3988-4226, 3988-4635,
	3989-4545, 3990-4247, 3998-4211, 3998-4298, 3998-4673, 4000-4304, 4000-4606, 4003-4265, 4003-4267, 4003-4285, 4003-4536,
	4003-4664, 4007-4230, 4013-4306, 4019-4552, 4025-4587, 4025-4631, 4025-4663, 4027-4289, 4031-4288, 4031-4310, 4031-4676,
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	4142-4697, 4144-4693, 4152-4486, 4170-4679, 4172-4711, 4174-4376, 4181-4678, 4189-4671, 4190-4459, 4190-4665, 4190-4678,

Table 4

Polynucleotide	Sequence Fragments
SEQ ID NO:/	
Incyte ID/ Sequence Length	
108 (cont.)	4190-4687, 4191-4363, 4192-4458, 4193-4439, 4197-4630, 4197-4702, 4200-4636, 4203-4685, 4206-4672, 4207-4678, 4212-4490,
	4223-4686, 4230-4678, 4231-4676, 4231-4680, 4233-4678, 4235-4678, 4236-4682, 4239-4678, 4243-4619, 4244-4682, 4246-4586,
	4250-4678, 4250-4692, 4251-4696, 4252-4671, 4254-4717, 4258-4678, 4260-4680, 4261-4672, 4261-4678, 4265-4684, 4266-4680,
	4269-4684, 4269-4703, 4270-4684, 4270-4707, 4271-4671, 4271-4678, 4272-4678, 4274-4678, 4275-4678, 4276-4687, 4281-4678,
	4285-4678, 4287-4681, 4289-4529, 4293-4513, 4293-4613, 4293-4641, 4293-4665, 4293-4674, 4295-4541, 4298-4581, 4306-4682,
	4307-4678, 4312-4684, 4315-4558, 4319-4679, 4322-4678, 4322-4681, 4323-4597, 4327-4669, 4328-4678, 4331-4678, 4332-4682,
	4334-4678, 4344-4597, 4351-4672, 4354-4678, 4355-4581, 4356-4625, 4358-4679, 4359-4678, 4360-4672, 4366-4580, 4366-4668,
	4366-4669, 4366-4676, 4369-4655, 4375-4685, 4377-4478, 4377-4663, 4382-4659, 4387-4677, 4389-4668, 4393-4679, 4407-4644,
	4429-4705, 4434-4679, 4459-4679, 4467-4678, 4469-4683, 4480-4708, 4482-4643, 4482-4681, 4482-4709, 4486-4575, 4499-4678,
	4501-4679, 4515-4702, 4518-4662, 4535-4678, 4541-4672, 4556-4678, 4571-4678, 4573-4678, 4595-4672, 4595-4709, 4609-4678

Table 5

Polynucleotide SEC	Incyte Project ID:	Representative Library
55	095765CB1	PITUNOT06
56	6399886CB1	PANCTUT01
57 ·	6024420CB1	TESTNOT11
58	7481067CB1	BRSTNOT07
59	3378720CB1	KERANOT02
60	938824CB1	CERVNOT01
61	1683721CB1	PROSNOT15
62	1694122CB1	COLNNOT23
63	1970615CB1	PROSTUT09
64	2314152CB1	CONUTUT01
65	2886225CB1	UTRSTMR02
66	6144418CB1	BRANDIN01
67	6834184CB1	BRSTNON02
68	6951005CB1	BRAITDR02
69	7250331CB1	KIDNTUT15
71	7011042CB1	BRAZNOT01
72	7427362CB1	BRSTTMR01
73	7485304CB1	SEMVTDE01
74	1422394CB1	MIXDUNB01
75	1336022CB1	COLNNOT13
76	7473674CB1	LUNGNON07
78	7475860CB1	ADRENON04
79	7950941CB1	BRABNOE02
80	7485334CB1	
81	7220001CB1	BSTMNON02 COLXTDT01
82	5956275CB1	BRAUNOR01
83	346472CB1	THYMNOT02
84	643526CB1	BRAIFEE05
85	1483418CB1	SINTBST01
86	2683477CB1	SINIUCT01
87	5580991CB1	UTRENON03
88	5605931CB1	MONOTXN03
89	6975241CB1	BRAHTDR04
90	6988529CB1	BRAIFER05
91	6996808CB1	BRAXTDR17
92	7472689CB1	LNODNOT12
93	876751CB1	THYRNOT03
	2512510CB1	
94 96		BRAUNOR01
97	1221545CB1	NEUTFMT01
	124737CB1	THYMNON04
98 99	1510784CB1	SINTFER03
	1901257CB1	BRSTTMT02
100	2044370CB1	HIPONON02
101	2820933CB1	ADRETUT06
102	2902793CB1	DRGCNOT01
103	7486536CB1	BRSTNOT05
104	8137305CB1	MIXIDTUE01

Table 5

Polynucleotide SEQ ID NO:	Incyte Project ID:	Representative Library
105	3793128CB1	BRSTNOT28
106	4001243CB1	PROSTMT03
107	6986717CB1	BRAIFER05
108	7503512CB1	BRSTNOT01

ADRENON04	Vector PSPORT1	Library Description Normalized library was constructed from 1.36 x 1e6 independent clones from an adrenal tissue library. Starting RNA was made from adrenal gland tissue removed from a 20-year-old Caucasian male, who died from head trauma. The library was normalized in two rounds using conditions adapted from Soares et al. (PNAS (1994) 91.9228-9232) and Bonaldo et al. (Genome Res (1996) 6: 791-806), using a significantly longer (48-hours/round) reannealing hybridization period.
ADRETUT06	pINCY	Library was constructed using RNA isolated from adrenal tumor tissue removed from a 57-year-old Caucasian female during a unilateral right adrenalectomy. Pathology indicated pheochromocytoma, forming a nodular mass completely replacing the medulla of the adrenal gland.
BRABNOE02	PBK-CMV	This 5' biased random primed library was constructed using RNA isolated from vermis tissue removed from a 35-year-old Caucasian male who died from cardiac failure. Pathology indicated moderate leptomeningeal fibrosis and multiple microinfarctions of the cerebral neocortex. Patient history included dilated cardiomyopathy, congestive heart failure, cardiomegaly, and an enlarged spleen and liver. Patient medications included simethicone, Lasix, Digoxin, Colace, Zantac, captopril, and Vasotec.
BRAHTDR04	PCDNA2.1	This random primed library was constructed using RNA isolated archaecortex, anterior hippocampus tissue removed from a 55-year-old Caucasian female who died from cholangiocarcinoma. Pathology indicated mild meningeal fibrosis predominately over the convexities, scattered axonal spheroids in the white matter of the cingulate cortex and the thalamus, and a few scattered neurofibrillary tangles in the entorhinal cortex and the periaqueductal gray region. Pathology for the associated tumor tissue indicated well-differentiated cholangiocarcinoma of the liver with residual or relapsed tumor. Patient history included cholangiocarcinoma, post-operative Budd-Chiari syndrome, biliary ascites, hydrothorax, dehydration, malnutrition, oliguria and acute renal failure. Previous surgeries included cholecystectomy and resection of 85% of the liver.
BRAIFEE05	PCDNA2.1	This 5' biased random primed library was constructed using RNA isolated from brain tissue removed from a Caucasian male fetus who was stillborn with a hypoplastic left heart at 23 weeks' gestation.
BRAIFER05	pINCY	Library was constructed using RNA isolated from brain tissue removed from a Caucasian male fetus who was stillborn with a hypoplastic left heart at 23 weeks' gestation.
BRAITDR02	PCDNA2.1	This random primed library was constructed using RNA isolated from allocortex, neocortex, anterior and frontal cingulate tissue removed from a 55-year-old Caucasian female who died from cholangiocarcinoma. Pathology indicated mild meningeal fibrosis predominately over the convexities, scattered axonal spheroids in the white matter of the cingulate cortex and the thalamus, and a few scattered neurofibrillary tangles in the entorhinal cortex and the periaqueductal gray region. Pathology for the associated tumor tissue indicated well-differentiated cholangiocarcinoma of the liver with residual or relapsed tumor. Patient history included cholangiocarcinoma, post-operative Budd-Chiari syndrome, biliary ascites, hydrothorax, dehydration, malnutrition, oliguria and acute renal failure. Previous surgeries included cholecystectomy and resection of 85% of the liver.

Library	Vector	Library Description
BRANDIN01	pINCY	This normalized pineal gland tissue library was constructed from .4 million independent clones from a pineal gland tissue library from two different donors. Starting RNA was made from pooled pineal gland tissue removed from two Caucasian females: a 68-year-old (donor A) who died from congestive heart failure and a 79-year-old (donor B) who died from pneumonia. Neuropathology for donor A indicated mild to moderate Alzheimer disease, atherosclerosis, and multiple infarctions. Neuropathology for donor B indicated severe Alzheimer disease, arteriolosclerosis, cerebral amyloid angiopathy and multiple infarctions. There were diffuse and neuritic amyloid plaques and neurofibrillary tangles throughout the brain sections examined in both donors. Patient history included diabetes mellitus, rheumatoid arthritis, hyperthyroidism, amyloid heart disease, and dementia in donor A; and pseudophakia, gastritis with bleeding, glaucoma, peripheral vascular disease, COPD, delayed onset tonic/clonic seizures, and transient ischemic attack in donor B. The library was normalized in one round using conditions adapted from Soares et al., PNAS (1994) 91:9228-9232 and Bonaldo et al., Genome Research 6 (1996):791, except that a significantly longer (48 hours/round) reannealing hybridization was used.
BRAUNOR01	pINCY	This random primed library was constructed using RNA isolated from striatum, globus pallidus and posterior putamen tissue removed from an 81-year-old Caucasian female who died from a hemorrhage and ruptured thoracic aorta due to atherosclerosis. Pathology indicated moderate atherosclerosis involving the internal carotids, bilaterally; microscopic infarcts of the frontal cortex and hippocampus; and scattered diffuse amyloid plaques and neurofibrillary tangles, consistent with age. Grossly, the leptomeninges showed only mild thickening and hyalinization along the superior sagittal sinus. The remainder of the leptomeninges was thin and contained some congested blood vessels. Mild atrophy was found mostly in the frontal poles and lobes, and temporal lobes, bilaterally. Microscopically, there were pairs of Alzheimer type II astrocytes within the deep layers of the neocortex. There was increased satellitosis around neurons in the deep gray matter in the middle frontal cortex. The amygdala contained rare diffuse plaques and neurofibrillary tangles. The posterior hippocampus contained a microscopic area of cystic cavitation with hemosiderin-laden macrophages surrounded by reactive gliosis. Patient history included sepsis, cholangitis, post-operative atelectasis, pneumonia CAD, cardiomegaly due to left ventricular hypertrophy, splenomegaly, arteriolonephrosclerosis, nodular colloidal goiter, emphysema, CHF, hypothyroidism, and peripheral vascular disease.
BRAXTDR17	PCDNA2.1	This random primed library was constructed using RNA isolated from temporal neocortex tissue removed from a 55-year-old Caucasian female who died from cholangiocarcinoma. Pathology indicated mild meningeal fibrosis predominately over the convexities, scattered axonal spheroids in the white matter of the cingulate cortex and the thalamus, and a few scattered neurofibrillary tangles in the entorthinal cortex and the periaqueductal gray region. Pathology for the associated tumor tissue indicated well-differentiated cholangiocarcinoma of the liver with residual or relapsed tumor. Patient history included cholangiocarcinoma, post-operative Budd-Chiari syndrome, biliary ascites, hydrothorax, dehydration, malnutrition, oliguria and acute renal failure. Previous surgeries included cholecystectomy and resection of 85% of the liver.

Library	Vector	Titran Daconintina
BRAZNOTOI	pINCY	Library was constructed using RNA isolated from striatum, globus pallidus and posterior putamen tissue removed from a 45-year-old Caucasian female who died from a dissecting aortic aneurysm and ischemic bowel disease. Pathology indicated mild arteriosclerosis involving the cerebral cortical white matter and basal ganglia. Grossly, there was mild meningeal fibrosis and mild focal atherosclerotic plaque in the middle cerebral artery, as well as vertebral arteries bilaterally. Microscopically, the cerebral hemispheres, brain stem and cerebellum reveal focal areas in the white matter that contain blood vessels that were barrel-shaped, hyalinized, with hemosiderin-laden macrophages in the Virchow-Robin space. In addition, there were scattered neurofibrillary tangles within the basolateral nuclei of the amygdala. Patient history included mild atheromatosis of aorta and coronary arteries, bowel and liver infarct due to aneurysm, physiologic fatty liver associated with obesity, mild diffuse emphysema, thrombosis of mesenteric and portal veins, cardiomegaly due to hypertrophy of left ventricle, arterial hypertension, acute pulmonary edema, splenomegaly, obesity (300 lb.), leiomyoma of uterus, sleep apnea, and iron deficiency anemia.
BRSTNON02	pINCY	This normalized breast tissue library was constructed from 6.2 million independent clones from a pool of two libraries from two different donors. Starting RNA was made from breast tissue removed from a 46-year-old Caucasian female during a bilateral reduction mammoplasty (donor A), and from breast tissue removed from a 60-year-old Caucasian female during a bilateral reduction mammoplasty (donor B). Pathology indicated normal breast parenchyma, bilaterally (A) and bilateral mammary hypertrophy (B). Patient history included hypertrophy of breast, obesity, lumbago, and glaucoma (A) and joint pain in the shoulder, thyroid cyst, colon cancer, normal delivery and cervical cancer (B). Family history included cataract, osteoarthritis, uterine cancer, benign hypertension, hyperlipidemia, and alcoholic cirrhosis of the liver, cerebrovascular disease, and type II diabetes (A) and cerebrovascular accident, atherosclerotic coronary artery disease, colon cancer, type II diabetes, hyperlipidemia, depressive disorder, and Alzheimer's Disease. The library was normalized in two rounds using conditions adapted from Soares et al., PNAS (1994) 91:9228-9232 and Bonaldo et al., Genome Research 6 (1996):791, except that a significantly longer (48 hours/round) reannealing hybridization was used.
BRSTNOT01	PBLUESCRIPT	Library was constructed using RNA isolated from the breast tissue of a 56-year-old Caucasian female who died in a motor vehicle accident.
BRSTNOT05	PSPORT1	Library was constructed using RNA isolated from breast tissue removed from a 58-year-old Caucasian female during a unilateral extended simple mastectomy. Pathology for the associated tumor tissue indicated multicentric invasive grade 4 lobular carcinoma. Patient history included skin cancer, rheumatic heart disease, osteoarthritis, and tuberculosis. Family history included cerebrovascular and cardiovascular disease, breast and prostate cancer, and type I diabetes.
BRSTNOT07	pINCY	Library was constructed using RNA isolated from diseased breast tissue removed from a 43-year-old Caucasian female during a unilateral extended simple mastectomy. Pathology indicated mildly proliferative fibrocystic changes with epithelial hyperplasia, papillomatosis, and duct ectasia. Pathology for the associated tumor tissue indicated invasive grade 4, nuclear grade 3 mammary adenocarcinoma with extensive comedo necrosis. Family history included epilepsy, cardiovascular disease, and type II diabetes.

Library	Vector	Library Description
BRSTNOT28	pINCY	Library was constructed using RNA isolated from diseased right breast tissue removed from a 40-year-old Caucasian female during a bilateral reduction mammoplasty. Pathology indicated bilateral mild fibrocystic and proliferative changes. Patient history included pure hypercholesterolemia. Family history included acute myocardial infarction, atherosclerotic coronary artery disease, type II diabetes, and prostate cancer.
BRSTTMR01	PCDNA2.1	This random primed library was constructed using RNA isolated from right breast tissue removed from a 62-year-old Caucasian female during open breast biopsy and unilateral extended simple mastectomy. Pathology indicated benign breast parenchyma. Pathology for the matched tumor tissue indicated residual grade 3 (of 4) ductal adenocarcinoma. The patient presented with breast cancer. Patient history included benign neoplasm of the large bowel and leg vein occlusion. Previous surgeries included dilation and curettage and spinal canal exploration. Patient medications included Lozal, Mevacor, and tamoxifen. Family history included heart murmur in the mother; skin cancer in the sibling(s); and prostate cancer in the grandfather.
BRSTTMT02	pINCY	Library was constructed using RNA isolated from diseased right breast tissue removed from a 46-year-old Caucasian female during a unilateral extended simple mastectomy and open breast biopsy. Pathology indicated mildly proliferative fibrocystic change, including intraductal duct ectasia, papilloma formation, and ductal hyperplasia. Pathology for the associated tumor tissue indicated multifocal ductal carcinoma in situ, both comedo and non-comedo types, nuclear grade 2 with extensive intraductal calcifications. Patient history included deficiency anemia, normal delivery, chronic sinusitis, extrinsic asthma, and kidney infection. Family history included type II diabetes, benign hypertension, cerebrovascular disease, skin cancer, and hyperlipidemia.
BSTMNON02	PSPORTI	This normalized brain stem library was constructed from 2.84 million independent clones from a brain stem library. Starting RNA was made from the brain stem tissue of a 72-year-old Caucasian male who died from myocardial infarction. Patient history included coronary artery disease, insulin-dependent diabetes mellitus, and arthritis. Normalization and hybridization conditions were adapted from Soares et al. (PNAS (1994) 91:9228).
CERVNOT01	PSPORT1	Library was constructed using RNA isolated from the uterine cervical tissue of a 35-year-old Caucasian female during a vaginal hysterectomy with dilation and curettage. Pathology indicated mild chronic cervicitis. Family history included atherosclerotic coronary artery disease and type II diabetes.
COLNNOT13	pINCY	Library was constructed using RNA isolated from ascending colon tissue of a 28-year-old Caucasian male with moderate chronic ulcerative colitis.
COLNNOT23	pINCY	Library was constructed using RNA isolated from diseased colon tissue removed from a 16-year-old Caucasian male during a total colectomy with abdominal/perineal resection. Pathology indicated gastritis and pancolonitis consistent with the acute phase of ulcerative colitis. Inflammation was more severe in the transverse colon, with inflammation confined to the mucosa. There was only mild involvement of the ascending and sigmoid colon, and no significant involvement of the cecum, rectum, or terminal ileum. Family history included irritable bowel syndrome.

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Library	Vector	Library Description
COLXIDT01	pINCY	Library was constructed using RNA isolated from colon tissue removed from the appendix of a 37-year-old Black female during myomectomy, dilation and curettage, right fimbrial region biopsy, and incidental appendectomy. Pathology indicated an unremarkable appendix. Pathology for the associated tumor tissue indicated multiple (12) uterine leiomyomata. Patient history included premenopausal menorrhagia and sarcoidosis of the lung. Family history included acute myocardial infarction and atherosclerotic coronary artery disease.
CONUTUT01	pINCY	Library was constructed using RNA isolated from sigmoid mesentery tumor tissue obtained from a 61-year-old female during a total abdominal hysterectomy and bilateral salpingo-oophorectomy with regional lymph node excision. Pathology indicated a metastatic grade 4 malignant mixed mullerian tumor present in the sigmoid mesentery at two sites.
DRGCNOT01	pINCY	Library was constructed using RNA isolated from dorsal root ganglion tissue removed from the cervical spine of a 32-year-old Caucasian male who died from acute pulmonary edema and bronchopneumonia, bilateral pleural and pericardial effusions, and malignant lymphoma (natural killer cell type). Patient history included probable cytomegalovirus infection, hepatic congestion and steatosis, splenomegaly, hemorrhagic cystitis, thyroid hemorrhage, and Bell's palsy. Surgeries included colonoscopy, large intestine biopsy, adenotonsillectomy, and nasopharyngeal endoscopy and biopsy; treatment included radiation therapy.
HIPONON02	PSPORT1	This normalized hippocampus library was constructed from 1.13M independent clones from a hippocampus tissue library. RNA was isolated from the hippocampus tissue of a 72-year-old Caucasian female who died from an intracranial bleed. Patient history included nose cancer, hypertension, and arthritis. The normalization and hybridization conditions were adapted from Soares et al. (PNAS (1994) 91:9228).
KERANOT02	PSPORT1	Library was constructed using RNA isolated from epidermal breast keratinocytes (NHEK). NHEK (Clontech #CC-2501) is human breast keratinocyte cell line derived from a 30-year-old black female during breast-reduction surgery.
KIDNTUT15	pINCY	Library was constructed using RNA isolated from left kidney tumor tissue removed from a 65-year-old Caucasian male during an exploratory laparotomy and nephroureterectomy. Pathology indicated grade 1 renal cell carcinoma within the upper pole of the left kidney. Patient history included malignant melanoma of the abdominal skin, benign neoplasm of colon, cerebrovascular disease, and umbilical hernia. Family history included myocardial infarction, atherosclerotic coronary artery disease, and cerebrovascular disease, and prostate cancer.
LNODNOT12	pINCY	Library was constructed using RNA isolated from lymph node tissue obtained from an 11-year-old Caucasian female who died from a motor vehicle accident. Previous surgeries included tonsilectomy.
LUNGNON07	pINCY	This normalized lung tissue library was constructed from 5.1 million independent clones from a lung tissue library. Starting RNA was made from RNA isolated from lung tissue. The library was normalized in two rounds using conditions adapted from Soares et al., PNAS (1994) 91:9228-9232 and Bonaldo et al., Genome Research (1996) 6:791, except that a significantly longer (48 hours/round) reannealing hybridization was used.

Library	Vector	Library Description
MIXDTUE01	РВК-СМV	This 5' biased random primed library was constructed using pooled cDNA from seven donors. cDNA was generated using mRNA isolated from placental tissue removed from a Caucasian fetus (A), who died after 16 weeks' gestation from fetal demise and hydrocephalus; from placental tissue removed from a Caucasian male fetus (B), who died after 18 weeks' gestation from fetal demise; from an untreated LNCaP cell line, derived from prostate carcinoma with metastasis to the left supraclavicular lymph nodes, removed from a 50-year-old Caucasian male (C); from endometrial tissue removed from a 32-year-old female (D); from diseased right ovary tissue removed from a 45-year-old Caucasian female (donor F) and from right fallopian tube tumor tissue removed from an 85-year-old Caucasian female (donor G). For donor F) and from right fallopian tube tumor tissue removed from an 85-year-old Caucasian female (donor G). For donor A, patient history included umbilical cord wrapped around the head (3 times) and the shoulders (1 time). Serology was positive for anti-CMV. Family history included multiple pregnancies and live births, and an abortion in the mother. For donor B, serologies were negative. For donor D, pathology indicated the endometrium was in secretory phase. For donor E, pathology indicated poorly differentiated mixed endometrioid (80%) and serous (20%) adenocarcinoma of the right fallopian tube. Patient history included medullary carcinoma of the thyroid.
MIXDUNB01	pINCY	Library was constructed using RNA isolated from myometrium removed from a 41-year-old Caucasian female (A) during vaginal hysterectomy with a dilatation and curettage and untreated smooth muscle cells removed from the renal vein of a 57-year-old Caucasian male. Pathology for donor A indicated the myometrium and cervix were unremarkable. The endometrium was secretory and contained fragments of endometrial polyps. Benign endo- and ectocervical mucosa were identified in the endocervix. Pathology for the associated tumor tissue indicated uterine leiomyoma. Medical history included an unspecified menstrual disorder, ventral hernia, normal delivery, a benign ovarian neoplasm, and tobacco abuse in donor A. Previous surgeries included a bilateral destruction of fallopian tubes, removal of a solitary ovary, and an exploratory laparotomy in donor A. Medications included ferrous sulfate in donor A.
MONOTXN03	pINCY	Normalized, treated monocyte tissue library was constructed from 7.6 million independent clones from a treated monocyte library. Starting RNA was made from RNA isolated from treated monocytes from peripheral blood obtained from a 42-year-old female. The cells were treated with anti-interleukin-10 (anti-LI-10) and lipopolysaccharide (LPS). The anti-LI-10 was added at time 0 at 10 ng/ml and LPS was added at 1 hour at 5ng/ml. The monocytes were isolated from buffy coat by adherence to plastic. Incubation time was 24 hours. cDNA synthesis was initiated using a Notl-anchored oligo(dT) primer. The libraries were normalized in two rounds using conditions adapted from Soares et al., PNAS (1994) 91:9228 and Bonaldo et al., Genome Research (1996 6):791, except that a significantly longer (48 -hours/round) reannealing hybridization was used. The libraries were then linearized and recircularized to select for insert containing clones as follows: plasmid DNA was prepped from approximately 1 million clones from the normalized, treated monocyte tissue libraries following soft agar transformation. The DNA was linearized with Notl and insert containing clones were size-selected by agarose gel electrophoresis and recircularized by ligation.

Library	Vector	Library Description
NEUTFMT01	PBLUESCRIPT	Library was constructed using total RNA isolated from peripheral blood granulocytes collected by density gradient centrifugation through Ficoll-Hypaque. The cells were isolated from buffy coat units obtained from unrelated male and female
		donors. Cells were cultured in 10 nM fMLP for 30 minutes, lysed in GuSCN, and spun through CsCl to obtain RNA for library
		coust uction. Decause this initially was made from total rates, it has an unusually inguished to mague singleton sequences, which may not all come from polyA RNA species.
PANCTUT01	pINCY	Library was constructed using RNA isolated from pancreatic tumor tissue removed from a 65-year-old Caucasian female during
		radical subtotal pancreatectomy. Pathology indicated an invasive grade 2 adenocarcinoma. Patient history included type II
		diabetes, osteoarthritis, cardiovascular disease, benign neoplasm in the large bowel, and a cataract. Previous surgeries included a
		total spienectomy, cholecystectomy, and abdominal bysterectomy. Family history included cardiovascular disease, type il diabetes, and stomach cancer.
PITUNOT06	pINCY	Library was constructed using RNA isolated from pituitary gland tissue removed from a 55-year-old male who died from
es en constant		chronic obstructive pulmonary disease. Neuropathology indicated there were no gross abnormalities, other than mild ventricular
		enlargement. There was no apparent microscopic abnormality in any of the neocortical areas examined, except for a number of
		silver positive neurons with apical dendrite staining, particularly in the frontal lobe. The significance of this was undetermined.
		The only other microscopic abnormality was that there was prominent silver staining with some swollen axons in the CA3
		region of the anterior and posterior hippocampus. Microscopic sections of the cerebellum revealed mild Bergmann's gliosis in
		the Purkinje celi layer. Patient fusiory included schizophrenia.
PROSNOT15	pINCY	Library was constructed using RNA isolated from diseased prostate tissue removed from a 66-year-old Caucasian male during
		radical prostatectomy and regional lymph node excision. Pathology indicated adenofibromatous hyperplasia. Pathology for the
		associated tumor tissue indicated an adenocarcinoma (Gleason grade 2+3). The patient presented with elevated prostate specific
		antigen (PSA). Family history included prostate cancer, secondary bone cancer, and benign hypertension.
PROSTMT03	pINCY	The library was constructed using RNA isolated from right prostate tissue removed from a 68-year-old Caucasian male during a
		radical prostatectomy and regional lymph node excision. Pathology for the associated tumor indicated adenocarcinoma, Gleason
		grade 4+3, which formed a predominant mass involving the left side peripherally. The patient presented with elevated prostate
		specific antigen (PSA) and induration. Patient history included pure hypercholesterolemia, kidney calculus, an unspecified
		allergy, and atopicdermatitis. Family history included colon cancer.
PROSTUT09	pINCY	Library was constructed using RNA isolated from prostate tumor tissue removed from a 66-year-old Caucasian male during a
		radical prostatectomy, radical cystectomy, and urinary diversion. Pathology indicated grade 3 transitional cell carcinoma. The
		patient presented with prostatic inflammatory disease. Patient history included lung neoplasm, and benign hypertension. Family
		history included a malignant breast neoplasm, tuberculosis, cerebrovascular disease, atherosclerotic coronary artery disease and
		lung cancer.

Library	Vector	Library Description
SEMVTDE01	PCDNA2.1	This S' biased random primed library was constructed using RNA isolated from seminal vesicle tissue removed from a 63-year-old Caucasian male during closed prostatic biopsy, radical prostatectomy, and regional lymph node excision. Pathology for the associated tumor tissue indicated Gleason grade 2+3 adenocarcinoma in the right side of the prostate. Adenofibromatous hyperplasia was present. The patient presented with prostate cancer, elevated prostate specific antigen and prostatic hyperplasia. Patient history included kidney calculus, extrinsic asthma, benign bowel neoplasm, backache, tremor, and tobacco abuse in remission. Previous surgeries included adenotonsillectomy. Patient medications included Ventolin and Vanceril. Family history included atherosclerotic coronary artery disease and acute myocardial infarction in the father; and stomach cancer and extrinsic asthma in the grandparent(s).
SINIUCT01	pINCY	Library was constructed using RNA isolated from ileum tissue obtained from a 42-year-old Caucasian male during a total intra- abdominal colectomy and endoscopic jejunostomy. Previous surgeries included polypectomy, colonoscopy, and spinal canal exploration. Family history included cerebrovascular disease, benign hypertension, atherosclerotic coronary artery disease, and type II diabetes.
SINTBST01	pINCY	Library was constructed using RNA isolated from ileum tissue obtained from an 18-year-old Caucasian female during bowel anastomosis. Pathology indicated Crohn's disease of the ileum, involving 15 cm of the small bowel. Family history included cerebrovascular disease and atherosclerotic coronary artery disease.
SINTFER03	PCDNA2.1	This random primed library was constructed using RNA isolated from small intestine tissue removed from a Caucasian male fetus who died from fetal demise.
TESTNOT11	pINCY	Library was constructed using RNA isolated from testicular tissue removed from a 16-year-old Caucasian male who died from hanging. Patient history included drug use (tobacco, marijuana, and cocaine use), and medications included Lithium, Ritalin, and Paxil.
THYMNON04	PSPORT1	This normalized library was constructed from a thymus tissue library. Starting RNA was made from thymus tissue removed from a 3-year-old Caucasian male, who died from anoxia. Serologies were negative. The patient was not taking any medications. The library was normalized in two rounds using conditions adapted from Soares et al., PNAS (1994) 91:9228 and Bonaldo et al., Genome Research (1996) 6:791, except that a significantly longer (48-hours/round) reannealing hybridization was used.
THYMINOT02	PBLUESCRIPT	Library was constructed using polyA RNA isolated from thymus tissue removed from a 3-year-old Caucasian male, who died from drowning. Serologies were negative.
THYRNOT03	pINCY	Library was constructed using RNA isolated from thyroid tissue removed from the left thyroid of a 28-year-old Caucasian female during a complete thyroidectomy. Pathology indicated a small nodule of adenomatous hyperplasia present in the left thyroid. Pathology for the associated tumor tissue indicated dominant follicular adenoma, forming a well-encapsulated mass in the left thyroid.

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Library	Vector	Library Description	_
UTRENON03 pINCY	pINCY	This normalized library was constructed from 1.2 x 10e7 independent clones from a uterine endometrial tissue library. Starting RNA was made from uterine endometrium tissue obtained from a 29-year-old Caucasian female during a vaginal hysterectomy and cystocele repair. Pathology indicated the endometrium was secretory and the cervix showed mild chronic cervicitis with	
		included hypothyroidism, Pervious surgeries included a normal delivery, a laminectomy, and a rhinoplasty. Family history	
		included benign hypertension, type II diabetes, and hyperlipidemia. The libraries were normalized in two rounds using conditions adapted from Soares et al., PNAS (1994) 91:9228 and Bonaldo et al., Genome Research (1996) 6:791, except that a significantly longer (48 hours/round) reannealing hybridization was used.	
UTRSTMR02 PCDNA2.1	PCDNA2.1	This random primed library was constructed using pooled cDNA from two different donors. cDNA was generated using mRNA isolated from endometrial tissue removed from a 32-year-old female (donor A) and using mRNA isolated from myometrium	
		removed from a 45-year-old female (donor B) during vaginal hysterectomy and bilateral salpingo-oophorectomy. In donor A, pathology indicated the endometrium was secretory phase. The cervix showed severe dysplasia (CIN III) focally involving the	
		squamocolumnar junction at the 1, 6 and 7 o'clock positions. Mild koilocytotic dysplasia was also identified within the cervix. In donor B, pathology for the matched tumor tissue indicated multiple (23) subserosal, intramural, and submucosal leiomyomata.	
		Patient history included stress incontinence, extrinsic asthma without status asthmaticus and normal delivery in donor B. Family	
		history included cerebrovascular disease, depression, and atherosclerotic coronary artery disease in donor B.	_

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Program	Description	Reference	Parameter Threshold
ProfileScan	An algorithm that searches for structural and sequence motifs in protein sequences that match sequence patterns defined in Prosite.	Gribskov, M. et al. (1988) CABIOS 4:61-66; Gribskov, M. et al. (1989) Methods Enzymol. 183:146-159; Bairoch, A. et al. (1997) Nucleic Acids Res. 25:217-221.	Normalized quality score>GCG-specified 'HIGH" value for that particular Prosite motif. Generally, score=1.4-2.1.
Phred	A base-calling algorithm that examines automated sequencer traces with high sensitivity and probability.	Ewing, B. et al. (1998) Genome Res. 8:175-185; Ewing, B. and P. Green (1998) Genome Res. 8:186-194.	
Phrap	A Phils Revised Assembly Program including SWAT and CrossMatch, programs based on efficient implementation of the Smith-Waterman algorithm, useful in searching sequence homology and assembling DNA sequences.	Smith, T.F. and M.S. Waterman (1981) Adv. Appl. Math. 2:482-489; Smith, T.F. and M.S. Waterman (1981) J. Mol. Biol. 147:195-197; and Green, P., University of Washington, Seattle, WA.	Score= 120 or greater; Match length= 56 or greater
Consed	A graphical tool for viewing and editing Phrap assemblies.	Gordon, D. et al. (1998) Genome Res. 8:195-202.	2.
SPScan	A weight matrix analysis program that scans protein sequences for the presence of secretory signal peptides.	Nielson, H. et al. (1997) Protein Engineering 10:1-6; Claverie, J.M. and S. Audic (1997) CABIOS 12:431-439.	Score=3.5 or greater
TMAP	A program that uses weight matrices to delineate transmembrane segments on protein sequences and determine orientation.	Persson, B. and P. Argos (1994) J. Mol. Biol. 237:182-192; Persson, B. and P. Argos (1996) Protein Sci. 5:363-371.	
TMHMMER	A program that uses a hidden Markov model (HMM) to delineate transmembrane segments on protein sequences and determine orientation.	Sonnhammer, E.L. et al. (1998) Proc. Sixth Intl. Conf. on Intelligent Systems for Mol. Biol., Glasgow et al., eds., The Am. Assoc. for Artificial Intelligence Press, Menio Park, CA, pp. 175-182.	ial 2.
Motifs	A program that searches amino acid sequences for patterns that matched those defined in Prosite.	Bairoch, A. et al. (1997) Nucleic Acids Res. 25:217-221; Wisconsin Package Program Manual, version 9, page M51-59, Genetics Computer Group, Madison, WI.	217-221; page VI.

What is claimed is:

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1. An isolated polypeptide selected from the group consisting of:

- a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-54,
- a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-53,
- c) a polypeptide consisting essentially of a naturally occurring amino acid sequence at least 91% identical to the amino acid sequence of SEQ ID NO:54,
- a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, and
- e) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-54.
- 2. An isolated polypeptide of claim 1 comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-54.
 - 3. An isolated polynucleotide encoding a polypeptide of claim 1.
 - 4. An isolated polynucleotide encoding a polypeptide of claim 2.
- 5. An isolated polynucleotide of claim 4 comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:55-108.
- 6. A recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide of claim 3.
 - 7. A cell transformed with a recombinant polynucleotide of claim 6.
 - 8. A transgenic organism comprising a recombinant polynucleotide of claim 6.
 - 9. A method of producing a polypeptide of claim 1, the method comprising:
- a) culturing a cell under conditions suitable for expression of the polypeptide, wherein said cell is transformed with a recombinant polynucleotide, and said recombinant

polynucleotide comprises a promoter sequence operably linked to a polynucleotide encoding the polypeptide of claim 1, and

- b) recovering the polypeptide so expressed.
- 5 10. A method of claim 9, wherein the polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO:1-54.
 - 11. An isolated antibody which specifically binds to a polypeptide of claim 1.
- 10 12. An isolated polynucleotide selected from the group consisting of:
 - a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:55-108,
 - a polynucleotide comprising a naturally occurring polynucleotide sequence at least
 90% identical to a polynucleotide sequence selected from the group consisting of
 SEQ ID NO:55-108,
 - c) a polynucleotide complementary to a polynucleotide of a),
 - d) a polynucleotide complementary to a polynucleotide of b), and
 - e) an RNA equivalent of a)-d).

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- 20 13. An isolated polynucleotide comprising at least 60 contiguous nucleotides of a polynucleotide of claim 12.
 - 14. A method of detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide of claim 12, the method comprising:
 - a) hybridizing the sample with a probe comprising at least 20 contiguous nucleotides comprising a sequence complementary to said target polynucleotide in the sample, and which probe specifically hybridizes to said target polynucleotide, under conditions whereby a hybridization complex is formed between said probe and said target polynucleotide or fragments thereof, and
- 30 b) detecting the presence or absence of said hybridization complex, and, optionally, if present, the amount thereof.
 - 15. A method of claim 14, wherein the probe comprises at least 60 contiguous nucleotides.
- 35 16. A method of detecting a target polynucleotide in a sample, said target polynucleotide

having a sequence of a polynucleotide of claim 12, the method comprising:

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a) amplifying said target polynucleotide or fragment thereof using polymerase chain reaction amplification, and

- b) detecting the presence or absence of said amplified target polynucleotide or fragment thereof, and, optionally, if present, the amount thereof.
- 17. A composition comprising a polypeptide of claim 1 and a pharmaceutically acceptable excipient.
- 18. A composition of claim 17, wherein the polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO:1-54.
- 19. A method for treating a disease or condition associated with decreased expression of functional SECP, comprising administering to a patient in need of such treatment the composition of
 claim 17.
 - 20. A method of screening a compound for effectiveness as an agonist of a polypeptide of claim 1, the method comprising:
 - a) exposing a sample comprising a polypeptide of claim 1 to a compound, and
 - b) detecting agonist activity in the sample.
 - 21. A composition comprising an agonist compound identified by a method of claim 20 and a pharmaceutically acceptable excipient.
- 22. A method for treating a disease or condition associated with decreased expression of functional SECP, comprising administering to a patient in need of such treatment a composition of claim 21.
- 23. A method of screening a compound for effectiveness as an antagonist of a polypeptide of claim 1, the method comprising:
 - a) exposing a sample comprising a polypeptide of claim 1 to a compound, and
 - b) detecting antagonist activity in the sample.
- 24. A composition comprising an antagonist compound identified by a method of claim 23and a pharmaceutically acceptable excipient.

25. A method for treating a disease or condition associated with overexpression of functional SECP, comprising administering to a patient in need of such treatment a composition of claim 24.

- 26. A method of screening for a compound that specifically binds to the polypeptide of claim
 5 1, the method comprising:
 - a) combining the polypeptide of claim 1 with at least one test compound under suitable conditions, and
 - b) detecting binding of the polypeptide of claim 1 to the test compound, thereby identifying a compound that specifically binds to the polypeptide of claim 1.

27. A method of screening for a compound that modulates the activity of the polypeptide of claim 1, the method comprising:

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- a) combining the polypeptide of claim 1 with at least one test compound under conditions permissive for the activity of the polypeptide of claim 1,
- b) assessing the activity of the polypeptide of claim 1 in the presence of the test compound, and
- c) comparing the activity of the polypeptide of claim 1 in the presence of the test compound with the activity of the polypeptide of claim 1 in the absence of the test compound, wherein a change in the activity of the polypeptide of claim 1 in the presence of the test compound is indicative of a compound that modulates the activity of the polypeptide of claim 1.
- 28. A method of screening a compound for effectiveness in altering expression of a target polynucleotide, wherein said target polynucleotide comprises a sequence of claim 5, the method comprising:
 - exposing a sample comprising the target polynucleotide to a compound, under conditions suitable for the expression of the target polynucleotide,
 - b) detecting altered expression of the target polynucleotide, and
 - c) comparing the expression of the target polynucleotide in the presence of varying amounts of the compound and in the absence of the compound.
 - 29. A method of assessing toxicity of a test compound, the method comprising:
 - a) treating a biological sample containing nucleic acids with the test compound,
 - b) hybridizing the nucleic acids of the treated biological sample with a probe comprising at least 20 contiguous nucleotides of a polynucleotide of claim 12 under conditions

whereby a specific hybridization complex is formed between said probe and a target polynucleotide in the biological sample, said target polynucleotide comprising a polynucleotide sequence of a polynucleotide of claim 12 or fragment thereof,

- c) quantifying the amount of hybridization complex, and
- d) comparing the amount of hybridization complex in the treated biological sample with the amount of hybridization complex in an untreated biological sample, wherein a difference in the amount of hybridization complex in the treated biological sample is indicative of toxicity of the test compound.
- 30. A diagnostic test for a condition or disease associated with the expression of SECP in a biological sample, the method comprising:
 - a) combining the biological sample with an antibody of claim 11, under conditions suitable for the antibody to bind the polypeptide and form an antibody:polypeptide complex, and
- b) detecting the complex, wherein the presence of the complex correlates with the presence of the polypeptide in the biological sample.
 - 31. The antibody of claim 11, wherein the antibody is:
 - a) a chimeric antibody,
 - b) a single chain antibody,
 - c) a Fab fragment,
 - d) a F(ab'), fragment, or
 - e) a humanized antibody.
- 25 32. A composition comprising an antibody of claim 11 and an acceptable excipient.
 - 33. A method of diagnosing a condition or disease associated with the expression of SECP in a subject, comprising administering to said subject an effective amount of the composition of claim 32.

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- 34. A composition of claim 32, wherein the antibody is labeled.
- 35. A method of diagnosing a condition or disease associated with the expression of SECP in a subject, comprising administering to said subject an effective amount of the composition of claim
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36. A method of preparing a polyclonal antibody with the specificity of the antibody of claim 11, the method comprising:

- immunizing an animal with a polypeptide consisting of an amino acid sequence a) selected from the group consisting of SEQ ID NO:1-54, or an immunogenic fragment thereof, under conditions to elicit an antibody response,
- b) isolating antibodies from said animal, and
- c) screening the isolated antibodies with the polypeptide, thereby identifying a polyclonal antibody which binds specifically to a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-54.

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- 37. A polyclonal antibody produced by a method of claim 36.
- 38. A composition comprising the polyclonal antibody of claim 37 and a suitable carrier.
- 15 39. A method of making a monoclonal antibody with the specificity of the antibody of claim 11, the method comprising:
 - a) immunizing an animal with a polypeptide consisting of an amino acid sequence selected from the group consisting of SEQ ID NO:1-54, or an immunogenic fragment thereof, under conditions to elicit an antibody response.
 - b) isolating antibody producing cells from the animal,
 - fusing the antibody producing cells with immortalized cells to form monoclonal c) antibody-producing hybridoma cells,
 - d) culturing the hybridoma cells, and
 - isolating from the culture monoclonal antibody which binds specifically to a e) polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-54.
 - 40. A monoclonal antibody produced by a method of claim 39.
- 30 41. A composition comprising the monoclonal antibody of claim 40 and a suitable carrier.
 - 42. The antibody of claim 11, wherein the antibody is produced by screening a Fab expression library.
- 35 43. The antibody of claim 11, wherein the antibody is produced by screening a recombinant

immunoglobulin library.

44. A method of detecting a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-54 in a sample, the method comprising:

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- a) incubating the antibody of claim 11 with a sample under conditions to allow specific binding of the antibody and the polypeptide, and
- b) detecting specific binding, wherein specific binding indicates the presence of a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-54 in the sample.

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- 45. A method of purifying a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-54 from a sample, the method comprising:
 - a) incubating the antibody of claim 11 with a sample under conditions to allow specific binding of the antibody and the polypeptide, and
 - separating the antibody from the sample and obtaining the purified polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-54.
- 46. A microarray wherein at least one element of the microarray is a polynucleotide of claim 20 13.
 - 47. A method of generating an expression profile of a sample which contains polynucleotides, the method comprising:
 - a) labeling the polynucleotides of the sample,
 - b) contacting the elements of the microarray of claim 46 with the labeled polynucleotides of the sample under conditions suitable for the formation of a hybridization complex, and
 - c) quantifying the expression of the polynucleotides in the sample.
- 48. An array comprising different nucleotide molecules affixed in distinct physical locations on a solid substrate, wherein at least one of said nucleotide molecules comprises a first oligonucleotide or polynucleotide sequence specifically hybridizable with at least 30 contiguous nucleotides of a target polynucleotide, and wherein said target polynucleotide is a polynucleotide of claim 12.

49. An array of claim 48, wherein said first oligonucleotide or polynucleotide sequence is completely complementary to at least 30 contiguous nucleotides of said target polynucleotide.

- 50. An array of claim 48, wherein said first oligonucleotide or polynucleotide sequence is completely complementary to at least 60 contiguous nucleotides of said target polynucleotide.
 - 51. An array of claim 48, wherein said first oligonucleotide or polynucleotide sequence is completely complementary to said target polynucleotide.
- 10 52. An array of claim 48, which is a microarray.
 - 53. An array of claim 48, further comprising said target polynucleotide hybridized to a nucleotide molecule comprising said first oligonucleotide or polynucleotide sequence.
- 15 54. An array of claim 48, wherein a linker joins at least one of said nucleotide molecules to said solid substrate.
 - 55. An array of claim 48, wherein each distinct physical location on the substrate contains multiple nucleotide molecules, and the multiple nucleotide molecules at any single distinct physical location have the same sequence, and each distinct physical location on the substrate contains nucleotide molecules having a sequence which differs from the sequence of nucleotide molecules at another distinct physical location on the substrate.
 - 56. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:1.
 - 57. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:2.
 - 58. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:3.
- 30 59. A polypeptide of claim 1, comprising the amino acid sequence of SEO ID NO:4.
 - 60. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:5.
 - 61. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:6.

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62. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:7. 63. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:8. 5 64. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:9. 65. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:10. 66. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:11. 10 67. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:12. 68. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:13. 15 69. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:14. 70. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:15. 71. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:16. 20 72. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:17. 73. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:18. 25 74. A polypeptide of claim 1, comprising the amino acid sequence of SEO ID NO:19. 75. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:20. 76. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:21. 30 77. A polypeptide of claim 1, comprising the amino acid sequence of SEO ID NO:22. 78. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:23. 79. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:24. 35

80. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:25. 81. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:26. 5 82. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:27. 83. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:28. 84. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:29. 10 85. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:30. 86. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:31. 15 87. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:32. 88. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:33. 89. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:34. 20 90. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:35. 91. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:36. 92. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:37. 25 93. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:38. 94. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:39. 30 95. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:40. 96. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:41. 35 97. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:42. 152

98. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:43. 99. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:44. 5 100. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:45. 101. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:46. 102. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:47. 10 103. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:48. 104. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:49. 15 105. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:50. 106. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:51. 107. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:52. 20 108. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:53. 109. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:54. 25 110. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:55. 111. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:56. 30 112. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:57. 113. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID

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 - 116. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:61.
- 10 117. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:62.
 - 118. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:63.
 - 119. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:64.

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- 120. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ IDNO:65.
 - 121. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:66.
- 25 122. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:67.
 - 123. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:68.
 - 124. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:69.
- 125. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ IDNO:70.

126. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:71.

- 127. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ IDNO:72.
 - 128. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:73.
- 10 129. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:74.
 - 130. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:75.
 - 131. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:76.

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- 132. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ IDNO:77.
 - 133. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:78.
- 25 134. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:79.
 - 135. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:80.
 - 136. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:81.
- 137. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ IDNO:82.

138. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:83.

- 139. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID 5 NO:84.
 - 140. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:85.
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 - 142. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:87.

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- 143. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:88.
- 144. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID

 20 NO:89.
 - 145. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:90.
- 25 146. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:91.
 - 147. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:92.
 - 148. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:93.
- 149. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ IDNO:94.

150. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:95.

- 151. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID 5 NO:96.
 - 152. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:97.
- 10 153. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:98.
 - 154. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:99.

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- 155. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:100.
- 156. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID 20 NO:101.
 - 157. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:102.
- 25 158. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:103.
 - 159. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:104.
 - 160. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:105.
- 161. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ IDNO:106.

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      DING, Li
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      LEE, Ernestine A.
      RAMKUMAR, Jayalaxmi
      THANGAVELU, Kavitha
      XU, Yuming
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      TANG, Y. Tom
      NGUYEN, Danniel B.
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      HONCHELL, Cynthia D.
      GIETZEN, Kimberly J.
      BAUGHN, Mariah R.
      GANDHI, Ameena R.
      ARVIZU, Chandra WALIA, Narinder K.
      LU, Yan
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      HAFALIA, April J.A.
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Thr Arg Gln Pro Leu Gln Thr Asp Ile Tyr Gly Leu Ala Lys Lys
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 Cys Asn Leu Thr Glu Arg Gln Val Glu Arg Trp Phe Arg Ser Arg
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 Arg Asn Gln Glu Arg Pro Ser Arg Leu Lys Lys Phe Gln Glu Ala
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                                     130
 Cys Trp Arg Phe Ala Phe Tyr Leu Met Ile Thr Val Ala Gly Ile
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 Ala Phe Leu Tyr Asp Lys Pro Trp Leu Tyr Asp Leu Trp Glu Val
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                                     160
 Trp Asn Gly Tyr Pro Lys Gln Pro Leu Leu Pro Ser Gln Tyr Trp
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                                     175
                                                          180
 Tyr Tyr Ile Leu Glu Met Ser Phe Tyr Trp Ser Leu Leu Phe Arg
                 185
                                     190
 Leu Gly Phe Asp Val Lys Arg Lys Asp Phe Leu Ala His Ile Ile
                 200
                                     205
                                                          210
 His His Leu Ala Ala Ile Ser Leu Met Ser Phe Ser Trp Cys Ala
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 Asn Tyr Ile Arg Ser Gly Thr Leu Val Met Ile Val His Asp Val
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 Ala Asp Ile Trp Leu Glu Ser Ala Lys Met Phe Ser Tyr Ala Gly
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 Trp Thr Gln Thr Cys Asn Thr Leu Phe Phe Ile Phe Ser Thr Ile
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 Phe Phe Ile Ser Arg Leu Ile Val Phe Pro Phe Trp Ile Leu Tyr
                 275
                                     280
                                                          285
 Cys Thr Leu Ile Leu Pro Met Tyr His Leu Glu Pro Phe Phe Ser
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                                     295
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 Tyr Ile Phe Leu Asn Leu Gln Leu Met Ile Leu Gln Val Leu His
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                                     310
 Leu Tyr Trp Gly Tyr Tyr Ile Leu Lys Met Leu Asn Arg Cys Ile
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 Phe Met Lys Ser Ile Gln Asp Val Arg Ser Asp Asp Glu Asp Tyr
                 335
                                     340
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 Glu Glu Glu Glu Glu Glu Glu Glu Glu Ala Thr Lys Gly Lys
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                                     355
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 Glu Met Asp Cys Leu Lys Asn Gly Leu Gly Ala Glu Arg His Leu
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Cys Val Arg Asn Lys Ala Ala Lys Leu His Val Val Ile Gln Gln
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Arg Lys Val Thr Gly Ala Asp Leu Thr Trp Ser Pro Gly Asp Gly
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                                     40
Ile Gln Phe Gln Val Pro Gly Thr Arg Lys Thr Lys Gln Tyr Cys
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Glu Phe Glu Asn Glu Ile Asn Phe Ile Met Pro His Met Lys Ile
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Cys Lys Glu Phe Met Gly Ile Met Thr Leu Gly Cys Leu Pro Thr
                 35
                                     40
Pro Ala Pro Leu His Leu Phe Phe Ser Leu Ser Pro Ala Arg Val
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Leu Arg Ala Pro Tyr Gly Ala Gln Glu Lys Lys Gly Arg Arg Val
                 65
                                     70
Arg Thr Thr Pro Trp Arg Arg Pro Pro Trp Arg Thr Ser Gly His
                 80
                                     85
Trp Gly Arg Asp Pro Ile Arg Glu Asn Cys Pro Gln Gln Ser Glu
                 95
                                    100
Glu Leu Ser Trp Pro Trp Ile Leu Arg Trp Ala Leu Leu Cys Ala
                110
                                    115
Leu Arg Gln Ala Thr Cys Pro Leu Ser Leu Ser Phe Leu Ile Cys
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125
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Thr Thr Gly Pro Ile Ser Leu Thr Ser Gln Val Ala Leu Gly Asp
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Arg Cys Ala Trp His Ile Val Gly Val Gln
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Ile Leu Leu Val Cys Leu His Arg Pro Asp Ala Arg Val Pro Cys
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                                     25
Leu Ile Leu His Leu Leu Ser His Trp Gly Ser Leu Pro Ser Asp
                 35
                                     40
Ala Leu Ala Lys Ile Ala Leu Val Cys Ser Arg Lys Glu Gly Gln
                                    55
                 50
Ile Pro Gly Ile Val Arg Ala Ala Glu Leu Tyr Arg Ile Gly Leu
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Pro Phe Pro Pro Val Trp Leu Ala Leu His Ser Leu Gln Ile Pro
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Pro Thr Ser Thr Gln
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Tyr Leu Ser Lys Arg Gln Glu Val Glu Arg Cys Gly Tyr Met Lys
                 35
                                     40
                                                         45
Pro Ser Leu Asn Thr Ile Ser Ser Pro Glu Ser His Pro Val Thr
                                     55
                50
Ser His Ile His Thr Ser Gln Asp Arg Arg Lys Trp Pro Ala Leu
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Ala Cys Lys Lys Gly Trp Glu Met Glu Ala Phe Phe Tyr Tyr Tyr
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Tyr Phe
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Leu Val
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                                     40
Asp Ser Pro Ala Ser Ile Ala Val Thr Asp Ile Thr Ile His Ile
                 50
                                     55
Gln Ile Val Leu Leu Ala Thr Leu Leu Ala Ser Ser Phe Thr Lys
                 65
                                     70
Ser Pro Asp Phe Ser Tyr Asn Pro Asp Leu Ser Phe Thr Ser Ser
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Tyr Met Thr Ser Gly Met Leu Leu Asp Ile Ser Glu Leu Gln Tyr
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Pro Tyr Val Gln Ser Glu Thr Ile
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Gln Cys Val Thr Cys Gln Leu Pro His His Pro Pro Pro Ser Leu
                 35
                                     40
Pro Pro Leu Leu Pro Gln Gly Pro Pro Pro Ile Ser Gly Ser Gln
                 50
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Ala Ile Asn Leu Glu Thr Glu Met Gly Leu Leu Ser Ile Leu Trp
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Pro Leu Phe Leu Ser Leu Gln Phe Val Pro
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                                      40
Ala His Gln Leu Ala Ile Asp Thr Tyr Gln Glu Phe Glu Glu Thr
                 50
                                     55
Tyr Ile Pro Lys Asp Gln Lys Tyr Ser Phe Leu His Asp Ser Gln
                 65
                                     70
Thr Ser Phe Cys Phe Ser Asp Ser Ile Pro Thr Pro Ser Asn Met
                 80
                                     85
                                                         90
Glu Glu Thr Gln Gln Lys Ser Asn Leu Glu Leu Leu Arg Ile Ser
                 95
                                    100
Leu Leu Leu Ile Glu Ser Trp Leu Glu Pro Val Arg Phe Leu Arg
                110
                                    115
                                                         120
Ser Met Phe Ala Asn Asn Leu Val Tyr Asp Thr Ser Asp Ser Asp
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                                    130
Asp Tyr His Leu Leu Lys Asp Leu Glu Glu Gly Ile Gln Thr Leu
                140
                                    145
                                                         150
Met Gly Val Arg Val Ala Pro Gly Val Thr Asn Pro Gly Thr Pro
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                                    160
Leu Ala Ser Arg Ala Gly Gly Glu Lys Tyr Cys Cys Pro Leu Phe
                170
                                    175
                                                         180
Ser Ser Lys Ala Leu Thr Gln Glu Asn Ser Pro Tyr Ser Ser Phe
                185
                                    190
Arg Leu Val Asn Pro Pro Gly Leu Ser Leu His Pro Glu Gly Glu
                200
                                    205
Gly Gly Lys Trp Ile Asn Glu Arg Gly Arg Glu Gln Cys Pro Ser
                215
                                    220
                                                         225
Ala Trp Pro Leu Leu Phe Leu His Phe Ala Glu Ala Gly Arg
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Arg Gln Pro Pro Asp Trp Ala Asp Pro Gln Ala Asp Leu Gln Gln
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                                     40
                                                          45
Ser Pro Cys Glu Gly Leu Pro Ala Ala Asp Ala Thr Ala Leu Thr
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                                     55
Leu Ala Asn Arg Asn Leu Glu Arg Leu Pro Gly Cys Leu Pro Arg
                 65
                                     70
Thr Leu Arg Ser Leu Asp Ala Ser His Asn Leu Leu Arg Ala Leu
                 80
                                     85
Ser Thr Ser Glu Leu Gly His Leu Glu Gln Leu Gln Val Leu Thr
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Leu Arg His Asn Arg Ile Ala Ala Leu Arg Trp Gly Pro Gly Gly

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110
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Pro Ala Gly Leu His Thr Leu Asp Leu Ser Tyr Asn Gln Leu Ala
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Ala Leu Pro Pro Cys Thr Gly Pro Ala Leu Ser Ser Leu Arg Ala
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Leu Ala Leu Ala Gly Asn Pro Leu Arg Ala Leu Gln Pro Arg Ala
                 155
                                     160
                                                          165
Phe Ala Cys Phe Pro Ala Leu Gln Leu Leu Asn Leu Ser Cys Thr
                170
                                     175
                                                          180
Ala Leu Gly Arg Gly Ala Gln Gly Gly Ile Ala Glu Ala Ala Phe
                 185
                                     190
Ala Gly Glu Asp Gly Ala Pro Leu Val Thr Leu Glu Val Leu Asp
                 200
                                     205
Leu Ser Gly Thr Phe Leu Glu Arg Val Glu Ser Gly Trp Ile Arg
                 215
                                     220
Asp Leu Pro Lys Leu Thr Ser Leu Tyr Leu Arg Lys Met Pro Arg
                 230
                                     235
                                                          240
Leu Thr Thr Leu Glu Gly Asp Ile Phe Lys Met Thr Pro Asn Leu
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                                     250
                                                          255
Gln Gln Leu Asp Cys Gln Asp Ser Pro Ala Leu Ala Ser Val Ala
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Thr His Ile Phe Gln Asp Thr Pro His Leu Gln Val Leu Leu Phe
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Leu Glu Pro Glu Ala Ala Ala Gln Ala His Tyr Lys Ala Cys Asp
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Arg Leu Lys Leu Glu Arg Lys Gln Arg Arg Met Cys Arg Arg Asp
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Pro Gly Val Ala Glu Thr Leu Val Glu Ala Val Ser Met Ser Ala
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                                      85
Leu Glu Cys Gln Phe Gln Phe Arg Phe Glu Arg Trp Asn Cys Thr
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Leu Glu Gly Arg Tyr Arg Ala Ser Leu Leu Lys Arg Gly Phe Lys
                                     115
Glu Thr Ala Phe Leu Tyr Ala Ile Ser Ser Ala Gly Leu Thr His
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                                     130
Ala Leu Ala Lys Ala Cys Ser Ala Gly Arg Met Glu Arg Cys Thr
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                                     145
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Cys Asp Glu Ala Pro Asp Leu Glu Asn Arg Glu Ala Trp Gln Trp
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.Gly Gly Cys Ser Glu Asp Ile Glu Phe Gly Gly Met Val Ser Arg
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Glu Phe Ala Asp Ala Arg Glu Asn Arg Pro Asp Ala Arg Ser Ala
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Met Asn Arg His Asn Asn Glu Ala Gly Arg Gln Val Ile Lys Ala

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200
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Gly Val Glu Thr Thr Cys Lys Cys His Gly Val Ser Gly Ser Cys
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Thr Val Arg Thr Cys Trp Arg Gln Leu Ala Pro Phe His Glu Val
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                                    235
Gly Lys His Leu Lys His Lys Tyr Glu Thr Ala Leu Lys Val Gly
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                                    250
                                                         255
Ser Thr Thr Asn Glu Ala Ala Gly Glu Ala Gly Ala Ile Ser Pro
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Pro Arg Gly Arg Ala Ser Gly Ala Gly Gly Ser Asp Pro Leu Pro
                275
                                    280
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Arg Thr Pro Glu Leu Val His Leu Asp Asp Ser Pro Ser Phe Cys
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                                    295
                                                         300
Leu Ala Gly Arg Phe Ser Pro Gly Thr Ala Gly Arg Arg Cys His
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                                    310
Arg Glu Lys Asn Cys Glu Ser Ile Cys Cys Gly Arg Gly His Asn
                320
                                    325
Thr Gln Ser Arg Val Val Thr Arg Pro Cys Gln Cys Gln Val Arg
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Trp Cys Cys Tyr Val Glu Cys Arg Gln Cys Thr Gln Arg Glu Glu
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Val Tyr Thr Cys Lys Gly
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Val Arg Gln Glu Val Gly Leu Cys Leu Glu Arg Gln Ser Leu Gln
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Leu Asp Pro Ala Leu Ser Ser Leu Ser Gln Gly Trp Pro Leu Arg
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Arg Pro Leu Pro Phe Ile Cys Pro Ser Pro Pro Ser Pro Arg Leu
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                                     70
Thr Cys Leu Pro Pro Leu Ala Leu Ser Ser Leu Thr Gly Arg Glu
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                                     85
Val Leu Thr Pro Phe Pro Gly Leu Gly Thr Ala Ala Ala Pro Ala
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                                    100
Gln Gly Gly Ala His Leu Lys Gln Cys Asp Leu Leu Lys Leu Ser
                110
                                    115
                                                         120
Arg Arg Gln Lys Gln Leu Cys Arg Arg Glu Pro Gly Leu Ala Glu
                125
                                    130
                                                         135
Thr Leu Arg Asp Ala Ala His Leu Gly Leu Leu Glu Cys Gln Phe
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                                    145
Gln Phe Arg His Glu Arg Trp Asn Cys Ser Leu Glu Gly Arg Met
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                                    160
Gly Leu Leu Lys Arg Gly Phe Lys Glu Thr Ala Phe Leu Tyr Ala
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                                    175
Val Ser Ser Ala Ala Leu Thr His Thr Leu Ala Arg Ala Cys Ser
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                                    190
Ala Gly Arg Met Glu Arg Cys Thr Cys Asp Asp Ser Pro Gly Leu
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                                    205
Glu Ser Arg Gln Ala Trp Gln Trp Gly Val Cys Gly Asp Asn Leu
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215
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 Lys Tyr Ser Thr Lys Phe Leu Ser Asn Phe Leu Gly Ser Lys Arg
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 Gly Asn Lys Asp Leu Arg Ala Arg Ala Asp Ala His Asn Thr His
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 Val Gly Ile Lys Ala Val Lys Ser Gly Leu Arg Thr Thr Cys Lys
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                                      265
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 Cys His Gly Val Ser Gly Ser Cys Ala Val Arg Thr Cys Trp Lys
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                                      280
                                                           285
 Gln Leu Ser Pro Phe Arg Glu Thr Gly Gln Val Leu Lys Leu Arg
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                                      295
                                                           300
 Tyr Asp Ser Ala Val Lys Val Ser Ser Ala Thr Asn Glu Ala Leu
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                                      310
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 Gly Arg Leu Glu Leu Trp Ala Pro Ala Arg Gln Gly Ser Leu Thr
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                                      325
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 Lys Gly Leu Ala Pro Arg Ser Gly Asp Leu Val Tyr Met Glu Asp
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                                                           345
 Ser Pro Ser Phe Cys Arg Pro Ser Lys Tyr Ser Pro Gly Thr Ala
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 Gly Arg Val Cys Ser Arg Glu Ala Ser Cys Ser Ser Leu Cys Cys
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 Gly Arg Gly Tyr Asp Thr Gln Ser Arg Leu Val Ala Phe Ser Cys
                  380
                                      385
                                                           390
 His Cys Gln Val Gln Trp Cys Cys Tyr Val Glu Cys Gln Gln Cys
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· Val Gln Glu Glu Leu Val Tyr Thr Cys Lys His
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                   35
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 Ile Leu Val Leu Thr Asp Asp Gln Asp Val Glu Leu Gly Ser Leu
                   50
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 Gln Val Met Asn Lys Thr Arg Lys Ile Met Glu His Gly Gly Ala
                   65
                                       70
 Thr Phe Ile Asn Ala Phe Val Thr Thr Pro Met Cys Cys Pro Ser
                   80
                                       85
 Arg Ser Ser Met Leu Thr Gly Lys Tyr Val His Asn His Asn Val
                   95
                                      100
 Tyr Thr Asn Asn Glu Asn Cys Ser Ser Pro Ser Trp Gln Ala Met
                  110
                                      115
                                                           120
 His Glu Pro Arg Thr Phe Ala Val Tyr Leu Asn Asn Thr Gly Tyr
                  125
                                      130
                                                           135
 Arg Thr Ala Phe Phe Gly Lys Tyr Leu Asn Glu Tyr Asn Gly Ser
                                      145
                 140
                                                           150
 Tyr Ile Pro Pro Gly Trp Arg Glu Trp Leu Gly Leu Ile Lys Asn
                  155
                                      160
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 Ser Arg Phe Tyr Asn Tyr Thr Val Cys Arg Asn Gly Ile Lys Glu
                  170
                                      175
 Lys His Gly Phe Asp Tyr Ala Lys Asp Tyr Phe Thr Asp Leu Ile
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200 205 206 205 206 205	Thr	Asn	Glu	Ser	185 Ile	Asn	Tvr	Phe	Lvs	190 Met	Ser	Lvs	Ara	Met	195 Tvr
215 220 221 220 221 220 221 220 221 220 221					200					205					210
Gly Pro Glu Asp Ser Ala Pro Gln Phe Ser Lys Leu Tyr Pro A 230 Ala Ser Gln His Ile Thr Pro Ser Tyr Asn Tyr Ala Pro Asn M 245 Asp Lys His Tyr Ile Met Gln Tyr Thr Gly Pro Met Leu Pro I 265 His Met Glu Phe Thr Asn Ile Leu Gln Arg Lys Arg Leu Gln Tyr 290 Val Glu Thr Gly Glu Leu Glu Asn Thr Tyr Ile Tyr Asn Met Leu Pro I 305 Asp His Gly Tyr His Ile Glu Asn Thr Tyr Ile I Ile Tyr Thr Ash 305 Asp His Gly Tyr His Ile Gly Gln Phe Gly Leu Val Lys Gly Ing 335 Gly Pro Ser Val Glu Pro Gly Ser Ile Val Pro Gl Ile Val Is 305 Asn Ile Asp Leu Ala Pro Thr Ile Leu Asp Ile Ala Gly Leu Ash 310 Asn Ile Asp Leu Ala Pro Thr Ile Leu Asp Ile Ala Gly Leu Ash 310 Asn Ile Asp Leu Ala Pro Thr Ile Leu Asp Ile Ala Gly Leu Ash 310 Asn Ile Asp Leu Ala Pro Thr Ile Leu Asp Ile Ala Gly Leu Ash 310 Asn Ile Asp Leu Ala Pro Thr Ile Leu Asp Ile Ala Gly Leu Ash 310 Asn Ile Asp Cash Ash Arg Phe Arg Thr Ash Lys Leu Leu Ash 380 Asn Ile Asp Asp Thr Phe Leu Val Glu Arg Gly Lys Fhe Leu Ash 380 Asn Ile Arg Cash Ash Arg Phe Arg Thr Ash Lys Lys Ala Ash 385 Fro Glu Lys Pro Gly Ash Asp Gly Lys Glu Leu Cys Gln Gln Ala Arg Ty Ash 400 Ala Cash Ala Cys Glu Gln Pro Gly Gln Lys Cys Lys Gly Pro Ser Ash Ala Cys Glu Gln Pro Gly Glu Lys Cys Lys Gly Fro Asp Ash Arg Gly Cys Ser Cys Lys Gly Fro Asp Ash Arg Gly Cys Ser Cys Lys Gly Pro Ser Ash Ala Cys Glu Gln Pro Gly Gln Lys Cys Lys Gly Pro Ser Asp Thr Arg Ash Cys Lys Asp Cys Lys Gly Pro So	Pro	HIS	Arg	Pro		Met	Met	Val	ITE		His	Ala	Ala	Pro	His 225
245					230					235					Asn 240
Asp Lys His Trp Tle Met Gln Tyr Thr Gly Pro Met Leu Pro Trophology Leu Met Glu Phe Thr Asn Tle Leu Gln Arg Lys Arg Leu Gln Trophology Leu Met Ser Val Asp Asp Ser Val Glu Arg Leu Tyr Asn Met Leu Glu Thr Gly Glu Leu Glu Arg Leu Tyr Thr Arg Asp Asp Ser Val Glu Arg Leu Val Lys Gly Lys Lys Gly Gly Gly Lys Gly Gl	Ala	Ser	Gln	His		Thr	Pro	Ser	Tyr		Tyr	Ala	Pro	Asn	Met 255
His Met Glu Phe Thr Asn Ile Leu Gln Arg Lys Arg Leu Gln Met Ser Val Asp Asp Ser Val Glu Arg Leu Tyr Asn Met Leu Glu Thr Glu Glu Leu Glu Glu Arg Leu Tyr Asn Met Leu Glu Thr Glu	Asp	Lys	His	Trp		Met	Gln	Tyr	Thr		Pro	Met	Leu	Pro	Ile 270
Leu Met Ser Val Asp Asp Asp Ser Val Glu Arg Leu Tyr Asn Met Can	His	Met	Glu	Phe	Thr	Asn	Ile	Leu	Gln	Arg	Lys	Arg	Leu	Gln	
Val Glu Thr Gly Glu Leu Glu Asn Thr Tyr Thr Asn Asn Asn Asn Thr Tyr Thr Asn Asn Asn Asn Asn Thr Tyr Thr Asn	Leu	Met	Ser	Val	qaA	Asp	Ser	Val	Glu	Arg	Leu	Tyr	Asn	Met	
Asp His Gly Tyr His J20 Ile Gly Gln Phe Gly Leu Val Lys Gly Days Leu Val Pro Phe Phe Ile Val Pro Phe Phe Ile Val And Say Jacob Jacob<	Val	Glu	Thr	Gly		Leu	Glu	Asn	Thr	Tyr	Ile	Ile	Tyr	Thr	
Ser Met Pro Tyr Asp Pro Asp 11e Arg 335 340 340 350 355 355 365	Asp	His	Gly	Tyr		Ile	Gly	Gln	Phe	Gly	Leu	Val	Lys	Gly	
Gly Pro Ser Val Glu Pro Gly Ser Ile Val Pro Gln Ile Val Val Asp Ile Asp Leu Ala Pro Thr Ile Leu Asp Ile Ala Gly Leu Asp Ile	Ser	Met	Pro	Tyr	_	Phe	qaA	Ile	Arg		Pro	Phe	Phe	Ile	Arg 345
Asn Ile Asp Leu Ala Pro Thr Ile Leu Asp Ile Ile <td>Gly</td> <td>Pro</td> <td>Ser</td> <td>Val</td> <td></td> <td>Pro</td> <td>Gly</td> <td>Ser</td> <td>Ile</td> <td>Va1</td> <td>Pro</td> <td>Gln</td> <td>Ile</td> <td>Val</td> <td></td>	Gly	Pro	Ser	Val		Pro	Gly	Ser	Ile	Va1	Pro	Gln	Ile	Val	
Second S	Asn	Ile	Asp	Leu		Pro	Thr	Ile	Leu	Asp	Ile	Ala	Gly	Leu	
Pro Glu Lys Pro Gly Asn Arg Phe Arg Thr Asn Lys Lys Ala Lys Ille Trp Arg Asp Thr Arg Asp Asp Ath	Thr	Pro	Pro	Asp		Asp	Gly	Lys	Ser	Val	Leu	Lys	Leu	Leu	
The Try Arg Asp	Pro	Glu	Lys	Pro		Asn	Arg	Phe	Arg	Thr	Asn	Lys	Lys	Ala	
Pro Lys Tyr Glu Arg Val Lys Glu Leu Cys Gln Gln Ala Arg Ty 440	Ile	Trp	Arg	Asp		Phe	Leu	Val	Glu		Gly	Lys	Phe	Leu	Arg 420
Gln Thr Ala Cys Glu Gln Pro Gly Gln Lys Trp Gln Cys Ile Gl 455 Asp Thr Ser Gly Lys Leu Arg Ile His Lys Cys Lys Gly Pro Ser 470 Asp Leu Leu Thr Val Arg Gln Ser Thr Arg Asn Leu Tyr Ala Asp Lys Foo So 500 Tyr Arg Ala Ser Arg Ser Gln Arg Lys Ser Gln Arg Gln Pro So 505 Arg Asn Gln Gly Thr Pro Lys Tyr Lys Pro Arg Phe Val His Tyr Say Asp Ile Asn Leu Glu Glu Glu Glu Leu Gln Pro So 505 Arg Asn Ile Asn Leu Glu Glu Glu Glu Glu Leu Gln Val Leu Gln Pro So 505 Arg Asn Ile Ala Lys Arg His Asp Glu Gly His Lys Gly Pro Arg Asp Leu Gln Pro So 505 Arg Asn Ile Ala Ser Ser Gln Glu Glu Glu Glu Leu Gln Val Leu Gln Pro So 505 Arg Asn Ile Ala Lys Arg His Asp Glu Gly His Lys Gly Pro Arg Asp Leu Gln Arg Gln Pro So 505 Arg Asn Ile Ala Ser Ser Gly Gly Asn Arg Gly Arg Met Leu Arg Ser Leu Glu Gly His Lys Gly Pro Arg So 505 Arg Asn Ile Ala Ser Ser Gly Gly Asn Arg Gly Arg Met Leu Arg Ser Ser Ser Asn Ala Val Gly Pro Pro Thr Thr Val Arg Val Thr 605 Asp Ser Ser Asn Ala Val Gly Pro Pro Thr Thr Val Arg Val Thr 605 Glu Leu Tyr Gln Ser Ala Arg Ala Trp Lys Asp His Lys Glu Arg Glu Leu Tyr Gln Ser Ala Arg Ala Trp Lys Asp His Lys Asn Leu Glu Leu Tyr Gln Ser Ala Arg Ala Trp Lys Asp Lys Ile Lys Asn Leu Glu Leu Tyr Gln Ser Ala Arg Ala Trp Lys Asp Lys Ile Lys Asn Leu Glu Leu Tyr Gln Ser Ala Arg Ala Trp Lys Asp Lys Ile Lys Asn Leu Glu Leu Tyr Gln Ser Ala Arg Ala Trp Lys Asp Lys Ile Lys Asn Leu Lys Ile Lys Asn Leu Lys Ile Lys Asn Leu Ileu Glu Ala Leu Gln Arg Lys Glu Ileu Glu Leu Gln Arg Lys Glu Arg Glu Glu Leu Tyr Gln Ser Ala Arg Ala Trp Lys Asp Lys Ile Lys Asn Leu Lys Arg Lys Ileu Lys Asn Leu Lys Ileu Ly	Lys	Lys	Glu	Glu		Ser	Lys	Asn	Ile		Gln	Ser	Asn	His	Leu 435
Asp Thr Ser Gly Lys Leu Arg Ile His Lys Cys Lys Gly Pro Ser Arg Leu Leu Thr Val Arg Gln Ser Thr Arg Asn Leu Tyr Ala Arg Gly Phe His Asp Lys Asp Lys Glu Cys Ser Cys Arg Glu Ser Gly Tyr Arg Ala Ser Arg Ser Gln Arg Lys Ser Gln Arg Gl	Pro	Lys	туг	Glu		Val	Lys	Glu	Leu		Gln	Gln	Ala	Arg	Tyr 450
Asp Leu Leu Thr Val Arg Gln Ser Thr Arg Asn Leu Tyr Ala Arg Gly Phe His Asp Lys Asp Lys Glu Cys Ser Cys Arg Glu Ser Gry Arg Asn Arg Gln Ser Gly Arg Asn Gln Gly Thr Pro Lys Tyr Lys Pro Arg Phe Val His Thr San Gln Gly Thr Pro Lys Tyr Lys Pro Arg Phe Val His Thr San Gln Gly Thr Pro Lys Tyr Lys Pro Arg Phe Val His Thr San Gln Gly Glu Glu Glu Glu Phe Glu Gly Glu Ile Thr San Gln Gly Glu	Gln	Thr	Ala	Суз		Gln	Pro	Gly	Gln		Trp	Gln	Сув	Ile	Glu 465
Sty Phe His Asp Lys Asp Lys Glu Cys Ser Cys Arg Glu Ser Glu Ser Glu Ser Sty Arg Ala Ser Arg Ser Gln Arg Lys Ser Gln Arg Gln Phe Log Ser Gln Arg Phe Val His The Ser Ser Log Ser	Asp	Thr	Ser	Gly		Leu	Arg	Ile	His		Суѕ	Lys	Gly	Pro	Ser 480
Tyr Arg Ala Ser Arg Ser Gln Arg Lys Ser Gln Arg Gln Phe Legar Arg Asn Gln Gly Thr Pro Lys Tyr Lys Pro Arg Phe Val His The Same of Same	qaA	Leu	Leu	Thr		Arg ·	Gln	Ser	Thr		Asn	Leu	Tyr	Ala	Arg 495
Arg Asn Gln Gly Thr Pro Lys Tyr Lys Pro Arg Phe Val His The San Gln Gly Thr Pro Lys Tyr Lys Pro Arg Phe Val His The San Gln Thr Arg Ser Leu Ser Val Glu Phe Glu Gly Glu Ile The San Leu Glu Glu Glu Glu Glu Glu Glu Glu Glu Gl	Gly	Phe	His	Asp		Asp	. Lys	Glu	Суз		Cys	Arg	Glu	Ser	Gly 510
Arg Gln Thr Arg Ser Leu Ser Val Glu Phe Glu Gly Glu Ile Ty 545 Asp Ile Asn Leu Glu Glu Glu Glu Glu Leu Gln Val Leu Gln Ph 560 Arg Asn Ile Ala Lys Arg His Asp Glu Gly His Lys Gly Pro Arg 575 Asp Leu Gln Ala Ser Ser Gly Gly Asn Arg Gly Arg Met Leu Glu Gly 590 Asp Ser Ser Asn Ala Val Gly Pro Pro Thr Thr Val Arg Val Th 605 His Lys Cys Phe Ile Leu Pro Asn Asp Ser Ile His Cys Glu As 620 Glu Leu Tyr Gln Ser Ala Arg Ala Trp Lys Asp His Lys Ala Ty 635 Ile Asp Lys Glu Ile Glu Ala Leu Gln Asp Lys Ile Lys Asn Le	Tyr	Arg	Ala	Ser	_	Ser	Gln	Arg	Lys		Gln	Arg	Gln	Phe	Leu 525
Asp Ile Asn Leu Glu Glu Glu Glu Glu Glu Leu Gln Val Leu Gln Pro S60 550 Arg Asn Ile Ala Lys Arg His Asp Glu Gly His Lys Gly Pro Arg S75 580 Asp Leu Gln Ala Ser Ser Gly Gly Asn Arg Gly Arg Met Leu Asp Ser Ser Asn Ala Val Gly Pro Pro Thr Thr Val Arg Val The G05 60 Asp Ser Ser Asn Ala Val Gly Pro Pro Thr Thr Val Arg Val The G05 60 His Lys Cys Phe Ile Leu Pro Asn Asp Ser Ile His Cys Glu Asp G25 625 Glu Leu Tyr Gln Ser Ala Arg Ala Trp Lys Asp His Lys Ala Ty G35 640 Ile Asp Lys Glu Ile Glu Ala Leu Gln Asp Lys Ile Lys Asn Leg	Arg	Asn	Gln	Gly		Pro	Lys	Tyr	Lys		Arg	Phe	Val	His	Thr 540
Arg Asn Ile Ala Lys Arg His Asp Glu Gly His Lys Gly Pro Asp Ser Ser Ser Ser Gly Gly Asn Arg Gly Arg Met Leu Asp Ser Ser Gly Gly Asn Arg Gly Arg Met Leu Asp Ser Ser Asn Ala Val Gly Pro Pro Thr Thr Val Arg Val The Got	Arg	Gln	Thr	Arg		Leu	Ser	Val	Glu		Glu	Gly	Glu	Ile	Tyr 555
Asp Leu Gln Ala Ser Ser Gly Gly Asn Arg Gly Arg Met Leu Al Ser Ser Gly Gly Pro Pro Thr Thr Val Arg Val The Glo	Asp	Ile	Asn	Leu		Glu	Glu	Glu	Glu		Gln	Val	Leu	Gln	Pro 570
Asp Ser Ser Asn Ala Val Gly Pro Pro Thr Thr Val Arg Val The Got Ser Asn Ala Val Gly Pro Pro Thr Thr Val Arg Val The Got Ser Ala Arg Val The Got Ser Ile His Cys Glu Arg Val The Got Ser Ala Arg Ala Trp Lys Asp His Lys Ala Trg Cys Glu Leu Tyr Gln Ser Ala Arg Ala Trp Lys Asp His Lys Ala Trg Got Ser Ser Ile Asp Lys Glu Ile Glu Ala Leu Gln Asp Lys Ile Lys Asn Leg Ile Asp Lys Glu Ile Glu Ala Leu Gln Asp Lys Ile Lys Asn Leg Ile Asp Lys Glu Ile Glu Ala Leu Gln Asp Lys Ile Lys Asn Leg Ile Asp Lys Glu Ile Glu Ala Leu Gln Asp Lys Ile Lys Asn Leg Ile Asp Lys Glu Ile Glu Ala Leu Gln Asp Lys Ile Lys Asn Leg Ile Asp Lys Ile Lys Asn Lys Ile Asp Lys Ile Lys Ile Asp Lys Ile Lys Asn Lys Ile Asp Lys Ile Lys	Arg	Asn	Ile	Ala		Arg	His	Asp	Glu		His	Lys	Gly	Pro	Arg 585
605 610 610 611 611 611 611 611 611 611 611	Asp	Leu	Gln	Ala		Ser	Gly	Gly	Asn		Gly	Arg	Met	Leu	Ala 600
Glu Leu Tyr Gln Ser Ala Arg Ala Trp Lys Asp His Lys Ala Tr 635 640 66 Ile Asp Lys Glu Ile Glu Ala Leu Gln Asp Lys Ile Lys Asn Le	Asp	Ser	Ser	Asn		Val	Gly	Pro	Pro		Thr	Val	Arg	Val	Thr 615
Glu Leu Tyr Gln Ser Ala Arg Ala Trp Lys Asp His Lys Ala Ty 635 640 66 Ile Asp Lys Glu Ile Glu Ala Leu Gln Asp Lys Ile Lys Asn Le					620					625					Arg 630
Ile Asp Lys Glu Ile Glu Ala Leu Gln Asp Lys Ile Lys Asn Le					635					640					Tyr 645
	Ile	Asp	Lys	Glu		Glu	Ala	Leu	Gln	Asp	Lys	Ile	ГЛа	Asn	Leu 660

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Arg Glu Val Arg Gly His Leu Lys Arg Arg Lys Pro Glu Glu Cys
                665
                                    670
Ser Cys Ser Lys Gln Ser Tyr Tyr Asn Lys Glu Lys Gly Val Lys
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                680
                                    685 .
Lys Gln Glu Lys Leu Lys Ser His Leu His Pro Phe Lys Glu Ala
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Ala Gln Glu Val Asp Ser Lys Leu Gln Leu Phe Lys Glu Asn Asn
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                                                         720
Arg Arg Arg Lys Lys Glu Arg Lys Glu Lys Arg Arg Gln Arg Lys
                725
                                    730
                                                         735
Gly Glu Glu Cys Ser Leu Pro Gly Leu Thr Cys Phe Thr His Asp
                740
                                    745
Asn Asn His Trp Gln Thr Ala Pro Phe Trp Asn Leu Gly Ser Phe
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                                                         765
                                    760
Cys Ala Cys Thr Ser Ser Asn Asn Asn Thr Tyr Trp Cys Leu Arg
                                                         780
                770
                                    775
Thr Val Asn Glu Thr His Asn Phe Leu Phe Cys Glu Phe Ala Thr
                785
                                    790
                                                         795
Gly Phe Leu Glu Tyr Phe Asp Met Asn Thr Asp Pro Tyr Gln Leu
                800
                                    805
Thr Asn Thr Val His Thr Val Glu Arg Gly Ile Leu Asn Gln Leu
                815
                                    820
His Val Gln Leu Met Glu Leu Arg Ser Cys Gln Gly Tyr Lys Gln
                                    835
                                                         840
                830
Cys Asn Pro Arg Pro Lys Asn Leu Asp Val Gly Asn Lys Asp Gly
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Gly Ser Tyr Asp Leu His Arg Gly Gln Leu Trp Asp Gly Trp Glu
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                                      40
Phe Thr Ser Thr Ser Pro Ala Thr Gly Thr Gln Met Val Asn Ala
                 50
                                      55
Ala Ala Asn Gly Leu Gly Ala Glu Pro Met Glu Ser Phe Lys Gln
                                      70
                 65
Ala Tyr Arg His Cys Ile Lys Ile Pro Asp Phe Lys Ile Pro Ser
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Gln Gly Ser His Lys Thr Ile Ile Phe Ser
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Glu Arg Tyr Asp Leu Arg Asn Ile Val Gln Pro Lys Pro Leu Pro
                 80
                                     85
                                                          90
Ser Gln Phe Gly His Tyr Phe Glu Thr Thr Tyr Asp Thr Ser Tyr
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                                    100
                                                         105
Asn Asn Lys Met Pro Leu Ser Thr His Arg Phe Lys Arg Glu Pro
                110
                                    115
                                                         120
His Trp Phe Pro Gly His Gln Pro Glu Leu Asp Pro Pro Arg Tyr
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Lys Cys Thr Glu Lys Ser Thr Tyr Met Asn Ser Tyr Ser Lys Pro
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                                                          30
Arg Lys Gln Ser Arg Asn Val Leu Ser His Gln Asp Gly His Ile
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                                     40
Leu Gln Cys Ser Phe Arg Pro Asp Arg Arg Met Lys Arg Lys Ala
                 50
                                     55
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                                                          30
Met Ile Cys Asp Pro Tyr Ser Val Ala Pro Ala Gly Gly Pro Ala
                 35
                                      40
                                                          45
Gly Ala Lys Ala Pro Pro Pro Gly Pro Ser Thr Ala Ala Leu Glu
                 50
                                     55
                                                          60
Val Met Gln Asp Leu Ser Ala Asn Pro Pro Pro Pro Phe Ile Gln
                 65
                                     70
Gly Pro Lys Gly Asp Pro Gly Arg Pro Gly Lys Pro Gly Pro Arg
                 80
                                     85
Gly Pro Pro Gly Glu Pro Gly Pro Gly Pro Arg Gly Pro Pro
                 95
                                    100
                                                         105
Gly Glu Lys Gly Asp Ser Gly Arg Pro Gly Leu Pro Gly Leu Gln
                110
                                    115
                                                         120
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Leu Thr Thr Ser Ala Ala Gly Gly Val Gly Val Val Ser Gly Gly
                125
                                    130
Thr Gly Gly Gly Asp Thr Glu Gly Glu Val Thr Ser Ala Leu
                140
                                    145
Ser Ala Ala Phe Ser Gly Pro Lys Ile Ala Phe Tyr Val Gly Leu
                155
                                    160
Lys Ser Pro His Glu Gly Tyr Glu Val Leu Lys Phe Asp Asp Val
                170
                                    175
Val Thr Asn Leu Gly Asn His Tyr Asp Pro Thr Thr Gly Lys Phe
                185
                                    190
                                                         195
Ser Cys Gln Val Arg Gly Ile Tyr Phe Phe Thr Tyr His Ile Leu
                200
                                    205
                                                         210
Met Arg Gly Gly Asp Gly Thr Ser Met Trp Ala Asp Leu Cys Lys
                215
                                    220
                                                         225
Asn Gly Gln Val Arg Ala Ser Ala Ile Ala Gln Asp Ala Asp Gln
                230
                                    235
Asn Tyr Asp Tyr Ala Ser Asn Ser Val Val Leu His Leu Asp Ser
                245
                                    250
Gly Asp Glu Val Tyr Val Lys Leu Asp Gly Gly Lys Ala His Gly
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                                    265
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Gly Asn Asn Asn Lys Tyr Ser Thr Phe Ser Gly Phe Leu Leu Tyr
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                                    280
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Pro Asp
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Thr Leu Pro Gln Ser His Val Thr Ser Arg Arg Ala Gly Trp Lys
                 35
                                      40
Met Pro Leu Phe Leu Ile Leu Cys Leu Leu Gln Gly Ser Ser Phe
                                     55
                 50
Ala Leu Pro Gln Lys Arg Pro His Pro Arg Trp Leu Trp Glu Gly
                 65
                                     70
Ser Leu Pro Ser Arg Thr His Leu Arg Ala Met Gly Thr Leu Arg
                 80
                                     85
Pro Ser Ser Pro Leu Cys Trp Arg Glu Glu Ser Ser Phe Ala Ala
                 95
                                    1.00
Pro Asn Ser Leu Lys Gly Ser Arg Leu Val Ser Gly Glu Pro Gly
                110
                                    115
                                                         120
Gly Ala Val Thr Ile Gln Cys His Tyr Ala Pro Ser Ser Val Asn
                125
                                    130
Arg His Gln Arg Lys Tyr Trp Cys Arg Leu Gly Pro Pro Arg Trp
                140
                                     145
Ile Cys Gln Thr Ile Val Ser Thr Asn Gln Tyr Thr His His Arg
                155
                                    160
Tyr Arg Asp Arg Val Ala Leu Thr Asp Phe Pro Gln Arg Gly Leu
                170
                                    175
                                                         180
Phe Val Val Arg Leu Ser Gln Leu Ser Pro Asp Asp Ile Gly
                                                         Сув
                                                         195
                185
                                    190
Tyr Leu Cys Gly Ile Gly Ser Glu Asn Asn Met Leu Phe Leu Ser
                200
                                    205
                                                         210
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Met Asn Leu Thr Ile Ser Ala Gly Pro Ala Ser Thr Leu Pro Thr
                215
                                    220
Ala Thr Pro Ala Ala Gly Glu Leu Thr Met Arg Ser Tyr Gly Thr
                230
                                    235
                                                         240
Ala Ser Pro Val Ala Asn Arg Trp Thr Pro Gly Ser His Pro Asp
                245
                                    250
Leu Arg Thr Gly Asp Ser Met Gly His Met Leu Leu Pro His Pro
                260
                                    265
Gly Thr Ser Lys Thr Thr Ala Ser Ala Glu Gly Arg Arg Thr Pro
                275
                                    280
Gly Ala Thr Arg Pro Ala Ala Pro Gly Thr Gly Ser Trp Ala Glu
                290
                                    295
                                                         300
Gly Ser Val Lys Ala Pro Ala Pro Ile Pro Glu Ser Pro Pro Ser
                305
                                    310
Lys Ser Arg Ser Met Ser Asn Thr Thr Glu Gly Val Arg Glu Gly
                320
                                    325
                                                         330
Thr Arg Ser Ser Val Thr Asn Arg Ala Arg Ala Ser Lys Asp Arg
                335
                                    340
Arg Glu Met Thr Thr Lys Ala Asp Arg Pro Arg Glu Asp Ile
                350
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                                                         360
Glu Gly Val Arg Ile Ala Leu Asp Ala Ala Lys Lys Val Leu Gly
                365
                                    370
                                                         375
Thr Ile Gly Pro Pro Ala Leu Val Ser Glu Thr Leu Ala Trp Glu
                380
                                    385
                                                         390
Ile Leu Pro Gln Ala Thr Pro Val Ser Lys Gln Gln Ser Gln Gly
                395
                                     400
Ser Ile Gly Glu Thr Thr Pro Ala Ala Gly Met Trp Thr Leu Gly
                410
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Thr Pro Ala Ala Asp Val Trp Ile Leu Gly Thr Pro Ala Ala Asp
                425
                                    430
                                                         435
Val Trp Thr Ser Met Glu Ala Ala Ser Gly Glu Gly Ser Ala Ala
                440
                                    445
                                                         450
Gly Asp Leu Asp Ala Ala Thr Gly Asp Arg Gly Pro Gln Ala Thr
                455
                                    460
                                                         465
Leu Ser Gln Thr Pro Ala Val Gly Pro Trp Gly Pro Pro Gly Lys
                470
                                    475
Glu Ser Ser Val Lys Arg Thr Phe Pro Glu Asp Glu Ser Ser Ser
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                                    490
Arg Thr Leu Ala Pro Val Ser Thr Met Leu Ala Leu Phe Met Leu
                500
                                    505
                                                         510
Met Ala Leu Val Leu Leu Gln Arg Lys Leu Trp Arg Arg Arg Thr
                515
                                    520
                                                         525
Ser Gln Glu Ala Glu Arg Val Thr Leu Ile Gln Met Thr His Phe
                530
                                    535
                                                         540
Leu Glu Val Asn Pro Gln Ala Asp Gln Leu Pro His Val Glu Arg
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Lys Met Leu Gln Asp Asp Ser Leu Pro Ala Gly Ala Ser Leu Thr
                560
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Ala Pro Glu Arg Asn Pro Gly Pro
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Thr Glu Val Pro Lys Asp Val Thr Val Arg Glu Gly Asp Asp Ile
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Glu Met Pro Cys Ala Phe Arg Ala Ser Gly Ala Thr Ser Tyr Ser
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                                     55
Leu Glu Ile Gln Trp Trp Tyr Leu Lys Glu Pro Pro Arg Glu Leu
                 65
                                     70
Leu His Glu Leu Ala Leu Ser Val Pro Gly Ala Arg Ser Lys Val
                 80
                                     85
Thr Asn Lys Asp Ala Thr Lys Ile Ser Thr Val Arg Val Gln Gly
                 95
                                    100
                                                        105
Asn Asp Ile Ser His Arg Leu Arg Leu Ser Ala Val Arg Leu Gln
                110
                                    115
                                                        120
Asp Glu Gly Val Tyr Glu Cys Arg Val Ser Asp Tyr Ser Asp Asp
                125
                                    130
                                                         135
Asp Thr Gln Glu His Lys Ala Gln Ala Met Leu Arg Val Leu Ser
                140
                                    145
Arg Phe Ala Pro Pro Asn Met Gln Ala Ala Glu Ala Val Ser His
                155
                                    160
                                                        165
Ile Gln Ser Ser Gly Pro Arg Arg His Gly Pro Ala Ser Ala Ala
                170
                                    175
                                                        180
Asn Ala Asn Asn Ala Gly Ala Ala Ser Arg Thr Thr Ser Glu Pro
                185
                                    190
Gly Arg Gly Asp Lys Ser Pro Pro Pro Gly Ser Pro Pro Ala Ala
                200
                                    205
Ile Asp Pro Ala Val Pro Glu Ala Ala Ala Ala Ser Ala Ala His
                215
                                    220
Thr Pro Thr Thr Val Ala Ala Ala Ala Ala Ser Ser Ala
                230
                                    235
                                                         240
Ser Pro Pro Ser Gly Gln Ala Val Leu Leu Arg Gln Arg His Gly
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                                    250
                                                        255
Ser Gly Lys Gly Arg Ser Tyr Thr Thr Asp Pro Leu Leu Ser Leu
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Leu Leu Leu Ala Leu His Lys Phe Leu Arg Leu Leu Gly His
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Arg Asn Arg Leu Arg Ile Pro Tyr Ser Gln Asn Cys Gly Ile Phe
                 35
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Lys Pro Gln Arg Lys Pro Arg Asp Pro Arg Arg Leu Phe Cys Gly
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Cys Gly Lys Phe Lys Tyr Pro Pro Arg Leu His Ser
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<212> PRT

<213> Homo sapiens

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Ala Ala Asp Leu Gly Leu Ala Leu Leu Asp Val Ile Leu Gln Pro
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Arg Gly Lys Leu Ser Leu Tyr Val Pro Ser Thr Ala Trp Gly Gln
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Thr Arg Thr Leu Thr Val Ala Met Ala Glu Gly Leu
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Ser Gly Ala Val Ser Gln Ala Trp Ala Ser Phe Asn Ile Phe Tyr
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                                      40
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Leu Ala Leu His Gly Ala Ala Pro Ala Met Val Pro Gln Gly Phe
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                                                          60
Phe Ser Gln Val Ser Ser Leu Glu Arg Ser Pro Arg Phe Pro Val
                 65
                                      70
Lys Gln Pro Cys Ser Leu Cys Leu Ser Gln Pro His His Pro Val
                 80
                                     85
Ala Ser Phe Thr Ala Cys Leu Thr Ile Cys Asn His Leu Ser Val
                 95
                                     100
                                                         105
Cys Arg Leu Val Asp Leu Leu Pro Pro His Cys Gln Leu Leu Gly
                110
                                     115
                                                         120
Asn Arg Asp Trp Phe Val Tyr Cys Ala Ser Leu Val Pro Arg Thr
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Gly His Gly Ile Leu Leu Val His Asn Lys Tyr Gly Gly Asn
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Pro Arg Ala Ala Gly Lys Ala Leu Met Val Trp Val Val Leu Phe
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Pro Trp Ala Glu Leu Gly Trp Arg Thr Leu Ser Arg Val Ala Ala
                 35
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Ser Leu Trp Gly Pro Tyr Leu Gly Thr Tyr Thr Asp Gln Ala Val
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Cys Leu Cys Ser Leu Ser Asn His Asn Tyr Ser Gln Lys Ala Cys
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Gly Leu Glu Ser Thr Thr Val Lys Pro Gly Arg Met Cys Tyr Pro
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Val Pro Glu Arg Leu Leu Val Cys Val Leu
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Leu Phe Ser Leu Pro Phe Ser Leu Cys Pro Ser Ser Leu Ser Leu
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                                     25
Leu Phe Phe Leu Leu Ala Val Gly Phe Tyr Phe Phe Phe Glu Thr
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Ser Leu Ala Leu Ser Pro Arg Leu Glu Cys Ser Gly Ala Ile Ser
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Ala His Cys Lys Leu Cys Leu Pro Gly Ser Cys Tyr Ser Trp Ala
Ser Ala Cys
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Pro Leu Val Thr Val Cys Leu Arg Ala Leu Gly Leu Ala Gly Trp
                 35
                                     40
Glu Gln Thr Gln Leu Cys Gly Gly His Gln Val Val Pro Phe Ile
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Ser Ser Gly Leu Ser Leu Leu Glu Cys Gly Arg Cys Gln Lys Gln
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Cys Lys Gln Glu Ile Leu Arg Glu Met Gly Met Trp Glu Asp Thr
                 35
Gly Val Ala Arg Ser Ser Cys Thr Glu Val Asn Lys Asn Pro Ala
                 50
                                     55
Gly Ser Ser Trp Met Gly Ile Gln Gln Thr Arg Ala His Asn Ser
                 65
                                     70
Gly Arg Ala Thr Tyr Thr Gly Ala Cys Asp Trp Leu Gln Trp Ser
                 80
                                     85
                                                          90
Pro Leu Arg Ala Arg Asp Pro Ala Ala Ile Lys Gln Glu Lys Leu
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Gln Val Gly Ser Arg Phe
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Glu Thr Glu Ile Ala Lys Pro Val Leu Tyr Lys Glu Cys Ala Ser
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                                     25
Ala Ile Glu Asp Thr Ala Arg Ile Gly Cys Trp Ser Ser Ala Gly
                                     40
Pro Ala Val Ile Thr Arg Val Gln Gln Arg Glu Ser Pro Pro Leu
                 50
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Pro Ser Leu Thr Gln His Leu Thr Leu Ser His Ser
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Met Phe Cys Ala Phe Leu Phe Leu Pro Phe Ser Gln Asp Val Leu
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Cys Met Cys Phe Gly Lys Val Val Leu Val Met Phe Ile Leu Leu
                 20
                                     25
Cys Ile Cys Ser Val Leu Glu Leu Phe Phe Ser Ser Gly Arg Cys
                 35
                                     40
                                                          45
Phe Glu Ser Thr Leu Phe Ile Val Ala His Val Ser Asn Leu Ile
                 50
                                     55
Ser Lys Ile Leu Gln Val Tyr Ser Leu Arg Arg Ile Leu Phe Ile
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                                     70
Tyr Cys Thr Asp Met Leu Cys Thr Arg His Cys Ala Met Ala Asn
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Ser Tyr Phe Pro Gly Lys Pro Gly Glu Leu Thr Leu Phe Ser Val
                 35
                                     40
Leu Pro Glu Leu Ser Gln Ser Leu Gly Leu Arg Glu Gln Glu Leu
                 50
                                     55
Gln Val Val Arg Ala Ser Gly Lys Glu Ser Ser Gly Leu Val Leu
                 65
                                     70
Leu Ser Ser Cys Pro Gln Thr Ala Ser Arg Leu Gln Lys Tyr Phe
                 80
                                     85
Thr His Ala Arg Arg Ala Gln Arg Pro Thr Ala Thr Tyr Cys Ala
                 95
                                    100
Val Thr Asp Gly Ile Pro Ala Ala Ser Glu Gly Lys Ile Gln Ala
                110
                                    115
                                                        120
Ala Leu Lys Leu Glu His Ile Asp Gly Val Asn Leu Thr Val Pro
                125
                                    130
                                                        135
Val Lys Ala Pro Ser Arg Lys Asp Ile Leu Glu Gly Val Lys Lys
                140
                                    145
Thr Leu Ser His Phe Arg Val Val Ala Thr Gly Ser Gly Cys Ala
                155
                                    160
                                                        165
Leu Val Gln Leu Gln Pro Leu Thr Val Phe Ser Ser Gln Leu Gln
               170
                                    175
                                                        180
Val His Met Val Leu Gln Leu Cys Pro Val Leu Gly Asp His Met
                185
                                    190
Tyr Ser Ala Arg Val Gly Thr Val Leu Gly Gln Arg Phe Leu Leu
                200
                                    205
Pro Ala Glu Asn Asn Lys Pro Gln Arg Gln Val Leu Asp Glu Ala
                215
                                    220
Leu Leu Arg Arg Leu His Leu Thr Pro Ser Gln Ala Ala Gln Leu
                230
                                    235
Pro Leu His Leu His Arg Leu Leu Pro Gly Thr Arg
               245
                                   250
                                                        255
Ala Arg Asp Thr Pro Val Glu Leu Leu Ala Pro Leu Pro Pro Tyr
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                                    265
Phe Ser Arg Thr Leu Gln Cys Leu Gly Leu Arg Leu Gln
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                                    280
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Gln Ala Ala Glu Phe Asp Gly Ser Arg Trp Pro Arg Gln Ile
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Val Ser Ser Ile Gly Leu Cys Arg Tyr Gly Gly Arg Ile Asp Cys Cys Trp Gly Trp Ala Arg Gln Ser Trp Gly Gln Cys Gln Pro Val Cys Gln Pro Arg Cys Lys His Gly Glu Cys Ile Gly Pro Asn Lys Cys Lys Cys His Pro Gly Tyr Ala Gly Lys Thr Cys Asn Gln Asp Leu Asn Glu Cys Gly Leu Lys Pro Arg Pro Cys Lys His Arg Cys Met Asn Thr Tyr Gly Ser Tyr Lys Cys Tyr Cys Leu Asn Gly Tyr Met Leu Met Pro Asp Gly Ser Cys Ser Ser Ala Leu Thr Cys Ser Met Ala Asn Cys Gln Tyr Gly Cys Asp Val Val Lys Gly Gln Ile Arg Cys Gln Cys Pro Ser Pro Gly Leu Gln Leu Ala Pro Asp Gly Arg Thr Cys Val Asp Val Asp Glu Cys Ala Thr Gly Arg Ala Ser Cys Pro Arg Phe Arg Gln Cys Val Asn Thr Phe Gly Ser Tyr Ile Cys Lys Cys His Lys Gly Phe Asp Leu Met Tyr Ile Gly Gly Lys Tyr Gln Cys His Asp Ile Asp Glu Cys Ser Leu Gly Gln Tyr Gln Cys Ser Ser Phe Ala Arg Cys Tyr Asn Val Arg Gly Ser Tyr Lys Cys Lys Cys Lys Glu Gly Tyr Gln Gly Asp Gly Leu Thr Cys Val Tyr Ile Pro Lys Val Met Ile Glu Pro Ser Gly Pro Ile His Val Pro Lys Gly Asn Gly Thr Ile Leu Lys Gly Asp Thr Gly Asn Asn Asn Trp Ile Pro Asp Val Gly Ser Thr Trp Trp Pro Pro Lys Thr Pro Tyr Ile Pro Pro Ile Ile Thr Asn Arg Pro Thr Ser Lys Pro Thr Thr Arg Pro Thr Pro Lys Pro Thr Pro Ile Pro Thr Pro Pro Pro Pro Pro Pro Leu Pro Thr Glu Leu Arg Thr Pro Leu Pro Pro Thr Thr Pro Glu Arg Pro Thr Thr Gly Leu Thr Thr Ile Ala Pro Ala Ala Ser Thr Pro Pro Gly Gly Ile Thr Val Asp Asn Arg Val Gln Thr Asp Pro Gln Lys Pro Arg Gly Asp Val Phe Ile Pro Arg Gln Pro Ser Asn Asp Leu Phe Glu Ile Phe Glu Ile Glu Arg Gly Val Ser Ala Asp Asp Glu Ala Lys Asp Asp Pro Gly Val Leu Val His Ser Cys Asn Phe Asp His Gly Leu Cys Gly Trp Ile Arg Glu Lys Asp Asn Asp Leu His Trp Glu Pro Ile Arg Asp Pro Ala Gly Gly Gln Tyr Leu Thr Val Ser Ala Ala Lys Ala Pro Gly Gly Lys Ala Ala Arg Leu Val Leu Pro Leu Gly Arg Leu Met His Ser Gly Asp Leu Cys Leu Ser Phe Arg His Lys Val Thr Gly Leu His Ser Gly Thr Leu Gln Val Phe Val Arg Lys His Gly Ala His Gly Ala

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500
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Ala Leu Trp Gly Arg Asn Gly Gly His Gly Trp Arg Gln Thr Gln
                515.
                                    520
Ile Thr Leu Arg Gly Ala Asp Ile Lys Ser Val Val Phe Lys Gly
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                                                         540
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Glu Lys Arg Arg Gly His Thr Gly Glu Ile Gly Leu Asp Asp Val
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                                    550
Ser Leu Lys Lys Gly His Cys Ser Glu Glu Arg
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Pro Cys Leu Leu Leu Leu Leu Leu Leu Leu Arg Leu Glu Pro
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                                     25
                                                          30
Val Thr Ala Ala Ala Gly Pro Arg Ala Pro Cys Ala Ala Ala Cys
                 35
                                      40
                                                          45
Thr Cys Ala Gly Asp Ser Leu Asp Cys Gly Gly Arg Gly Leu Ala
                 50
                                     55
Ala Leu Pro Gly Asp Leu Pro Ser Trp Thr Arg Ser Leu Asn Leu
                 65
                                     70
Ser Tyr Asn Lys Leu Ser Glu Ile Asp Pro Ala Gly Phe Glu Asp
                 80
                                     85
Leu Pro Asn Leu Gln Glu Val Tyr Leu Asn Asn Asn Glu Leu Thr
                 95
                                    100
Ala Val Pro Ser Leu Gly Ala Ala Ser Ser His Val Val Ser Leu
                110
                                     115
                                                         120
Phe Leu Gln His Asn Lys Ile Arg Ser Val Glu Gly Ser Gln Leu
                125
                                    130
Lys Ala Tyr Leu Ser Leu Glu Val Leu Asp Leu Ser Leu Asn Asn
                140
                                    145
Ile Thr Glu Val Arg Asn Thr Cys Phe Pro His Gly Pro Pro Ile
                155
                                    160
Lys Glu Leu Asn Leu Ala Gly Asn Arg Ile Gly Thr Leu Glu Leu
                170
                                    175
Gly Ala Phe Asp Gly Leu Ser Arg Ser Leu Leu Thr Leu Arg Leu
                185
                                    190
Ser Lys Asn Arg Ile Thr Gln Leu Pro Val Arg Ala Phe Lys Leu
                200
                                    205
Pro Arg Leu Thr Gln Leu Asp Leu Asn Arg Asn Arg Ile Arg Leu
                215
                                    220
                                                         225
Ile Glu Gly Leu Thr Phe Gln Gly Leu Asn Ser Leu Glu Val Leu
                230
                                    235
Lys Leu Gln Arg Asn Asn Ile Ser Lys Leu Thr Asp Gly Ala Phe
                245
                                     250
Trp Gly Leu Ser Lys Met His Val Leu His Leu Glu Tyr Asn Ser
                260
                                    265
Leu Val Glu Val Asn Ser Gly Ser Leu Tyr Gly Leu Thr Ala Leu
                                    280
                275
His Gln Leu His Leu Ser Asn Asn Ser Ile Ala Arg Ile His Arg
                                    295
                290
                                                         300
Lys Gly Trp Ser Phe Cys Gln Lys Leu His Glu Leu Val Leu Ser
                305
                                    310
Phe Asn Asn Leu Thr Arg Leu Asp Glu Glu Ser Leu Ala Glu Leu
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G	a	T	0	320	T	3	T	a	325	7	G	T1.	C	330
ser	ser	ьец	ser	Val 335	ren	Arg	ьeu	ser	340	Asn	ser	TTE	ser	345
Ile	Ala	Glu	Gly	Ala 350	Phe	Lys	Gly	Leu		Ser	Leu	Arg	Va1	
Asp	Leu	Asp	His	Asn 365	Glu	Ile	Ser	Gly		Ile	Glu	Asp	Thr	
Gly	Ala	Phe	Ser	Gly 380	Leu	Asp	Ser	Leu	_	Lys	Leu	Thr	Leu	
Gly	Asn	Lys	Ile	Lys 395	Ser	Val	Ala	Lys	-	Ala	Phe	Ser	Gly	Leu 405
Glu	Gly	Leu	Glu	His 410	Leu	Asn	Leu	Gly	Gly 415	Asn	Ala	Ile	Arg	Ser 420
Val	Gln	Phe	qaA	Ala 425	Phe	Val	ГЛЗ	Met	Lys 430	Asn	Leu	Lys	Glu	Leu 435
His	Ile	Ser	Ser	Asp 440	Ser	Phe	Leu	Сув	Asp 445	Сув	Gln	Leu	Lys	Trp 450
				Leu 455					460					465
				His 470					475					480
				Glu 485					490					495
				Gln 500					505				_	510
_				Thr 515	-				520					525
				Trp 530					535					540
-				Phe 545					550		_	_		555
				Thr 560					565					570
				Tyr 575 Lys					580					585
				590 Pro					595					600
				605 Cys					610					615
				620 Asp					625	•				630
				635 Val					640					645
	_			650 Asp			_	_	655					660
				665 Ser					670					675
			_	680 Leu					685					690
				695 Val					700					705
			_	710 Thr				_	715					720
		_		725 His	_		_	_	730	_				735
				740 Ala					745					750
				755 Gly					760					765
				770 Gly					775					780
				785					790					795

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Phe Thr Ile Ala Val Val Ser Ser Ile Val Leu Thr Ser Leu Val
                800
                                    805
                                                         810
Trp Val Cys Ile Ile Tyr Gln Thr Arg Lys Lys Ser Glu Glu Tyr
                815
                                    820
Ser Val Thr Asn Thr Asp Glu Thr Val Val Pro Pro Asp Val Pro
                830
                                    835
                                                         840
Ser Tyr Leu Ser Ser Gln Gly Thr Leu Ser Asp Arg Gln Glu Thr
                845
                                    850
                                                         855
Val Val Arg Thr Glu Gly Gly Pro Gln Ala Asn Gly His Ile Glu
                860
                                    865
                                                         870
Ser Asn Gly Val Cys Pro Arg Asp Ala Ser His Phe Pro Glu Pro
                875
                                    880
                                                         885
Asp Thr His Ser Val Ala Cys Arg Gln Pro Lys Leu Cys Ala Gly
                890
                                    895
                                                         900
Ser Ala Tyr His Lys Glu Pro Trp Lys Ala Met Glu Lys Ala Glu
                905
                                    910
Gly Thr Pro Gly Pro His Lys Met Glu His Gly Gly Arg Val Val
                920
                                    925
                                                         930
Cys Ser Asp Cys Asn Thr Glu Val Asp Cys Tyr Ser Arg Gly Gln
                935
                                    940
                                                         945
Ala Phe His Pro Gln Pro Val Ser Arg Asp Ser Ala Gln Pro Ser
                                    955
                950
                                                         960
Ala Pro Asn Gly Pro Glu Pro Gly Gly Ser Asp Gln Glu His Ser
                965
                                    970
                                                         975
Pro His His Gln Cys Ser Arg Thr Ala Ala Gly Ser Cys Pro Glu
                980
                                    985
                                                         990
Cys Gln Gly Ser Leu Tyr Pro Ser Asn His Asp Arg Met Leu Thr
                995
                                   1000
                                                        1005
Ala Val Lys Lys Pro Met Ala Ser Leu Asp Gly Lys Gly Asp
               1010
                                   1015
                                                        1020
Ser Ser Trp Thr Leu Ala Arg Leu Tyr His Pro Asp Ser Thr Glu
                                   1030
               1025
                                                        1035
Leu Gln Pro Ala Ser Ser Leu Thr Ser Gly Ser Pro Glu Arg Ala
               1040
                                   1045
                                                        1050
Glu Ala Gln Tyr Leu Leu Val Ser Asn Gly His Leu Pro Lys Ala
               1055
                                   1060
Cys Asp Ala Ser Pro Glu Ser Thr Pro Leu Thr Gly Gln Leu Pro
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Gly Lys Gln Arg Val Pro Leu Leu Leu Ala Pro Lys Ser
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Leu Leu Leu Gly Ser Arg Pro Ala Arg Gly Ala Gly Pro Glu
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Pro Pro Val Leu Pro Ile Arg Ser Glu Lys Glu Pro Leu Pro Val
                 35
                                      40
Arg Gly Ala Ala Gly Cys Thr Phe Gly Gly Lys Val Tyr Ala Leu
                 50
                                      55
Asp Glu Thr Trp His Pro Asp Leu Gly Glu Pro Phe Gly Val Met
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Arg Cys Val Leu Cys Ala Cys Glu Ala Pro Gln Trp Gly Arg Arg

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Thr Arg Gly Pro Gly Arg Val Ser Cys Lys Asn Ile Lys Pro Glu
                  95
                                      100
 Cys Pro Thr Pro Ala Cys Gly Gln Pro Arg Gln Leu Pro Gly His
                 110
                                      115
 Cys Cys Gln Thr Cys Pro Gln Glu Arg Ser Ser Ser Glu Arg Gln
                                      130
 Pro Ser Gly Leu Ser Phe Glu Tyr Pro Arg Asp Pro Glu His Arg
                 140
                                     145
 Ser Tyr Ser Asp Arg Gly Glu Pro Gly Ala Glu Glu Arg Ala Arg
                 155
                                     160
 Gly Asp Gly His Thr Asp Phe Val Ala Leu Leu Thr Gly Pro Arg
                 170
                                      175
 Ser Gln Ala Val Ala Arg Ala Arg Val Ser Leu Leu Arg Ser Ser
                 185
                                      190
 Leu Arg Phe Ser Ile Ser Tyr Arg Arg Leu Asp Arg Pro Thr Arg
                 200
                                      205
 Ile Arg Phe Ser Asp Ser Asn Gly Ser Val Leu Phe Glu His Pro
                 215
                                      220
 Ala Ala Pro Thr Gln Asp Gly Leu Val Cys Gly Val Trp Arg Ala
                 230
                                      235
 Val Pro Arg Leu Ser Leu Arg Leu Leu Arg Ala Glu Gln Leu His
                 245
                                      250
                                                          255
 Val Ala Leu Val Thr Leu Thr His Pro Ser Gly Glu Val Trp Gly
                 260
                                      265
 Pro Leu Ile Arg His Arg Ala Leu Ala Ala Glu Thr Phe Ser Ala
                 275
                                      280
 Ile Leu Thr Leu Glu Gly Pro Pro Gln Gln Gly Val Gly Gly Ile
                 290
                                      295
 Thr Leu Leu Thr Leu Ser Asp Thr Glu Asp Ser Leu His Phe Leu
                 305
                                      310
 Leu Leu Phe Arg Gly Leu Leu Glu Pro Arg Ser Gly Gly Leu Thr
                 320
                                      325
 Gln Val Pro Leu Arg Leu Gln Ile Leu His Gln Gly Gln Leu Leu
                 335
                                      340
 Arg Glu Leu Gln Ala Asn Val Ser Ala Gln Glu Pro Gly Phe Ala
                                      355
 Glu Val Leu Pro Asn Leu Thr Val Gln Glu Met Asp Trp Leu Val
                 365
                                      370
 Leu Gly Glu Leu Gln Met Ala Leu Glu Trp Ala Gly Arg Pro Gly
                 380
                                     385
' Leu Arg Ile Ser Gly His Ile Ala Ala Arg Lys Ser Cys Asp Val
                 395
                                      400
                                                          405
 Leu Gln Ser Val Leu Cys Gly Ala Asp Ala Leu Ile Pro Val Gln
                 410
                                      415
 Thr Gly Ala Ala Gly Ser Ala Ser Leu Thr Leu Leu Gly Asn Gly
                 425
                                      430
 Ser Leu Ile Tyr Gln Ala Val Gly Ile Cys Pro Gly Leu Gly Ala
                 440
                                      445
 Arg Gly Ala His Met Leu Leu Gln Asn Glu Leu Phe Leu Asn Val
                 455
                                      460
 Gly Thr Lys Asp Phe Pro Asp Gly Glu Leu Arg Gly His Val Ala
                 470
                                      475
                                                          480
 Ala Leu Pro Tyr Cys Gly His Ser Ala Arg His Asp Thr Leu Pro
                 485
                                      490
 Val Pro Leu Ala Gly Ala Leu Val Leu Pro Pro Val Lys Ser Gln
                 500
                                      505
 Ala Ala Gly His Ala Trp Leu Ser Leu Asp Thr His Cys His Leu
                 515
                                      520
 His Tyr Glu Val Leu Leu Ala Gly Leu Gly Gly Ser Glu Gln Gly
                 530
                                      535
 Thr Val Thr Ala His Leu Leu Gly Pro Pro Gly Thr Pro Gly Pro
                 545
                                      550
 Arg Arg Leu Leu Lys Gly Phe Tyr Gly Ser Glu Ala Gln Gly Val
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Val Lys Asp Leu Glu Pro Glu Leu Leu Arg His Leu Ala Lys Gly

Met Ala Ser Leu Leu Ile Thr Thr Lys Gly Ser Pro Arg Gly Glu

Leu Arg Gly Gln Val His Ile Ala Asn Gln Cys Glu Val Gly Gly

Leu Arg Leu Glu Ala Ala Gly Ala Glu Gly Val Arg Ala Leu Gly

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620
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Ala Pro Asp Pro Ala Ser Ala Ala Pro Pro Val Val Pro Gly Leu
                635
                                     640
Pro Ala Leu Ala Pro Ala Lys Pro Gly Gly Pro Gly Arg Pro Arg
                                     655
                650
                                                         660
Asp Pro Asn Thr Cys Phe Phe Glu Gly Gln Gln Arg Pro His Gly
                665
                                     670
                                                         675
Ala Arg Trp Ala Pro Asn Tyr Asp Pro Leu Cys Ser Leu Cys Thr
                                     685
                680
Cys Gln Arg Arg Thr Val Ile Cys Asp Pro Val Val Cys Pro Pro
                695
                                     700
Pro Ser Cys Pro His Pro Val Gln Ala Pro Asp Gln Cys Cys Pro
                710
                                     715
                                                         720
Val Cys Pro Glu Lys Gln Asp Val Arg Asp Leu Pro Gly Leu Pro
                725
                                     730
Arg Ser Arg Asp Pro Gly Glu Gly Cys Tyr Phe Asp Gly Asp Arg
                740
                                     745
                                                         750
Ser Trp Arg Ala Ala Gly Thr Arg Trp His Pro Val Val Pro Pro
                755
                                     760
Phe Gly Leu Ile Lys Cys Ala Val Cys Thr Cys Lys Gly Gly Thr
                770
                                    775
Gly Glu Val His Cys Glu Lys Val Gln Cys Pro Arg Leu Ala Cys
                785
                                     790
                                                         795
Ala Gln Pro Val Arg Val Asn Pro Thr Asp Cys Cys Lys Gln Cys
                800
                                     805
Pro Val Gly Ser Gly Ala His Pro Gln Leu Gly Asp Pro Met Gln
                815
                                     820
Ala Asp Gly Pro Arg Gly Cys Arg Phe Ala Gly Gln Trp Phe Pro
                830
                                     835
Glu Ser Gln Ser Trp His Pro Ser Val Pro Pro Phe Gly Glu Met
                                     850
                845
Ser Cys Ile Thr Cys Arg Cys Gly Ala Gly Val Pro His Cys Glu
                860
                                     865
                                                         870
Arg Asp Asp Cys Ser Leu Pro Leu Ser Cys Gly Ser Gly Lys Glu
                875
                                     880
Ser Arg Cys Cys Ser Arg Cys Thr Ala His Arg Arg Pro Ala Pro
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Glu Thr Arg Thr Asp Pro Glu Leu Glu Lys Glu Ala Glu Gly Ser
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Val Leu Asp Ser Gly Met Cys Val Arg Ala Gly Glu Cys Leu Asp
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Gly Asp Val Val Ser Leu Leu His Phe Trp His Ser Val Thr Thr
                                     40
Gln Glu Asn Gln Ile Glu Asn Leu Glu Ser Val Leu Gln Trp Ile
                 50
                                     55
                                                          60
Glu Thr Gly Leu Gln Ser Leu Arg Lys Lys Ser Lys Gln Asn Thr
                 65
                                     70
Gln Glu Phe Arg Glu Asn Ile Phe Leu Pro Lys Asn Asn Phe Ser
                 80
                                     85
Phe Met Leu Phe Leu Ile Trp Val Asn Thr Pro Met Glu Lys Ile
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                                    100
Asp Arg Leu Val Lys Ser Ser Ile
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Met Gly Lys Gly Arg Trp Ala Thr Val Gly Val Ser Pro Cys Leu
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Pro Pro Leu Trp Ala Ala Ala Gly Ala His Ala Ser Lys Ser Ser
                 20
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Leu Arg Glu Arg Glu Leu Arg Cys Leu Tyr Pro Ser Ser Val Arg
                 35
                                     40
His Trp Leu Asn Val His Thr Pro Gly Ser Pro Pro Leu Ile Leu
                 50
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Met Met Ser His Gly Pro His Phe Thr Ser Glu Leu Trp Val His
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Gly Glu His Gln Ser His Pro Gly Ser Val Pro Gln Leu Ser Leu
Thr
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Met Arg Met Phe Pro Leu Pro Leu Pro Val Cys Leu Pro Leu Gly
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Val His Leu Gln Ser Thr Ser Pro Pro Phe Pro Ala Ser His Thr
                                     25
Gln Val Ser Leu Ser Asp Ser His Thr Cys Leu Thr Ala Ser Pro
                 35
                                     40
Ala Lys Val Leu Phe Lys Cys Leu Phe Ser Val Cys Leu Cys His
                 50
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Ser Gln Cys Asp His Ser Cys Ser Ala Val Ser Gln Gln Glu Asp
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                                     70
Arg Cys Arg Ser Ser Ser Cys Ser
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Ile Leu Leu Pro Thr Val Ala Asn Ile Ala Leu Ser Ser Ser Arg
                 20
                                     25
Thr Gly Arg Ser Lys Glu His Thr Gln Asp Asp Ala Thr Ala Tyr
                 35
                                     40
Met Leu Ser Arg His Leu His Ala Leu Ser Ala Pro Thr Cys Ser
                                     55
Leu Gly Ser Leu His Ala Leu Ser Ala Ala Tyr Thr Leu Ser Trp
                 65
                                     70
His Val Gln Gln Val Leu Gln Pro Cys Pro Gly Gly Leu Gly Leu
                 80
                                     85
Arg Gly Leu Ser Leu Ser Trp Val Leu Asp Leu Pro Pro His Phe
                 95
                                    100
                                                         105
His His Cys Asn Phe Cys Phe Thr Cys Trp Lys Gly Ala Ser Tyr
                110
                                    115
Asn Met Pro Leu Lys Glu Lys Asp
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Met Ala Leu Leu Trp Trp Ile Ser Thr Val Ala Ile Leu Leu Phe
 1
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Thr Ser Thr Ile Leu Gly Thr Tyr Val Glu Ala Gly Ala Ala Lys
                 20
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Ser Asn Glu Glu Glu Ile Val Asn Lys Ser Glu Phe Gly Arg Phe
Pro Arg Gly Ser Arg Lys Asp Ala Ser Gly Cys His Lys Pro Gly
                 50
                                     55
                                                          60
Tyr Pro Val Pro Pro His Ser Arg Cys Pro Pro Pro Pro His Val
                 65
Gln Arg Pro Arg Pro Ile Leu His Ala
                 80
<210> 47
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Met Gly Trp Pro Pro Pro Pro Gly Ser Ser Phe Cys Leu Cys Phe
Ile His Gly Ala Phe Ser Ser Phe Ser Pro His Pro Pro Ser His
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Glu Cys Ser Ser Arg Cys Cys Ser Leu Cys Leu Ala Arg Phe Leu
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                                     40
Ala Ser Pro Leu Pro Trp Ser Asn Ser Glu Ser Ser Ser Thr Leu
                 50
                                     55
                                                          60
Tyr Leu Lys Ser Arg Leu Ala Gly Ser Leu Ser Gly Ser Ala His
                 65
                                     70
Cys Ser Pro Thr Ser Leu Pro Phe Ser Leu Gly Thr Leu Ile Thr
                 80
                                     85
Pro Glu Thr Val Asp Ser Ser Pro Lys Tyr Ser Phe Trp Leu Ile
Val Gly Ala Gln
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  1
His Val Leu Ser Leu Ser Cys Ala Gln Cys Asn Cys Val His Val
Phe Leu Ile Pro Pro Pro Ala Leu Pro Ala Arg Phe Thr Glu Gly
                 35
                                     40
Leu Arg Asn Glu Glu Ala Met Glu Gly Ala Thr Ala Thr Leu Gln
                 50
                                     55
Cys Glu Leu Ser Lys Ala Ala Pro Val Glu Trp Arg Lys Gly Leu
                 65
                                     70
Glu Ala Leu Arg Asp Gly Asp Lys Tyr Ser Leu Arg Gln Asp Gly
Ala Val Cys Glu Leu Gln Ile His Gly Leu Ala Met Ala Asp Asn
                                    100
Gly Val Tyr Ser Cys Val Cys Gly Gln Glu Arg Thr Ser Ala Thr
                110
                                    115
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Leu Thr Val Arg Gly Lys Asp Pro Met Trp Pro Cys Gly Leu Val
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Arg Tyr Leu Trp Val Leu Val Ser Leu Ser Ala Thr Glu Ser Val
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Gln Asp Val Leu Leu Glu Gly His Pro Ser Trp Lys Tyr Leu Gln
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Glu Val Glu Thr Leu Leu Leu Asn Val Gln Gln Gly Leu Thr Asp
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Val Glu Val Ser Pro Lys Val Glu Ser Val Leu Ser Leu Leu Asn
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Ala Pro Gly Pro Asn Leu Lys Leu Val Arg Pro Lys Ala Leu Leu
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Asp Asn Cys Phe Arg Val Met Glu Leu Leu Tyr Cys Ser Cys
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Lys Gln Ser Ser Val Leu Asn Trp Gln Asp Cys Glu Val Pro Ser
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Pro Gln Ser Cys Ser Pro Glu Pro Ser Leu Gln Tyr Ala Ala Thr
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Glu Arg Asp Lys Gly Thr Gln Tyr Glu Leu Phe Phe Lys Lys Ala
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Lys Lys Ile His Leu Thr Val Val Tyr Phe Gly Lys Glu Gly Leu
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